

# QuES&T

Quality Environmental Solutions & Technologies, Inc.

September 12<sup>th</sup>, 2018

North Salem Central School District  
230 June Road  
North Salem, NY 10560

Attn: Gary Green

Re: Project No. Q18-1941 Pequenakonck Elementary School, Post-Remediation Assessment and Clearance Summary

Dear Mr. Green:

The following is a summary of the inspection(s) performed from August 20<sup>th</sup> – September 4<sup>th</sup>, 2018 within entirety of Pequenakonck Elementary School located at 230 June Road, North Salem, NY. QuES&T was asked to perform an initial and post-remediation assessment and clearance of the building in accordance with Article 32 of the New York State Labor Law. Article 32 of the New York State Labor Law states that the “post remediation assessment shall determine whether: (a) the work area is free from all visible mold; and (b) all work has been completed in compliance with the remediation plan and remediation work plan and meets clearance criteria specified in the plan.” In addition, analytical-based clearance criteria was established by the Client, North Salem Central School District & QuES&T. The clearance criteria was defined by the Client as such; Levels of total fungal spore counts within the area of concern (AOC) shall be less than total fungal spore counts of exterior and/or indoor control samples. Analytical results for this investigation are enclosed.

## **1.0 Visual Observations**

During the dates of August 22, 2018 through September 4<sup>th</sup>, 2018 Mr. Louis N Johnson III, Mold Assessor Cert. # MA00532 & Mr. Tanay Ranadive of QuES&T Mold Assessor Cert. # MA00534 arrived on site to conduct an initial visual inspection of the building which were observed to have mold growth on multiple types of surfaces (porous & non-porous). The impacted areas were remediated as per the remediation plan required by Article 32 of the New York State Labor Law and with agreements made between North Salem Central School District and a licensed Mold Remediation Contractor. Upon inspection, QuES&T made the following observations:

1. Preliminary inspection of the building identified visual suspected fungal growth occurring on various surfaces and materials within the building. The growth appeared to be surficial in nature and likely the result of high humidity. See photos: Appendix A
2. Further inspection of the building identified visual suspected fungal growth behind cabinets, various room contents, HVAC vents, on and behind cove base moldings along sheetrock walls. Fungal growth was also observed above ceilings on various pipe insulations.

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## **2.0 Remediation**

In addition to performing the visual inspections of the building, Mr. Louis N. Johnson III and Mr. Tanay Ranadive were onsite throughout the entire remediation process to continually evaluate the growth and remediation of each of the AOC's. All Pro Restoration & Cleaning, a New York State Licensed Mold remediation firm was mobilized to remediate the conditions present. The remediation began with each of the wings of the school being cordoned off from each other using 6-mil fire retardant polyethylene sheeting. This created a series of smaller work areas, each of which could be remediated and cleared and then returned to the district. A remediation plan was developed which provided an outline of how the individual AOC's would be remediated. General procedures used for remediation were to initially determine if an item was to be cleaned or to be disposed of. If the decision to clean was made, then the item was HEPA vacuumed, wet wiped with a mild disinfecting solution and allowed to dry. Upon completion of a successful clearance inspection the area was lightly misted with SteraMist followed by a sufficient drying period. Final clearance samples of each of the AOC's were collected and forwarded to QLab Microbiology of Metuchen NJ for analysis for total fungal structures. Additional inspections identified the following additional conditions:

1. On August 24<sup>th</sup>, mold growth was observed on and behind cove base moldings along sheetrock walls. Therefore, the licensed Mold Contractor removed 6in. of sheetrock from the ground up in various rooms. A map showing the various rooms where sheetrock was removed, is listed in Appendix C (See Photos).
2. On August 26<sup>th</sup>, it was determined that all carpets in the rooms would be covered in plastic sheeting and removed to another location. These carpets will be assessed at a later time on possible mold growth (See Photos).
3. Due to the ongoing roof work, water leaks did occur in the South Wing. Head Custodian Chris was notified of these issues. It was also observed that the humidity throughout the building increased when the HVAC system was activated. Therefore, the system was turned off; more negative air scrubber machines & dehumidifiers were placed throughout the building.
4. On September 2<sup>nd</sup>, Tanay Ranadive performed visual inspections on all carpets as well as various books from classrooms and the library. Approximately 10 carpets were taken by licensed Mold Contractor to be steam cleaned. 7 carpets were determined to be highly impacted by fungal growth and were therefore disposed of. Several library books and classroom books were determined to have been impacted by moisture and microbial growth and were disposed of.

A list of all materials & contents cleaned and disposed of are listed in the: Mold Assessment Documentation Sheets (See Appendix. D).

## **3.0 Sampling Data**

### **3.1 Sampling Protocol**

Air samples were collected between August 26<sup>th</sup>, 2018 and September 4<sup>th</sup>, 2018 for determination of Total Fungal Spore Levels throughout the building. Each wing of the school was sampled separately, one (1) sample was placed in various locations in the different wings. Two (2) samples were placed outside; pre-sample & post-sampling. Two (2) additional blank samples were submitted for QA/QC purposes.

For both sampling events, samples were collected using Air-O-Cell Spore Trap cassettes and a vacuum air pump calibrated to a uniform flow of fifteen (15) liters per minute for five (5) minutes for a total sample volume of 75 Liters. Exterior sampling was conducted to a uniform flow of fifteen (15) liters per minute for ten (10) minutes for a total sample volume of 150 Liters. Samples were sent to QLab Environmental Microbiology of Metuchen, NJ for presumptive identification and enumeration of fungi. The laboratory

results for the air samples are contained in Appendix B of this report. Samples were collected at the following locations:

**Table 1.0- Clearance Sample Locations**

<b>August 26<sup>th</sup>, 2018:</b> South Wing, Multipurpose Room & Cafeteria
<b>August 27<sup>th</sup>, 2018:</b> West Wing + Adjacent Rooms. Resampled South Wing except for Kindergarten Section
<b>August 28<sup>th</sup>, 2018:</b> North Wing. Resampled Cafeteria, West Wing and Adjacent Rooms
<b>August 29<sup>th</sup>, 2018:</b> East Wing. Resampled Main Office, Rm. N21
<b>August 30<sup>th</sup>, 2018:</b> Resampled East Wing
<b>September 1<sup>st</sup>, 2018:</b> Library Wing, Gym Wing & Kitchen Pantry
<b>September 2<sup>nd</sup>, 2018:</b> Gym Storage Room. Resampled Kitchen Pantry, Women's & Men's Restrooms by the Gym
<b>September 4<sup>th</sup>, 2018:</b> Resampled Mechanical Room by Library

### **3.2 Data Interpretation**

Mold is an omnipresent organism in the environment and will be detected in almost all air samples collected. Several factors are considered when evaluating indoor total fungal spore levels. Three major factors considered involve 1) comparison of indoor (AOC) to outdoor ambient concentrations 2) commonality of species between indoor and outdoor samples and 3) the presence of indicator species. Currently, no regulatory or health based standard exists for indoor levels of microbiological contaminants. Consistent with the clearance criteria established by the Client, analytical results have only been interpreted in regards to factor 1 listed above; comparison of AOC to control samples concentrations.

A health based numerical standard for acceptable exposure to microbial contaminants is not feasible for a variety of reasons. Microbial contaminants in air as well as dust are ubiquitous throughout the environment, and are composed of fungal spores, fragments of fungi, bacteria, (toxic) complex organic compounds, as well as fragments and feces of insects and similar organisms. In addition, human responses to microbial contaminants vary over a tremendous range and it is not possible to sample and analyze for all possible microbial contaminants by a single method. Therefore, the standard model for acceptable indoor environmental conditions prescribes that the quantity and types of fungi present in the indoor environment should not be significantly different from the general outdoor environment or a suitable indoor control zone.

Typically, total fungal spore levels detected on samples collected inside occupied structures should be less than levels detected in outdoor ambient air. Additionally, comparison of indoor and outdoor air samples should demonstrate a similarity in the fungal species identified. Outdoor ambient spore levels can vary dramatically with changes in environmental conditions. Variations in weather conditions may affect ambient outdoor spore levels and result in conditions where indoor levels may exceed outdoor levels. Indicator species are those that are commonly found in moisture impacted structures and in some case are capable of producing mycotoxins. The presence of indicator species is used to evaluate the potential impact that any moisture intrusion has had on a building. In the absence of specific regulatory and health-based standards, regarding acceptable indoor levels of microbiological contaminants, careful qualitative evaluation of the data obtained is used to determine if bio-amplification is occurring. For the purposes of this report, the clearance criteria was defined by the Client as such; Levels of total fungal spore counts within the area of concern (AOC) shall be less than total fungal spore counts of exterior and/or indoor control samples.

Laboratory results of AOC and outdoor environmental air samples were evaluated based on both total spore levels and commonality of species detected.

South Wing, Multi-Purpose Room & Cafeteria results on August 27<sup>th</sup> indicated elevated levels of fungi with *Aspergillus/Penicillium-like* being the predominant species. Re-cleaning and SteraMist of South Wing was performed and passed on August 28<sup>th</sup>.

West Wing and Adjacent Rooms results on August 28<sup>th</sup> indicated elevated levels of *Aspergillus/Penicillium*. Re-cleaning and SteraMist of West Wing was performed and passed on August 29<sup>th</sup>.

North Wing clearance results passed on August 29<sup>th</sup>. Main Office and Room. N21 were re-cleaned and SteraMist. These areas were passed the following day

East Wing was resampled on August 30<sup>th</sup> after being re-cleaned and SteraMist, and clearance results were passed the following day.

Library / Core Wing & Gym were tested on September 1<sup>st</sup> and passed. Kitchen Pantry was re-cleaned on September 2<sup>nd</sup> along with Women's & Men's Restrooms by the Gym, and passed the following day by clearance results.

The Mechanical Room by Library were re-cleaned on September 4<sup>th</sup> and passed that night following clearance results.

Clearance and Re-Sampling Locations are identified in Table 1.0

It was a pleasure to work with you and your staff while dealing with this issue. I hope that the information contained within this letter is sufficient for your needs and we look forward to working with you again in the safety and environmental consulting area.

Should you have any questions or concerns please feel free to contact my office for assistance.

Regards,



Tanay Ranadive  
Safety & Environmental Services



**Report Limitations and Disclaimer**

Microbiological organisms are ubiquitous opportunistic allergenic organisms whose concentration is greatly affected by changes in localized ambient environmental conditions. Assessment for microbiological contamination is limited to collection and evaluation of data relating to general ambient environmental conditions, detected as present, at the time of the evaluation. Demolition or disassembly of building surfaces and installed equipment are not performed as part of the evaluation. QuES&T believes this report is based on reliable current industry practices/references/sources and accurately reflects the general conditions existing in the area inspected at the time of our assessment. However, unobserved or concealed conditions and/or variations in localized ambient environmental conditions may significantly affect reported microbiological contamination levels.

The Parties agree and understand that the presence of mold and the evolving understanding of risks which may be associated with human exposure to certain types of mold represent an area of medical, scientific and industry knowledge which is only beginning to mature and that this area of knowledge at present is, at best, incomplete. The parties agree and understand that mold is mobile; it can arise in new places and recur in areas which have been remediated due to limitations in detection or removal methods (spores are microscopic), limitations in time and cost, new and modified or previously unknown water intrusion and/or accumulation events and processes beyond the control of QuES&T. Accordingly, QuES&T is not liable for such new or recurring mold growths. Further, due to the microscopic nature of mold spores, it is agreed and understood that **no warranty or promise that all mold has been identified or removed is made or intended by QuES&T.** Assessments of water intrusion or accumulation risk by QuES&T, if any, are not to be understood as a complete list of potential ways in which water intrusion or accumulation may occur at the Site(s) subject to this Agreement. Client further recognizes the unsettled liability environment surrounding mold. Therefore, as a fundamental incentive to **Quality Environmental Solutions & Technologies Inc. (QuES&T)** to undertake the provision of services to Client, Client agrees that QuES&T will be deemed to have fully complied with any contractual standards of performance or any legal mandate of non-negligent behavior by providing QuES&T's services consistent with Proposal No. P16-4635, and signed by all parties as of February 2, 2016. Limitation of Liability shall be the cost of services. Client hereby agrees to indemnify, defend and hold harmless QuES&T its joint ventures, affiliates, parent and subsidiary entities and the employees, officers, directors, representatives and agents of QuES&T, and all of the foregoing from and against any and all claims, suits, causes of actions, liabilities, costs (including but not limited to reasonable attorney's fees) and judgments which are based in whole or in part upon (or which sound in) mold-based liability, except to the extent of the sole negligence of QuES&T and the other Indemnitees set out immediately preceding, but subject always to the Limitation of Liability. ***NO OTHER WARRANTY, EXPRESS OR IMPLIED, IS MADE OR INTENDED HEREBY AND ANY AND ALL OTHER SUCH WARRANTIES ARE HEREBY FULLY AND COMPLETELY DISCLAIMED BY QuES&T.***

# Appendix A

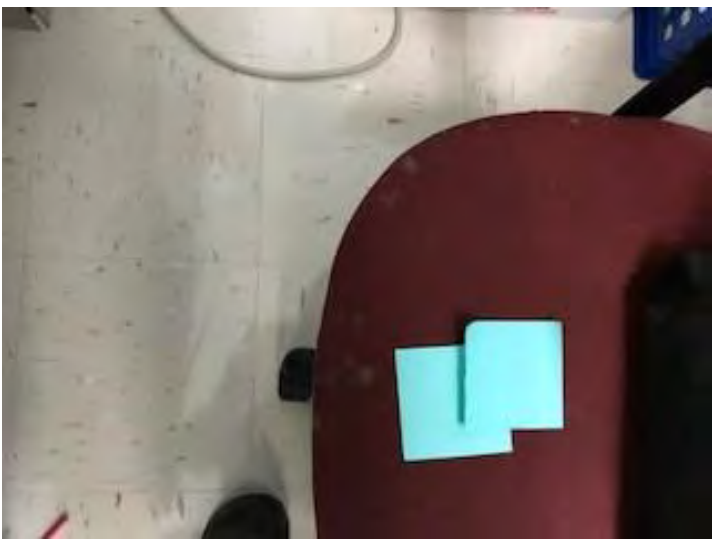
## Photos



1.1



1.2



1.3



1.4



1.5

Figures 1.1-1.5 Above Show Spots of Microbial Growth On Desks and Chairs



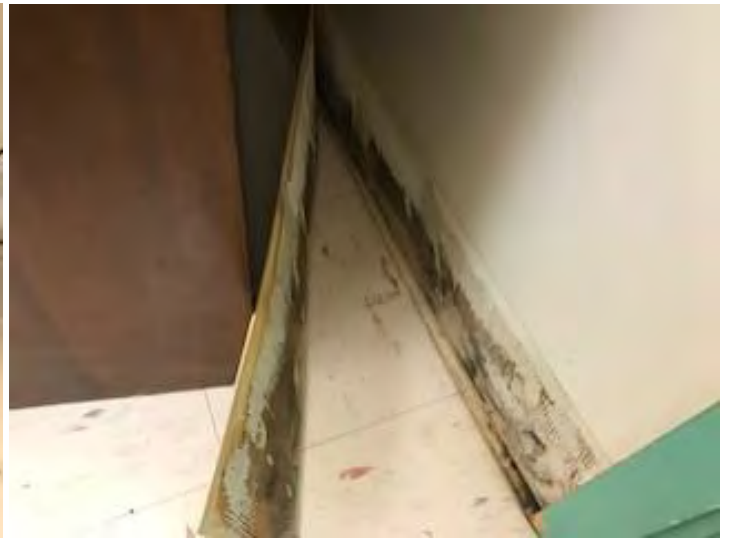
1.6



1.7



1.8



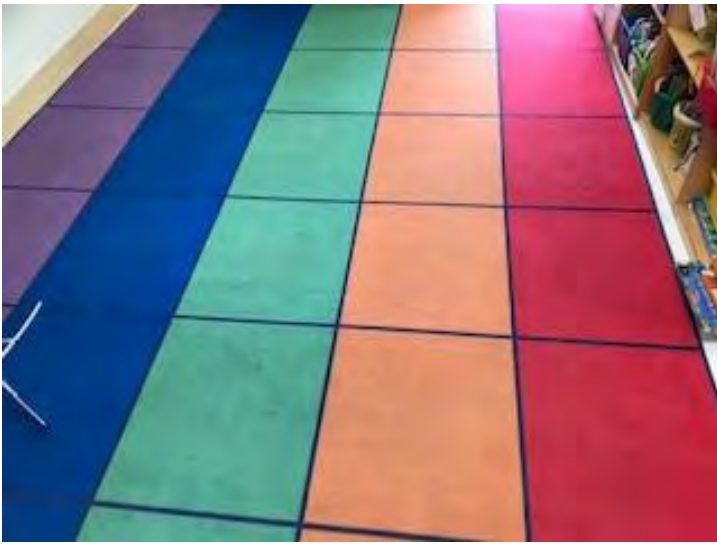
1.9



1.10

Figures 1.6-1.10 Above Show Microbial Growth Hidden Behind Cove Base Molding & On Adjacent Sheetrock





1.11



1.12



1.13



1.14



1.15



1.16

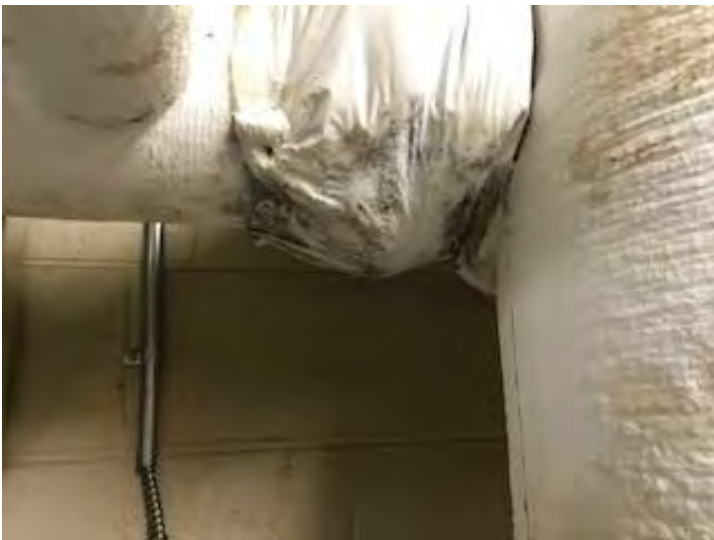
Figures 1.11-1.16 Above Show Microbial Growth Hidden Behind Rugs, Mats, and Boards



1.17



1.18



1.19



1.20



1.21

Figures 1.17-1.21 Above Show Microbial Growth On and Around Pipes and Pipe Insulation





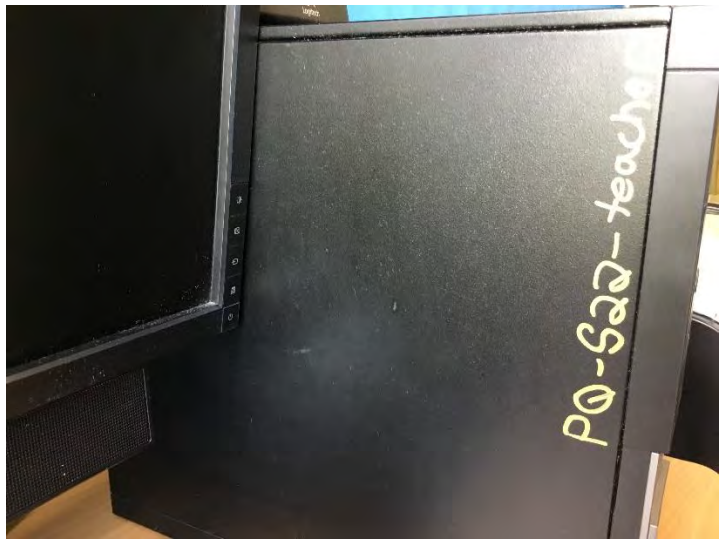
1.22



1.23



1.24



1.25

Figures 1.22-1.25 Above Show Microbial Growth on Various Materials (Wood, Plastic, Cloth)

# Appendix B

## Analytical Data





# EXPEDITE

256 Bridge Street, Metuchen, NJ 08840, USA

## Chain of Custody

# EXPEDITE

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: <small>(lab use only)</small> ME180826-02	Telephone No.: 825-559-8537	Company Contact: Louis N Johnson III
Company Name: QUEST	Please select: Fax Report ( ) or Email Report (X)	Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY 12590	Fax No.:	Date/Time sampled: 08/26/18 12:00
	Email address: tranadive@qualityenv.com	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Rm. S25	FD-01HP			3 Hr	Air-O-cell	165 L	2574-0533
1941-02	Southwing 1 <sup>st</sup> grade Hallway	"			"	"	150 L	2574-0517
1941-03	Rm. S23, Kindergarten	"			"	"	150 L	2574-0531
1941-04	South wing, Kindergarten Hall	"			"	"	150 L	2574-0595
1941-05	South wing by Bathroom	"			"	"	150 L	2574-0548
1941-06	Rm. SPO	"			"	"	150 L	2574-0493
1941-07	Environmental, Rear of Bld	"			"	"	150 L	2574-0525
1941-08	outside Bld, Kindergarten Hall	"			"	"	150 L	2574-0459
1941-09	Batch Blank	"			"	"	1	2574-0623
1941-10	Lab <sup>(M)</sup> Field Blanks	"			"	"		2574-0613

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.

**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Tanya Kanadive (print) Tanya Kanadive Date submitted: 08/26/18  
 Received by: (sign) [Signature] (print) Wei Tang Date and time received: 08/26/18 3:13 PM



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

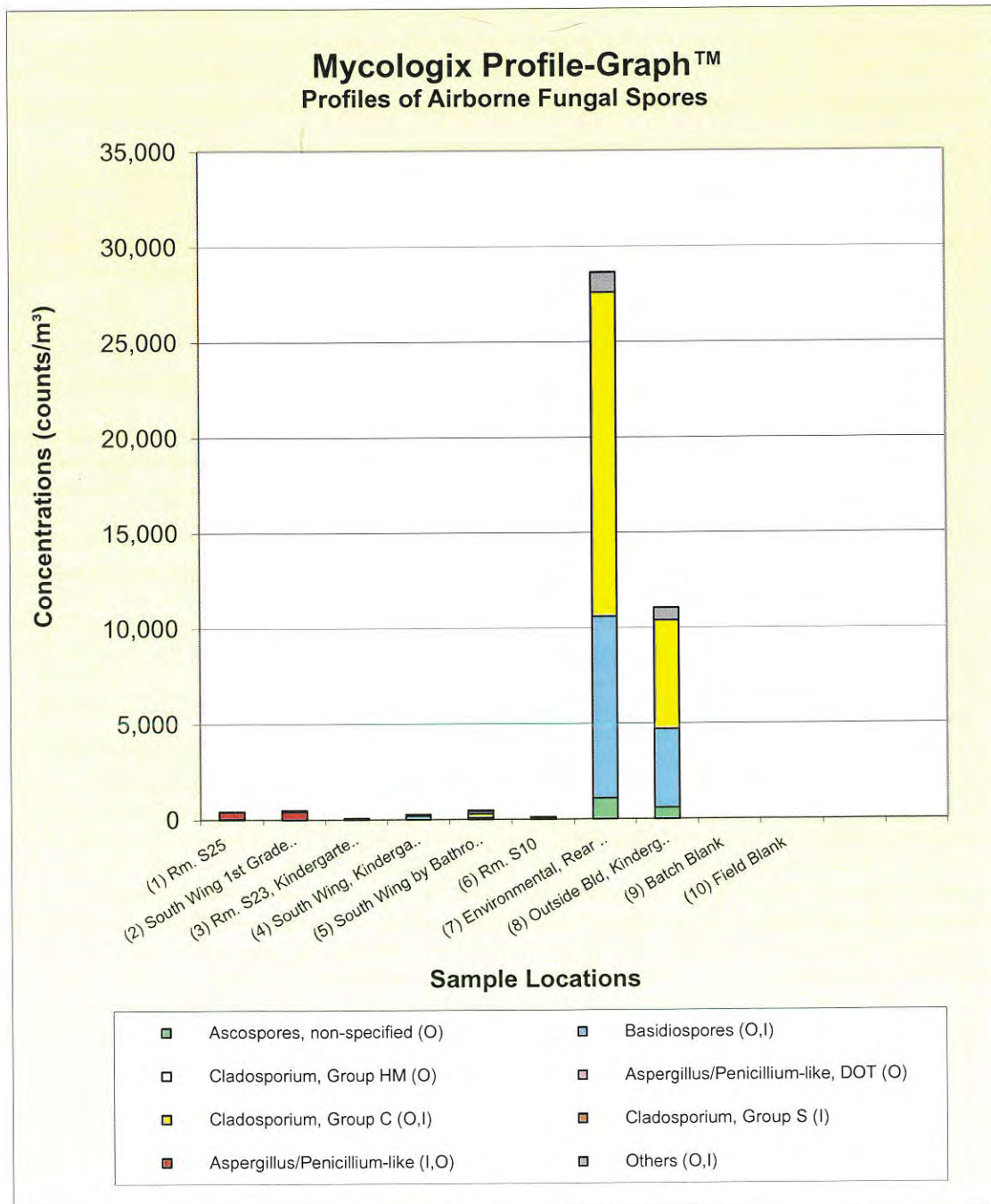
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







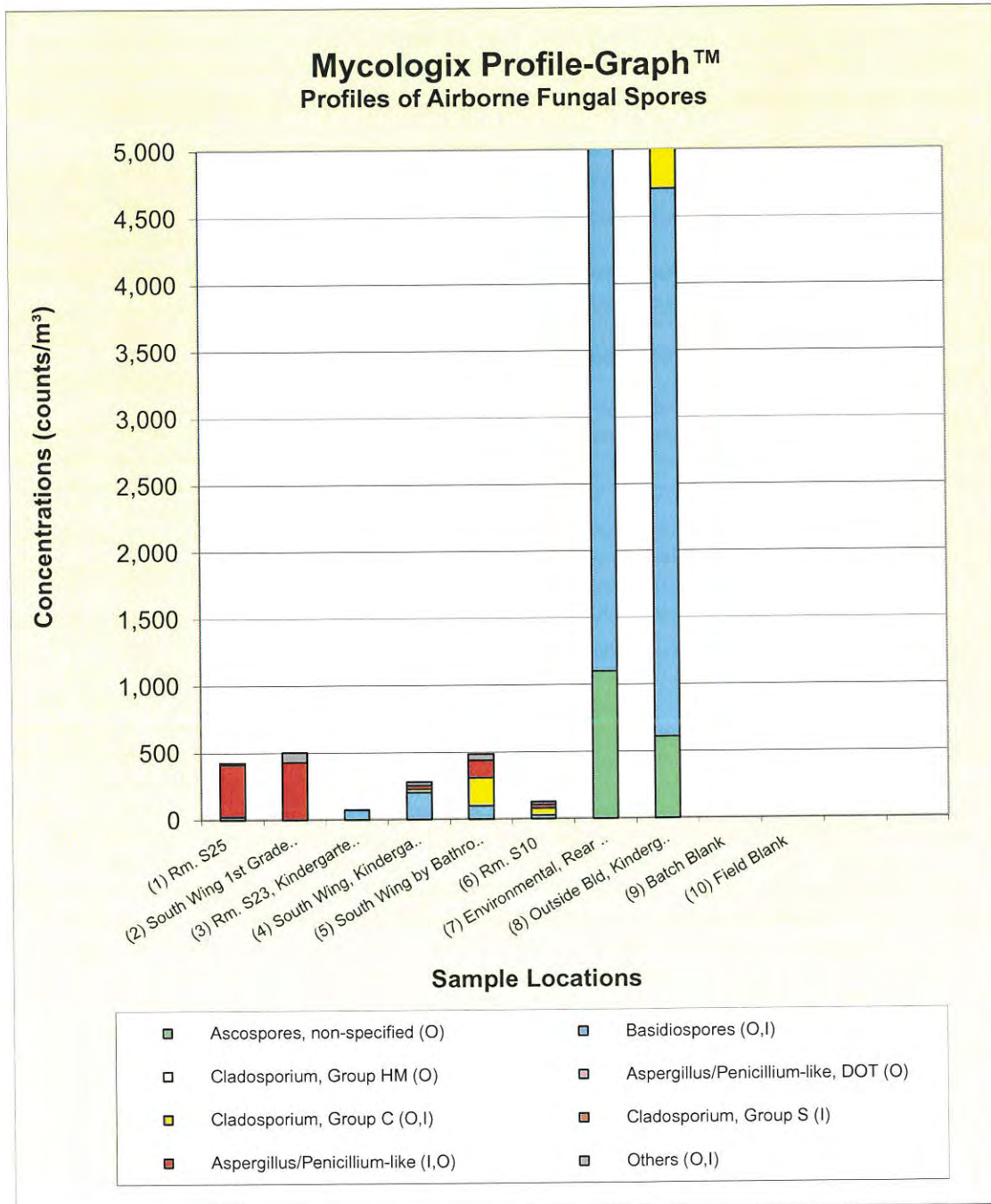
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AccuScience™  
Analysis Report

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**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180826-02(1)			ME180826-02(2)			ME180826-02(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Rm. S25			South Wing 1st Grade Hallway			Rm. S23, Kindergarten		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	165 L			150 L			150 L		
Total Concentration (counts/m³)**	430 cts/m³			500 cts/m³			73 cts/m³		
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 24; LQL = 480 cts/m³			DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	4	24	6				11	73	100
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O)									
#Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O)	64	390	92	64	430	85			
## Cluster-Chain-Loose Spore Profile™		0% - 53% - 47%			0% - 0% - 100%				
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 6; LQL = 120 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 6; LQL = 120 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	1	6	1	1	7	1			
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)				2	13	3			
Nigrospora (O)									
Pithomyces (O)				7	47	9			
Rusts (O)									
Unknown (O,I)	1	6	1	1	7	1			
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

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AIHA EMPAT Lab ID: 178794

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**Contact:** Johnson, Louis, III  
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**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(4)			ME180826-02(5)			ME180826-02(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	South Wing, Kindergarten Hall			South Wing by Bathroom			Rm. S10		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			150 L			150 L		
Total Concentration (counts/m³)**	280 cts/m³			490 cts/m³			130 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	30	200	71	15	100	21	4	27	21
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	4	27	10	31	210	43	8	53	42
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	4	27	10	19	130	27	4	27	21
Cluster(s)	0% - 0% - 100%			0% - 0% - 100%			0% - 0% - 100%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)				1	7	1			
Chaetomium (I)	1	7	2	1	7	1			
Ulocladium (I)				1	7	1			
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	1	7	2				1	7	5
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)				1	7	1			
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)									
Nigrospora (O)									
Pithomyces (O)				2	13	3	1	7	5
Rusts (O)									
Unknown (O,I)	2	13	5	1	7	1	1	7	5
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(7)			ME180826-02(8)			ME180826-02(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	Environmental, Rear of Bld			Outside Bld, Kindergarten Hall			Batch Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			150 L			1 smp		
Total Concentration (counts/m³)**	29,000 cts/m³			11,000 cts/m³			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 4 cts/smp		
Ascospores, non-specified (O)	166	1,100	4	91	610	6			
Basidiospores (O,I)	1,419	9,500	33	612	4,100	37			
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	2,567	17,000	59	853	5,700	52			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™									
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp		
Hyphal fragment (O,I)	4	27	<1	7	47	<1			
Alternaria (O,I)	2	13	<1	6	40	<1			
Cercospora (O)									
Curvularia (O,I)	22	150	<1	7	47	<1			
Drechslera/Bipolaris-like (O)				2	13	<1			
Epicoccum (O)	8	53	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	41	270	<1	4	27	<1			
Nigrospora (O)	2	13	<1						
Pithomyces (O)	38	250	<1	63	420	4			
Rusts (O)	19	130	<1						
Unknown (O,I)	21	140	<1	8	53	<1			
<b>Skin Cells Rating</b>	None			None			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			0 (None detected)		
<b>Note</b>							No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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Analysis Report

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info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(10)		
Sample ID	1941-10		
Sample Location	Field Blank		
Sample Type (Device)	Air (Air-O-Cell)		
Air Volume	1 smp		
Total Concentration (counts/m <sup>3</sup> )**	< DL cts/smp		
<b>Mycologix Profile Group 1, 2 &amp; 3</b>	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp		
Ascospores, non-specified (O)			
Basidiospores (O,I)			
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)			
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™			
Cluster(s)			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp		
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memnoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 1 cts/smp		
Hyphal fragment (O,I)			
Alternaria (O,I)			
Cercospora (O)			
Curvularia (O,I)			
Drechslera/Bipolaris-like-(O)			
Epicoccum (O)			
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)			
Nigrospora (O)			
Pithomyces (O)			
Rusts (O)			
Unknown (O,I)			
<b>Skin Cells Rating</b>	None		
<b>Debris Rating</b>	0 (None detected)		
<b>Note</b>	No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





# EXPEDITE

## Chain of Custody

# EXPEDITE

256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: <small>(lab use only)</small> ME180826-01	Telephone No.: 845-559-8537	Company Contact: Tara Banadive
Company Name: QUES&T	Please select: Fax Report ( ) or Email Report (X)	Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY 12590	Fax No.:	Date/Time sampled: 08/26/18 13:00
	Email address: taradive@qualityenv.com	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01-S18	Rm. S18 Southwing	FD-01HP			3 Hr	Air-O-cell	150L	2574-0523
1941-02-S18	Batch Blanks	"			"	"		2574-0513
1941-03-S18	Field Blank	"			"	"		2574-0527

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.

**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (4) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Tara Banadive (print) Tara Banadive Date submitted: 08/26/18

Received by: (sign) Wei Tang (print) Wei Tang Date and time received: 08/26/18 3:13 PM





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

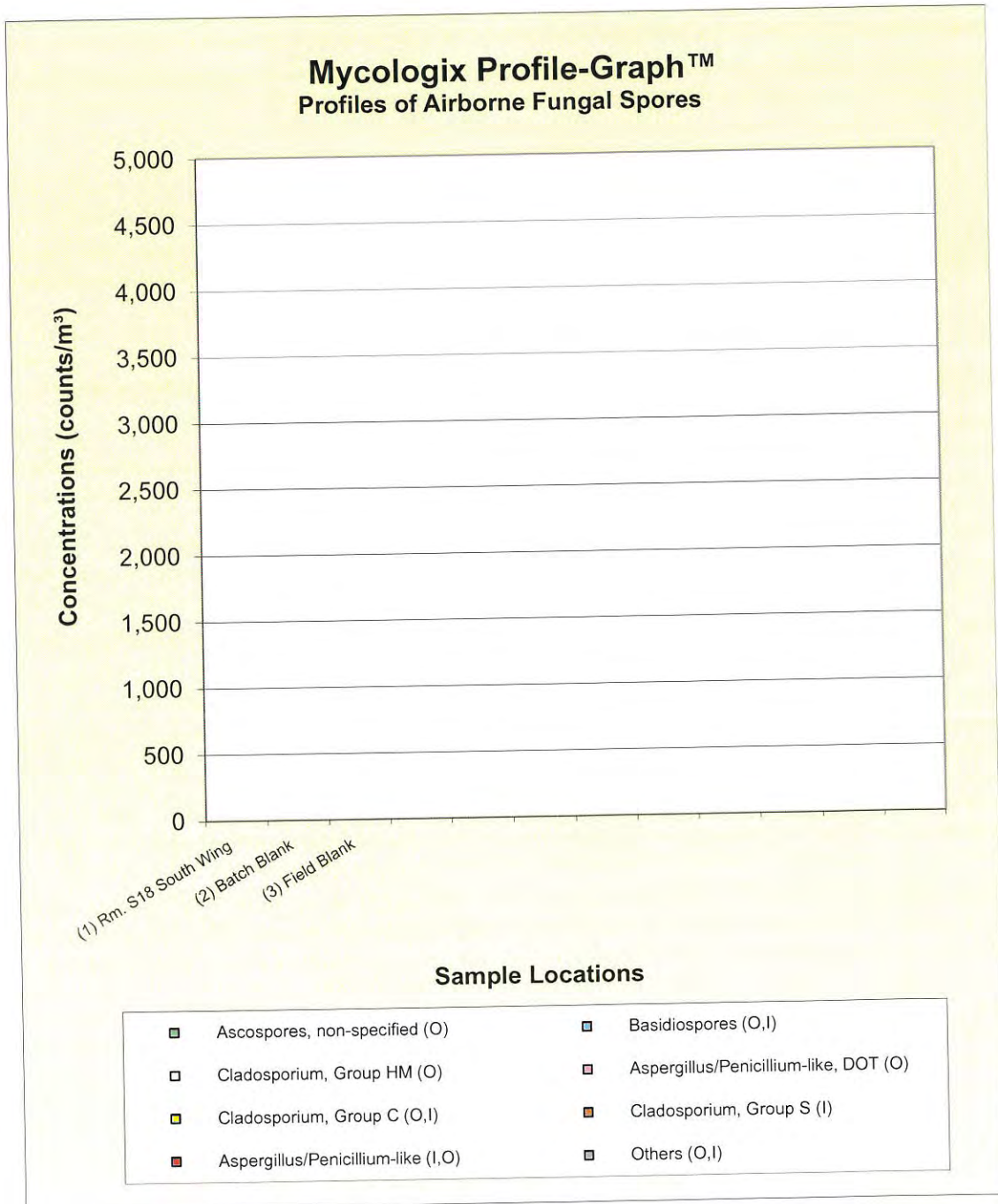
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-01  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





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Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-01  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180826-01(1)	ME180826-01(2)	ME180826-01(3)
<b>Sample ID</b>	<b>1941-01-S18</b>	<b>1941-02-S18</b>	<b>1941-03-S18</b>
<b>Sample Location</b>	Rm. S18 South Wing	Batch Blank	Field Blank
<b>Sample Type (Device)</b>	Air (Air-O-Cell)	Air (Air-O-Cell)	Air (Air-O-Cell)
<b>Air Volume</b>	150 L	1 smp	1 smp
<b>Total Concentration (counts/m³)**</b>	< DL cts/m³	< DL cts/smp	< DL cts/smp
<b>Mycologix Profile Group 1, 2 &amp; 3</b>	cts/smp* counts/m³ %	cts/smp* counts/smp %	cts/smp* counts/smp %
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³	DL = 4 cts/smp	DL = 4 cts/smp
Ascospores, non-specified (O)			
Basidiospores (O,I)			
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)			
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™			
Cluster(s)			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³	DL = 1 cts/smp	DL = 1 cts/smp
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memnoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³	DL = 1 cts/smp	DL = 1 cts/smp
Hyphal fragment (O,I)			
Alternaria (O,I)			
Cercospora (O)			
Curvularia (O,I)			
Drechslera/Bipolaris-like (O)			
Epicoccum (O)			
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)			
Nigrospora (O)			
Pithomyces (O)			
Rusts (O)			
Unknown (O,I)			
<b>Skin Cells Rating</b>	None	None	None
<b>Debris Rating</b>	1 (≤ 5%)	0 (None detected)	0 (None detected)
<b>Note</b>	No fungal structure observed	No fungal structure observed	No fungal structure observed

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





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Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180826-02(1)			ME180826-02(2)			ME180826-02(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Rm. S25			South Wing 1st Grade Hallway			Rm. S23, Kindergarten		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	165 L			150 L			150 L		
Total Concentration (counts/m³)**	430 cts/m³			500 cts/m³			73 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 24; LQL = 480 cts/m³			DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	4	24	6				11	73	100
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	64	390	92	64	430	85			
Cluster(s)		0% - 53% - 47%			0% - 0% - 100%				
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 6; LQL = 120 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 6; LQL = 120 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Hypal fragment (O,I)	1	6	1	1	7	1			
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)				2	13	3			
Nigrospora (O)				7	47	9			
Pithomyces (O)									
Rusts (O)									
Unknown (O,I)	1	6	1	1	7	1			
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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AIHA EMPAT Lab ID: 178794

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**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(4)			ME180826-02(5)			ME180826-02(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	South Wing, Kindergarten Hall			South Wing by Bathroom			Rm. S18		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			150 L			150 L		
Total Concentration (counts/m³)**	280 cts/m³			490 cts/m³			130 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³			DL = 27; LQL = 530 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	30	200	71	15	100	21	4	27	21
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	4	27	10	31	210	43	8	53	42
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™	4	27	10	19	130	27	4	27	21
Cluster(s)	0% - 0% - 100%			0% - 0% - 100%			0% - 0% - 100%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)				1	7	1			
Chaetomium (I)	1	7	2	1	7	1			
Ulocladium (I)				1	7	1			
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	1	7	2				1	7	5
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)				1	7	1			
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)									
Nigrospora (O)									
Pithomyces (O)				2	13	3	1	7	5
Rusts (O)									
Unknown (O,I)	2	13	5	1	7	1	1	7	5
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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AIHA EMPAT Lab ID: 178794

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**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
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**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(7)			ME180826-02(8)			ME180826-02(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	Environmental, Rear of Bld			Outside Bld, Kindergarten Hall			Batch Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			150 L			1 smp		
Total Concentration (counts/m³)**	29,000 cts/m³			11,000 cts/m³			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 4 cts/smp		
Ascospores, non-specified (O)	166	1,100	4	91	610	6			
Basidiospores (O,I)	1,419	9,500	33	612	4,100	37			
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	2,567	17,000	59	853	5,700	52			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™ Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp		
Hyphal fragment (O,I)	4	27	<1	7	47	<1			
Alternaria (O,I)	2	13	<1	6	40	<1			
Cercospora (O)									
Curvularia (O,I)	22	150	<1	7	47	<1			
Drechslera/Bipolaris-like (O)				2	13	<1			
Epicoccum (O)	8	53	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	41	270	<1	4	27	<1			
Nigrospora (O)	2	13	<1						
Pithomyces (O)	38	250	<1	63	420	4			
Rusts (O)	19	130	<1						
Unknown (O,I)	21	140	<1	8	53	<1			
<b>Skin Cells Rating</b>	None			None			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			0 (None detected)		
<b>Note</b>							No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180826-02  
**Date Received:** 8/26/2018  
**Date Analyzed:** 8/26/2018  
**Date Reported:** 8/26/2018

Lab Sample No.	ME180826-02(10)		
Sample ID	1941-10		
Sample Location	Field Blank		
Sample Type (Device)	Air (Air-O-Cell)		
Air Volume	1 smp		
Total Concentration (counts/m³)**	< DL cts/smp		
<b>Mycologix Profile Group 1, 2 &amp; 3</b>	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp		
Ascospores, non-specified (O)			
Basidiospores (O,I)			
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)			
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp		
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memnoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 1 cts/smp		
Hyphal fragment (O,I)			
Alternaria (O,I)			
Cercospora (O)			
Curvularia (O,I)			
Drechslera/Bipolaris-like (O)			
Epicoccum (O)			
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)			
Nigrospora (O)			
Pithomyces (O)			
Rusts (O)			
Unknown (O,I)			
<b>Skin Cells Rating</b>	None		
<b>Debris Rating</b>	0 (None detected)		
<b>Note</b>	No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.

RUSH!

RUSH!



Chain of Custody

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)
Tel: 856-489-0011 www.QLabUSA.com

256 Bridge Street, Metuchen, NJ 08840, USA

Lab Job No.: ME180827-05
Telephone No.: 845-559-8537
Company Contact: Tanay Ranadive
Company Name: QUES&T
Please select: Fax Report ( ) or Email Report (x)
Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY 12590
Fax No.:
Date/Time sampled: 08/26/18 15:00
Email address: tranadive@qualityenv.com
P.O. No.:

Table with 7 columns: Sample ID, Sample Location, Analysis Code, Turnaround Time (Std, Day, Hr), Sample Type, Volume (L) or Area (in^2), Note. Rows include: 1941-01 Multipurpose Room, 1941-02 Cafeteria, 1941-03 Environmental, 1941-04 outside Bld. by Multipurpose, 1941-05 Batch Blanks, 1941-06 Field Blanks.

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP.
Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive Date submitted: 08/26/18
Received by: (sign) Maggie Lim (print) Maggie Lim Date and time received: 08/27/18 11:21 AM
Page 1 of 2 QLAB\_C-O-C\_V4.01



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™

**Client:** QuES&T  
Wappingers Falls, NY

**Contact:** Ranadive, Tanay

**Project ID:** Q18-1941

**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180827-05

**Date Received:** 8/27/2018

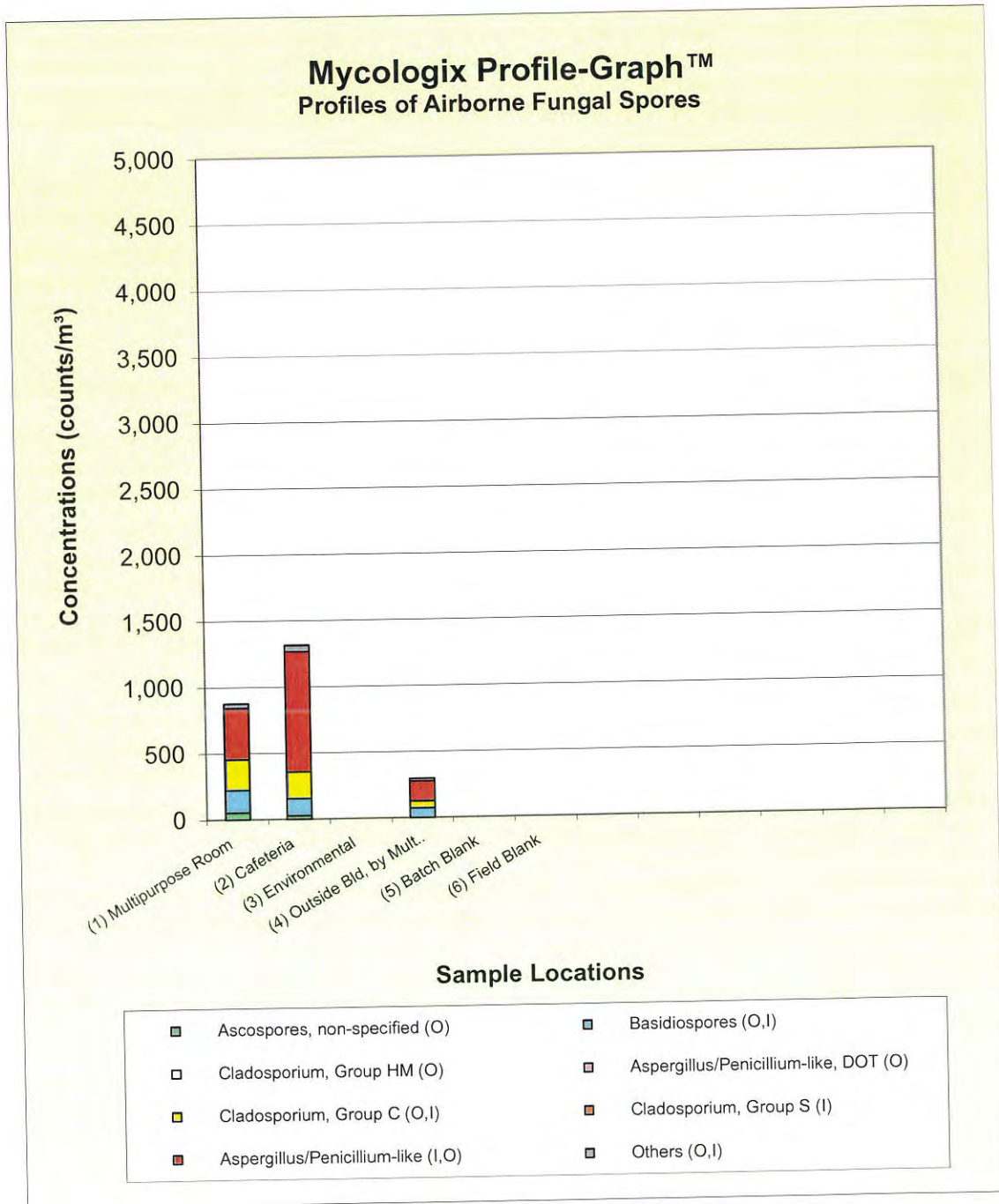
**Date Analyzed:** 8/27/2018

**Date Reported:** 8/27/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180827-05  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No. Sample ID	ME180827-05(1) 1941-01	ME180827-05(2) 1941-02	ME180827-05(3) 1941-03
Sample Location	Multipurpose Room	Cafeteria	Environmental
Sample Type (Device)	Air (Air-O-Cell)	Air (Air-O-Cell)	Air (Air-O-Cell)
Air Volume	150 L	150 L	150 L
Total Concentration (counts/m³)**	880 cts/m³	1,300 cts/m³	< DL cts/m³
<b>Mycologix Profile Group 1, 2 &amp; 3</b>	cts/smp*    counts/m³    %	cts/smp*    counts/m³    %	cts/smp*    counts/m³    %
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³	DL = 27; LQL = 530 cts/m³	DL = 27; LQL = 530 cts/m³
Ascospores, non-specified (O)	8    53    6	4    27    2	
Basidiospores (O,I)	26    170    19	19    130    10	
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)	34    230    26	30    200    15	
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™	58    390    45	137    910    69	
Cluster(s)	0% - 28% - 72%	9% - 57% - 34%	1 cluster(s) of 13 spores
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³	DL = 7; LQL = 130 cts/m³	DL = 7; LQL = 130 cts/m³
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memnoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³	DL = 7; LQL = 130 cts/m³	DL = 7; LQL = 130 cts/m³
Hyphal fragment (O,I)	1    7    <1		
Alternaria (O,I)			
Cercospora (O)			
Curvularia (O,I)		1    7    <1	
Drechslera/Bipolaris-like (O)			
Epicoccum (O)			
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)		1    7    <1	
Nigrospora (O)			
Pithomyces (O)	3    20    2	1    7    <1	
Rusts (O)			
Unknown (O,I)	1    7    <1	4    27    2	
<b>Skin Cells Rating</b>	Trace	Trace	None
<b>Debris Rating</b>	2 (6 - 25%)	1 (≤ 5%)	1 (≤ 5%)
<b>Note</b>			No fungal structure observed

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/26/2018

**QLab Job No.:** ME180827-05  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Lab Sample No.	ME180827-05(4)	ME180827-05(5)	ME180827-05(6)
Sample ID	1941-04	1941-05	1941-06
Sample Location	Outside Bld, by Multipurpose	Batch Blank	Field Blank
Sample Type (Device)	Air (Air-O-Cell)	Air (Air-O-Cell)	Air (Air-O-Cell)
Air Volume	150 L	1 smp	1 smp
Total Concentration (counts/m³)**	300 cts/m³	< DL cts/smp	< DL cts/smp
Mycologix Profile Group 1, 2 & 3	cts/smp* counts/m³ %	cts/smp* counts/smp %	cts/smp* counts/smp %
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³	DL = 4 cts/smp	DL = 4 cts/smp
Ascospores, non-specified (O)			
Basidiospores (O,I)	11 73 25		
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O)			
#Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)	8 53 18		
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O)	23 150 51		
## Cluster-Chain-Loose Spore Profile™	0% - 17% - 83%		
Cluster(s)			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³	DL = 1 cts/smp	DL = 1 cts/smp
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memnoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³	DL = 1 cts/smp	DL = 1 cts/smp
Hyphal fragment (O,I)			
Alternaria (O,I)			
Cercospora (O)			
Curvularia (O,I)			
Drechslera/Bipolaris-like (O)			
Epicoccum (O)	1 7 2		
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)	2 13 4		
Nigrospora (O)			
Pithomyces (O)			
Rusts (O)			
Unknown (O,I)			
<b>Skin Cells Rating</b>	Trace	None	None
<b>Debris Rating</b>	1 (≤ 5%)	0 (None detected)	0 (None detected)
<b>Note</b>		No fungal structure observed	No fungal structure observed

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





**EXPEDITE**

**Chain of Custody**

**EXPEDITE**

256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: <small>(lab use only)</small> ME180827-18	Telephone No.: 845-559-8537	Company Contact: Tanay Ranadive
Company Name: QUES&T	Please select: Fax Report ( ) or Email Report (✓)	Project ID: Q18-1941 Centerwing
Company Address: 1376 Route 9 Wappingers Falls, NY 12590	Fax No.:	Date/Time sampled: 08'27'18'16'00
	Email address: tranadive@qualityenv.com	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note Serial number (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	O/S Background Pre-Sample 1614-1624	FD-01HP			3 Hr	Air-O-Cell	150 L	2574-0539
1941-02	Main Office 1628-1633	"			"	"	75 L	2574-0624
1941-03	Center Wing Hallway 1629-1634	"			"	"	75 L	2574-0559
1941-04	Multi-purpose 1630-1636	"			"	"	90 L	2574-0548
1941-05	Hallway by Rm. 58 1631-1637	"			"	"	90 L	2574-0515
1941-06	O/S Background post sample 1636-1646	"			"	"	150 L	2574-0588
1941-07	Batch Blank							
1941-08	Field Blank							

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.

**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive Date submitted: 08/27/18  
 Received by: (sign) [Signature] (print) Wen Tang Date and time received: 08/27/18 7:23 PM



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

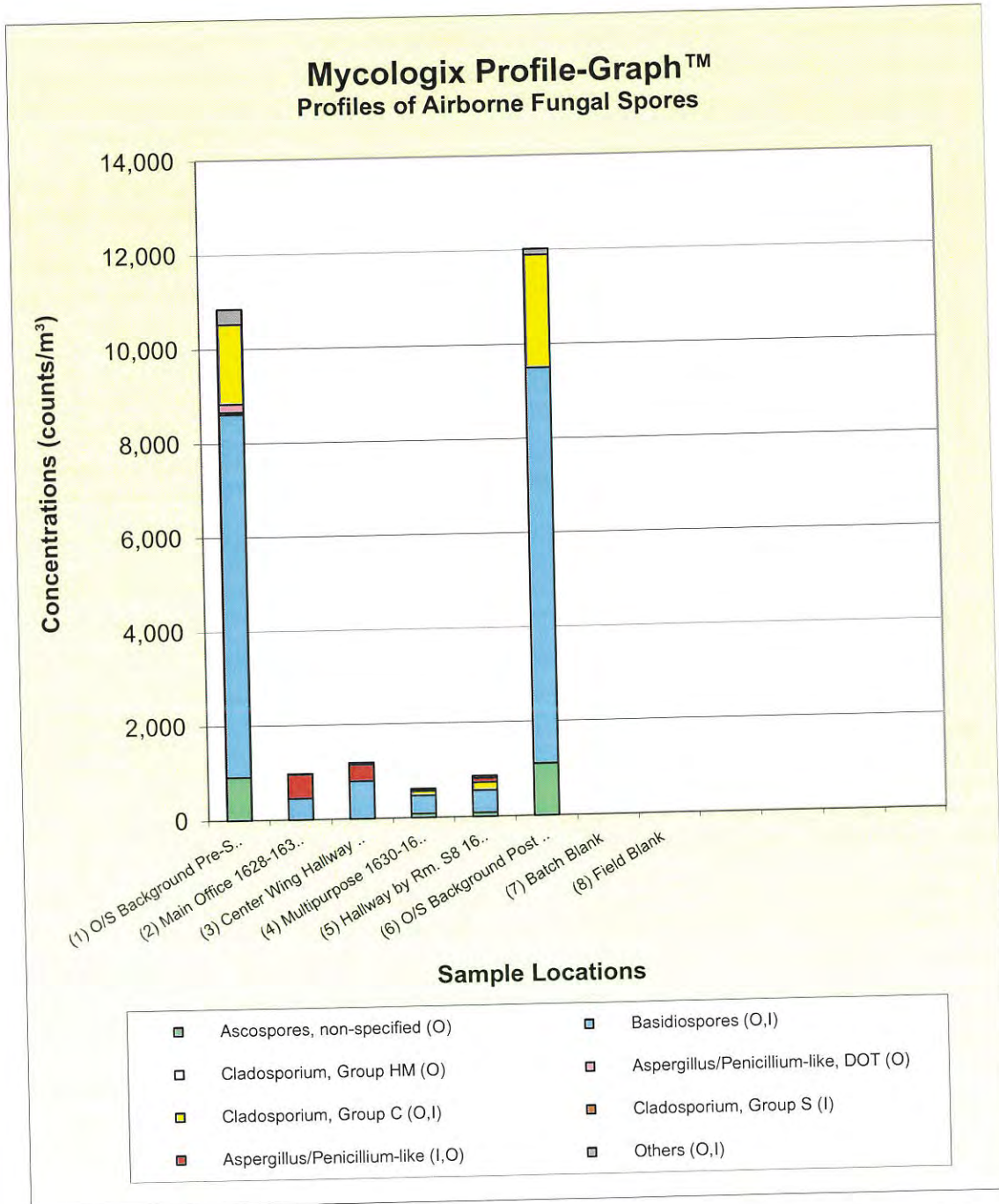
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Center Wing  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-18  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Center Wing  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-18  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180827-18(1)			ME180827-18(2)			ME180827-18(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	O/S Background Pre-Sample 1614-1624			Main Office 1628-1633			Center Wing Hallway 1629-1634		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	11,000 cts/m³			970 cts/m³			1,200 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	136	910	8						
Basidiospores (O,I)	1,155	7,700	71	34	450	46	60	800	67
Cladosporium, Group HM (O)	8	53	<1						
Aspergillus/Penicillium-like, DOT (O)	26	170	2						
#Cluster-Chain-Loose Spore Profile™		73% - 27% - 0%							
Cladosporium, Group C (O,I)	249	1,700	16						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O)				38	510	52	26	350	29
### Cluster-Chain-Loose Spore Profile™					0% - 60% - 40%			0% - 0% - 100%	
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	4	27	<1						
Alternaria (O,I)	1	7	<1						
Cercospora (O)	1	7	<1						
Curvularia (O,I)	16	110	1						
Drechslera/Bipolaris-like (O)	2	13	<1						
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	7	47	<1				3	40	3
Nigrospora (O)									
Pithomyces (O)	14	93	<1						
Rusts (O)									
Unknown (O,I)	2	13	<1	1	13	1			
<b>Skin Cells Rating</b>	None			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Center Wing  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-18  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Lab Sample No.	ME180827-18(4)			ME180827-18(5)			ME180827-18(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Multipurpose 1630-1636			Hallway by Rm. S8 1631-1637			O/S Background Post Sample 1636-1646		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	90 L			90 L			150 L		
Total Concentration (counts/m³)**	610 cts/m³			860 cts/m³			12,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 44; LQL = 890 cts/m³			DL = 44; LQL = 890 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	8	89	15	8	89	10	159	1,100	9
Basidiospores (O,I)	34	380	62	42	470	55	1,253	8,400	70
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O)									
#Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	8	89	15	15	170	20	355	2,400	20
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O)	4	44	7	8	89	10			
### Cluster-Chain-Loose Spore Profile™		0% - 0% - 100%			0% - 0% - 100%				
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 11; LQL = 220 cts/m³			DL = 11; LQL = 220 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 11; LQL = 220 cts/m³			DL = 11; LQL = 220 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)				1	11	1	1	7	<1
Alternaria (O,I)									
Cercospora (O)							1	7	<1
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)							7	47	<1
Nigrospora (O)							1	7	<1
Pithomyces (O)				2	22	3	4	27	<1
Rusts (O)									
Unknown (O,I)	1	11	2	1	11	1	4	27	<1
<b>Skin Cells Rating</b>	Trace			Low			None		
<b>Debris Rating</b>	1 (≤ 5%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Center Wing  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-18  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Lab Sample No.	ME180827-18(7)			ME180827-18(8)		
Sample ID	1941-07			1941-08		
Sample Location	Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m <sup>3</sup> )**	< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™						
Cluster(s)						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memnoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected)			0 (None detected)		
<b>Note</b>	No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





256 Bridge Street, Metuchen, NJ 08840, USA

### Chain of Custody

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: (lab use only) <b>ME180827-19</b>	Telephone No.: <b>845-559-8537</b>	Company Contact: <b>Taney Ranadive</b>
Company Name: <b>QES&amp;T</b>	Please select: Fax Report ( ) or Email Report (✓)	Project ID: <b>Q18-1941 (South Wing)</b>
Company Address: <b>1376 Route 9, Wappingers Falls, NY 12590</b>	Fax No.:	Date/Time sampled: <b>08 '27 '18 '16 '50</b>
	Email address: <b>tranadive@qualityenv.com</b>	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1 1941-01	o/s Background Pre-Sample 1653-1703	FD-01HP			3Hr	Air-O-Cell	150L 2574-0576	
2 1941-02	Rm. 510 1706-1711	"			"	"	75L 2574-0498	
3 1941-03	2 <sup>nd</sup> Grade Hallway by Bathroom 1707-1712	"			"	"	75L 2574-0556	
4 1941-04	2 <sup>nd</sup> Grade Hallway 1708-1713	"			"	"	75L 2574-0594	
(7) 1941-0507	o/s Background Post-Sample 1715-1725	"			"	"	150L 2574-0567	
6 1941-06	Library Pre-Clean 1653-1658	"			"	"	75L 2574-0645	
(5) 1941-05	Rm. 525 1709-1714	"			"	"	75L 2574-0609	
8 1941-08	Batch Blank							
9 1941-09	Field Blank							

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Taney Ranadive (print) Taney Ranadive Date submitted: 08/27/18  
Received by: (sign) Wayne Wang (print) WAYNE WANG Date and time received: 08/27/18 7:23PM  
Page 1 of 1 QLAB\_C-O-C\_V4.01





# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

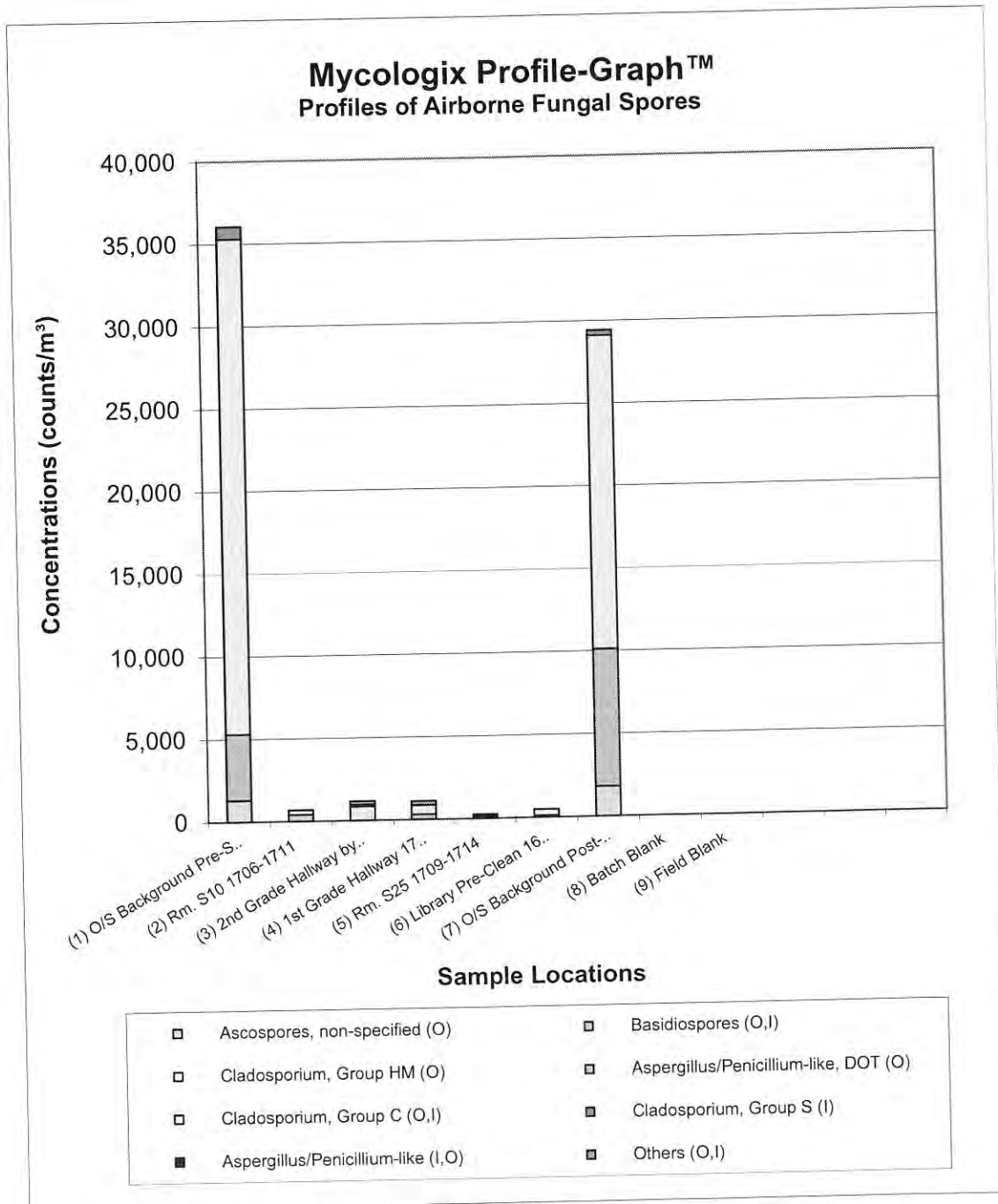
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 (South Wing)  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-19  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





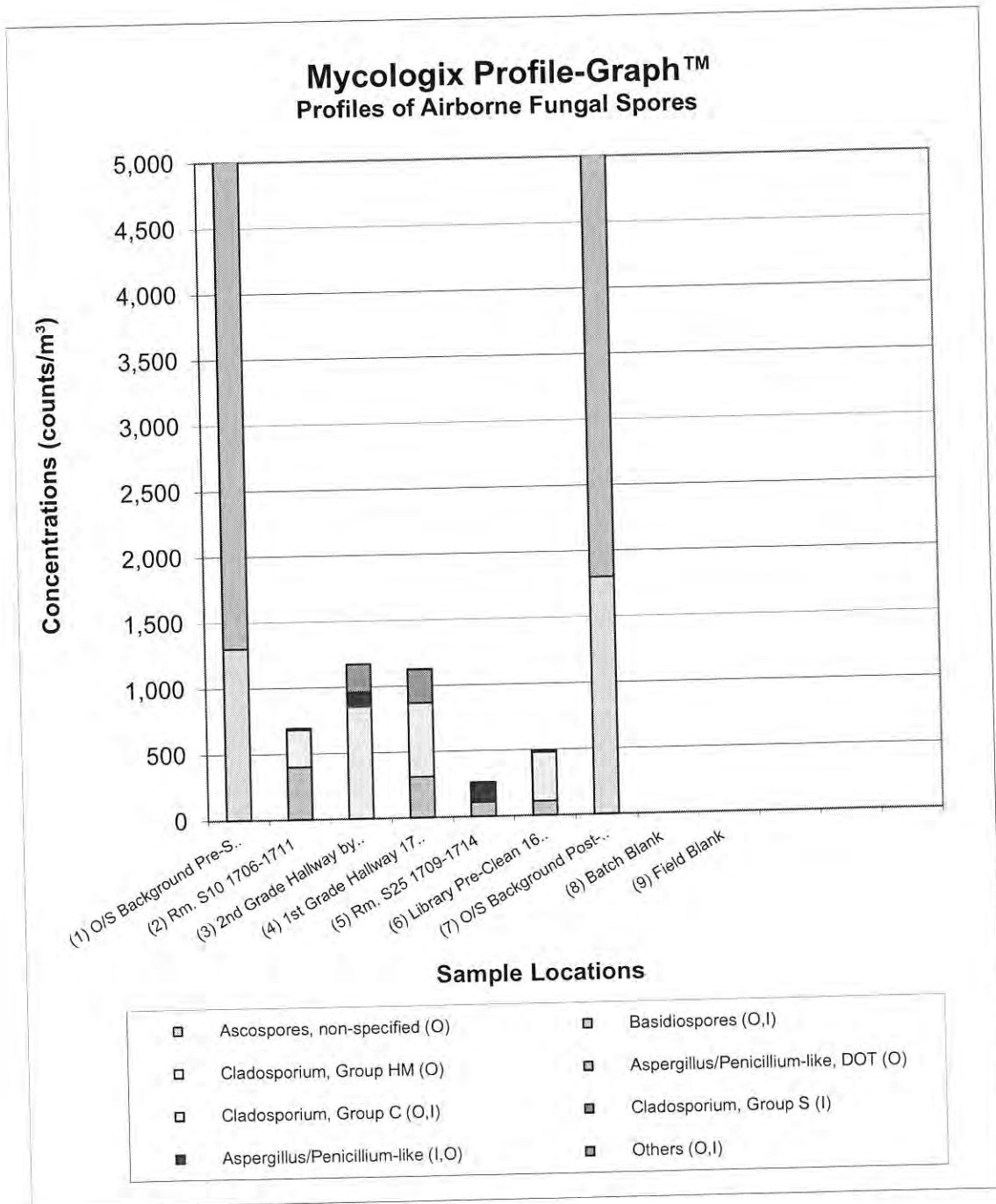
# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 (South Wing)  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-19  
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**Date Reported:** 8/27/2018

Please see original data for complete interpretation.





# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
 info@qlabusa.com www.QLABusa.com  
 AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 (South Wing)  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-19  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180827-19(1)			ME180827-19(2)			ME180827-19(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	O/S Background Pre-Sample 1653-1703			Rm. S10 1706-1711			2nd Grade Hallway by Bathroom 1707-1712		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	36,000 cts/m³			690 cts/m³			1,200 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	196	1,300	4						
Basidiospores (O,I)	604	4,000	11	30	400	58			
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	4,560	30,000	83	21	280	40	64	850	72
Cladosporium, Group S (I)							8	110	9
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™									0% - 0% - 100%
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	3	20	<1	1	13	2			
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	25	170	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)							1	13	1
Myxomycetes/Smuts/Periconia (O,I)	6	40	<1						
Nigrospora (O)							9	120	10
Pithomyces (O)	74	490	1						
Rusts (O)									
Unknown (O,I)	6	40	<1				6	80	7
<b>Skin Cells Rating</b>	None			Trace			Low		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 (South Wing)  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-19  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Lab Sample No.	ME180827-19(4)			ME180827-19(5)			ME180827-19(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	1st Grade Hallway 1708-1713			Rm. S25 1709-1714			Library Pre-Clean 1653-1658		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	1,100 cts/m³			260 cts/m³			490 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	23	310	28	8	110	42	8	110	22
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	42	560	50				28	370	75
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				11	150	58			
Cluster(s)				0% - 0% - 100%					
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)							1	13	3
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	5	67	6						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	2	27	2						
Nigrospora (O)									
Pithomyces (O)	8	110	10						
Rusts (O)									
Unknown (O,I)	4	53	5						
<b>Skin Cells Rating</b>	Low			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 (South Wing)  
**Date Sampled:** 8/27/2018

**QLab Job No.:** ME180827-19  
**Date Received:** 8/27/2018  
**Date Analyzed:** 8/27/2018  
**Date Reported:** 8/27/2018

Lab Sample No.	ME180827-19(7)			ME180827-19(8)			ME180827-19(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	O/S Background Post-Sample 1715-1725			Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			1 smp			1 smp		
Total Concentration (counts/m³)**	29,000 cts/m³			< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)	272	1,800	6						
Basidiospores (O,I)	1,238	8,300	28						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	2,790	19,000	65						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)	5	33	<1						
Alternaria (O,I)	2	13	<1						
Cercospora (O)									
Curvularia (O,I)	8	53	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	5	33	<1						
Nigrospora (O)									
Pithomyces (O)	23	150	<1						
Rusts (O)									
Unknown (O,I)	4	27	<1						
<b>Skin Cells Rating</b>	Trace			None			None		
<b>Debris Rating</b>	1 (≤ 5%)			0 (None detected)			0 (None detected)		
<b>Note</b>				No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





RUSH!

Chain of Custody

RUSH!

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256 Bridge Street, Metuchen, NJ 08840, USA

Lab Job No.: <del>ME180828-17</del> (lab use only) <del>ME1828-17</del> (1) 8/28/18	Telephone No.:	Company Contact:
Company Name: <b>QUEST</b>	Please select: Fax Report ( ) or Email Report (✓)	Project ID: <b>Q18-1941</b>
Company Address: <b>1376 Route 9 Wappingers Falls, NY 12590</b>	Fax No.:	Date/Time sampled: <b>08 '28 '18</b>
Email address: <b>keek@qualityenv.com ljohnson@qualityenv.com + ronald@qualityenv.com</b>		P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Environmental <sup>1443</sup> outside <sup>1453</sup> Room 5-B	FD-01 HP			3hr	Air-o-cell	150L	25740516
1941-02	Main Entry Hallway <sup>1455</sup> <sup>1500</sup>	FD-01 HP			3hr	Air-o-cell	75L	25740557
1941-03	Main office vestibule <sup>1501</sup>	FD-01 HP			3hr	Air-o-cell	75L	25740530
1941-04	Environmental <sup>1502</sup> outside Room 5-B Post	FD-01 HP			3hr	Air-o-cell	150L	25740564
1941-05	Field blank	FD-01 HP				Air o cell		25741135
1941-06	Batch blank	FD-01 HP				Air o cell		

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP. Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) [Signature] (print) Zach Timpano / Luis Jimenez Date submitted: 8/28/18

Received by: (sign) [Signature] (print) WAYNE WANG Date and time received: 8/28/18 6:30PM

Page 1 of 1



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

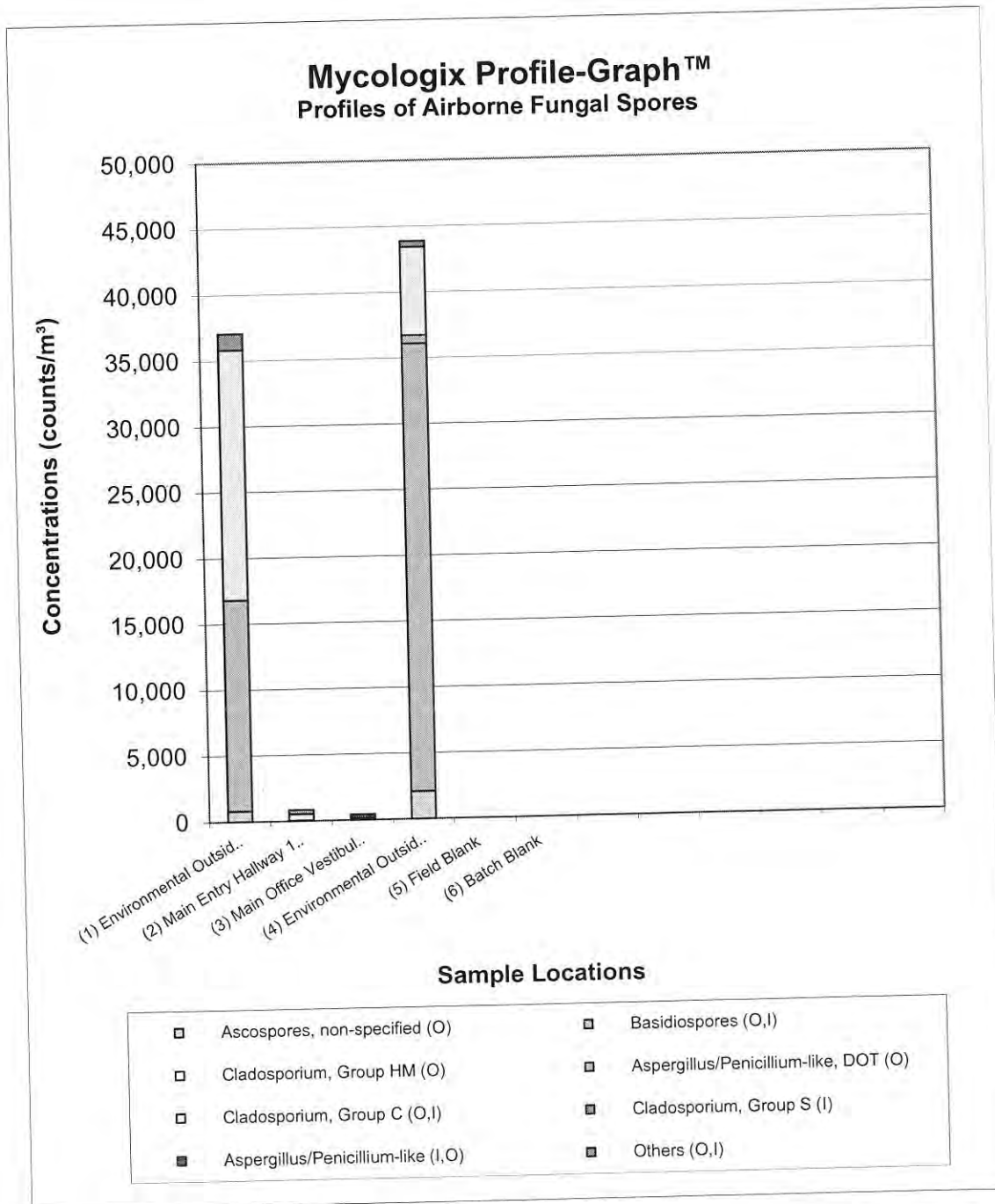
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/28/2018

**QLab Job No.:** ME180828-17  
**Date Received:** 8/28/2018  
**Date Analyzed:** 8/28/2018  
**Date Reported:** 8/28/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





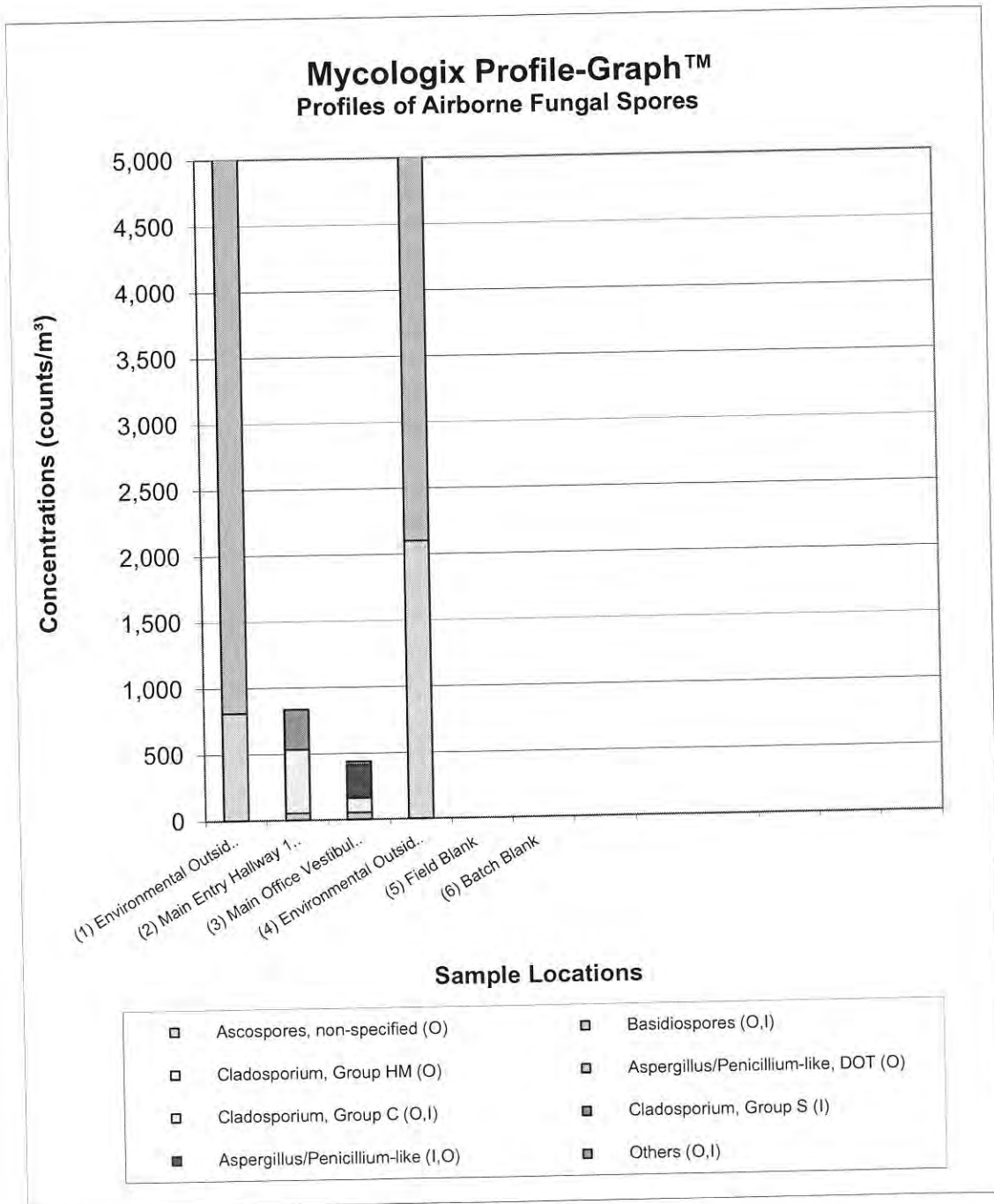
# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/28/2018

**QLab Job No.:** ME180828-17  
**Date Received:** 8/28/2018  
**Date Analyzed:** 8/28/2018  
**Date Reported:** 8/28/2018

Please see original data for complete interpretation.





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**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180828-17(1)			ME180828-17(2)			ME180828-17(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Environmental Outside Room S-8 1443 1453			Main Entry Hallway 1455 1500			Main Office Vestibule 1456 1501		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	37,000 cts/m³			840 cts/m³			440 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	121	810	2						
Basidiospores (O,I)	2,461	16,000	43	4	53	6	4	53	12
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	2,805	19,000	51	36	480	57	8	110	25
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™							19	250	57
Cluster(s)									0% - 58% - 42%
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	5	33	<1						
Alternaria (O,I)	2	13	<1						
Cercospora (O)	3	20	<1						
Curvularia (O,I)	95	630	2	3	40	5	1	13	3
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	4	27	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)				1	13	2			
Nigrospora (O)									
Pithomyces (O)	74	490	1	13	170	20			
Rusts (O)									
Unknown (O,I)	6	40	<1	6	80	10	1	13	3
<b>Skin Cells Rating</b>	None			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





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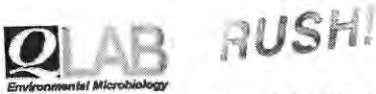
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**Client:** QuES&T  
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**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/28/2018

**QLab Job No.:** ME180828-17  
**Date Received:** 8/28/2018  
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Lab Sample No.	ME180828-17(4)			ME180828-17(5)			ME180828-17(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Environmental Outside Rm S-8 Post 1502			Field Blank			Batch Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			1 smp			1 smp		
Total Concentration (counts/m³)**	44,000 cts/m³			< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)	317	2,100	5						
Basidiospores (O,I)	5,074	34,000	78						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O)	92	610	1						
#Cluster-Chain-Loose Spore Profile™	100% - 0% - 0%								
Cladosporium, Group C (O,I)	1,012	6,700	15						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O)									
## Cluster-Chain-Loose Spore Profile™									
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)	2	13	<1						
Alternaria (O,I)	3	20	<1						
Cercospora (O)	3	20	<1						
Curvularia (O,I)	22	150	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	4	27	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)									
Nigrospora (O)	2	13	<1						
Pithomyces (O)	26	170	<1						
Rusts (O)	3	20	<1						
Unknown (O,I)	4	27	<1						
<b>Skin Cells Rating</b>	None			None			None		
<b>Debris Rating</b>	2 (6 - 25%)			0 (None detected)			0 (None detected)		
<b>Note</b>				No fungal structure observed			No fungal structure observed		

\*: cts/smp; counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



256 Bridge Street, Metuchen, NJ 08840, USA

### Chain of Custody

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: <small>(lab use only)</small> ME 0808 28-18	Telephone No.:	Company Contact:
Company Name: QoES&T	Please select: Fax Report ( ) or Email Report (✓)	Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY, 12590	Fax No.:	Date/Time sampled: 08'28'18
Email address: keck@qualityenv.com jschansen@qualityenv.com *ranadive@qualityenv.com		P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Environmental outside N-15 Prec 1522	FD-01 HP			3hr	Airocell	150L	25740495
1941-02	inside Classroom N-2V 1533	FD-01 HP			3hr	Airocell	75L	25740502
1941-03	North wing Hallway outside N-1531	FD-01 HP			3hr	Airocell	75L	25740537
1941-04	inside classroom N-11 1540	FD-01 HP			3hr	Airocell	75L	25740549
1941-05	inside classroom N-11 1538	FD-01 HP			3hr	Airocell	75L	25740543
1941-06	inside Cafeteria 1543	FD-01 HP			3hr	Airocell	75L	25740529
1941-07	Environmental outside N-15 Post 1545	FD-01 HP			3hr	Airocell	100L	25740577

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.

**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) (print) Zach Timpano / Luis Jimenez Date submitted: 8/28/18  
Received by: (sign) (print) WAYNE WANG Date and time received: 8/28/18 6:30PM



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AIHA EMPAT Lab ID: 178794

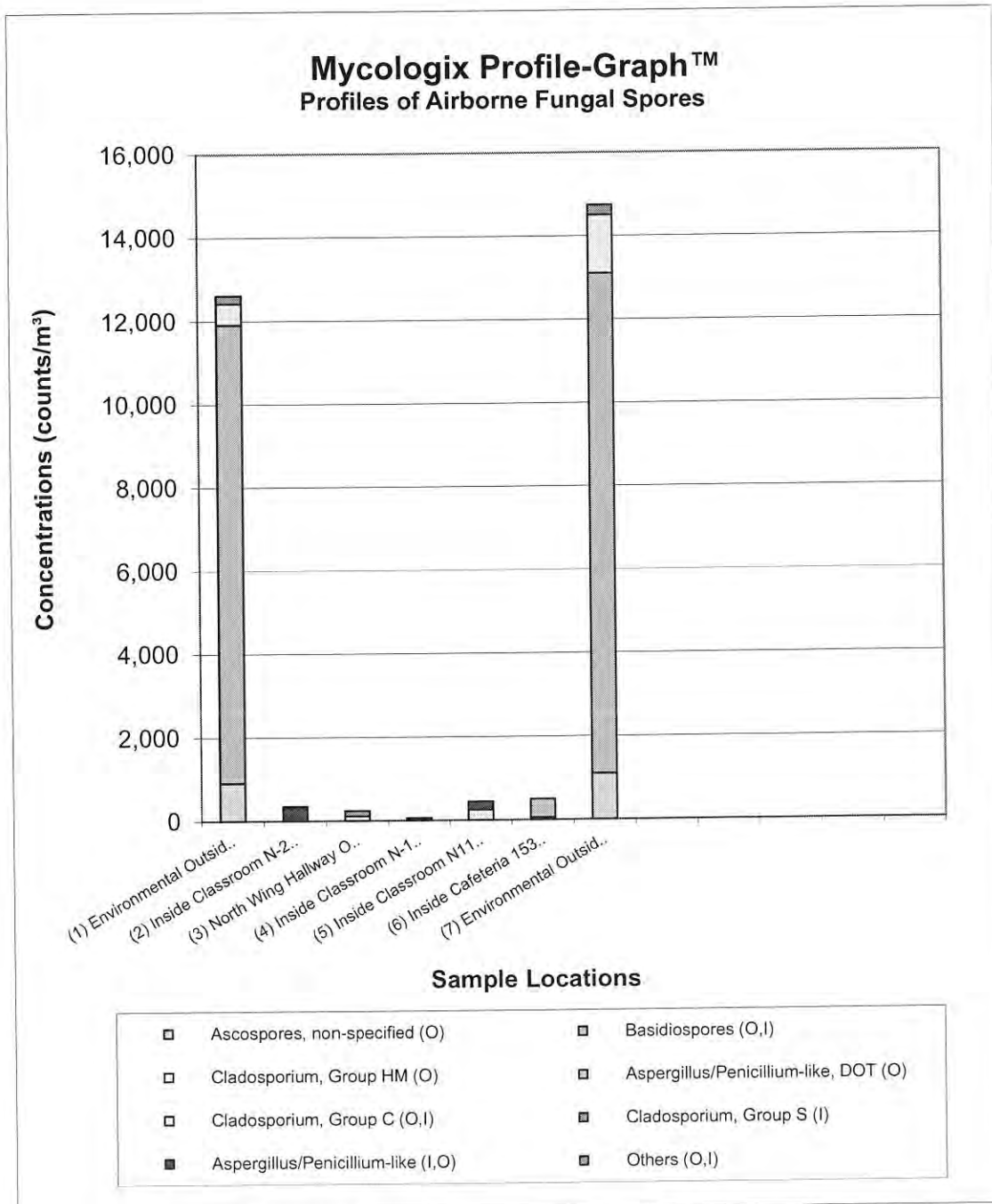
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Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
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**Date Sampled:** 8/28/2018

**QLab Job No.:** ME180828-18  
**Date Received:** 8/28/2018  
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Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





# AccuScience™ Analysis Report

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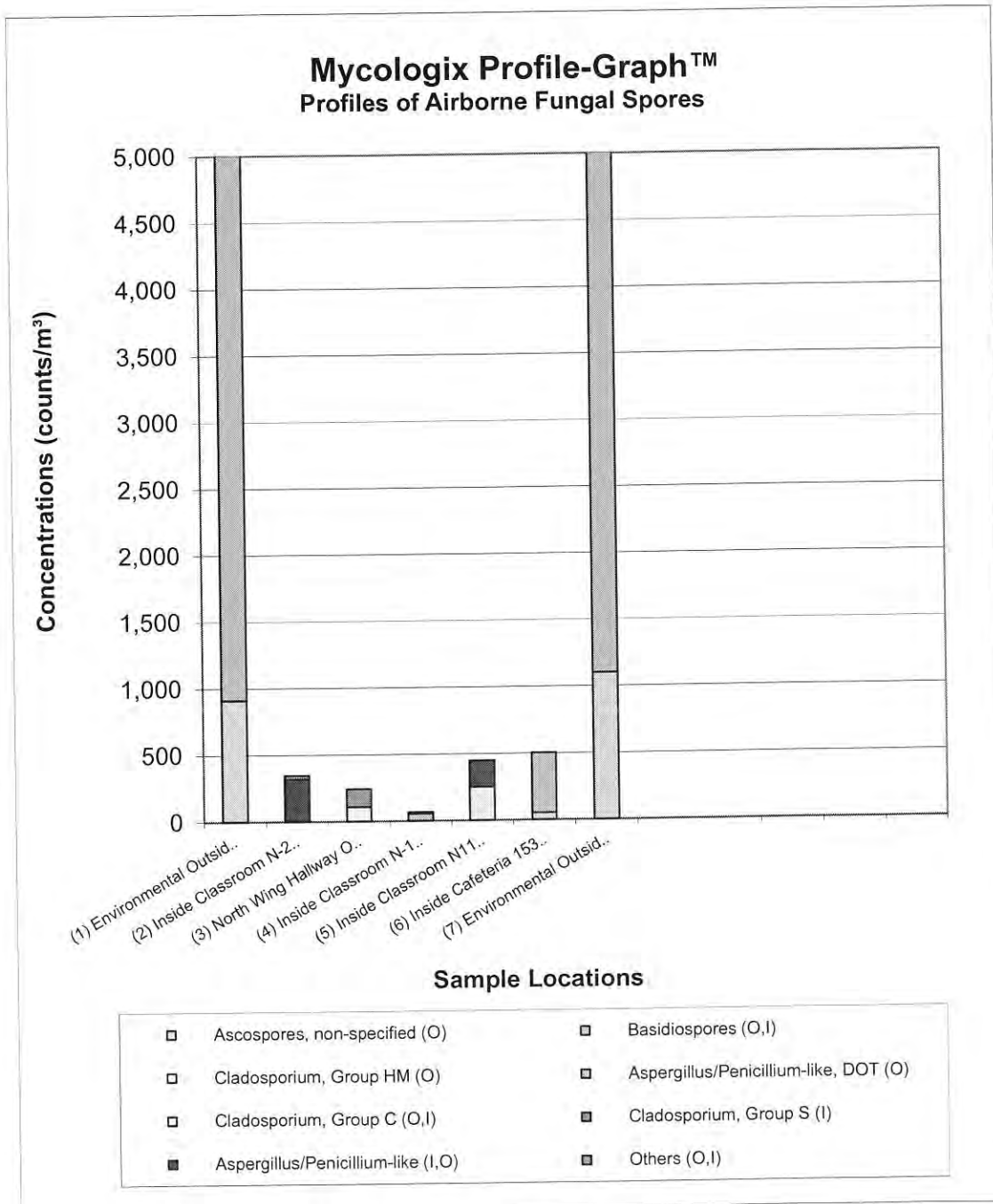
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Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180828-18(1)			ME180828-18(2)			ME180828-18(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Environmental Outside N-15 Pre 1522 1532			Inside Classroom N-21 1533 1538			North Wing Hallway Outside N17 1534 1539		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	13,000 cts/m³			350 cts/m³			240 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	136	910	7						
Basidiospores (O,I)	1,578	11,000	87						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	76	510	4				8	110	45
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				24	320	92			
Cluster(s)				67% - 0% - 33%					
				1 cluster(s) of 16 spores					
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	4	27	<1	2	27	8			
Alternaria (O,I)									
Cercospora (O)	6	40	<1						
Curvularia (O,I)	1	7	<1				2	27	11
Drechslera/Bipolaris-like (O)									
Epicoccum (O)							1	13	5
Fusarium (O,I)	1	7	<1						
Myxomycetes/Smuts/Periconia (O,I)	4	27	<1						
Nigrospora (O)	1	7	<1						
Pithomyces (O)	6	40	<1				6	80	33
Rusts (O)	3	20	<1						
Unknown (O,I)	3	20	<1				1	13	5
<b>Skin Cells Rating</b>	None			None			Low		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			2 (6 - 25%)		
<b>Note</b>									

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Lab Sample No.	ME180828-18(4)			ME180828-18(5)			ME180828-18(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Inside Classroom N-121 1535 1540			Inside Classroom N11 1536 1541			Inside Cafeteria 1538 1543		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	66 cts/m³			450 cts/m³			500 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)							4	53	11
Basidiospores (O,I)	4	53	80				34	450	89
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)				19	250	56			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				15	200	44			
Cluster(s)						0% - 0% - 100%			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	1	13	20						
Nigrospora (O)									
Pithomyces (O)									
Rusts (O)									
Unknown (O,I)									
<b>Skin Cells Rating</b>	Trace			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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**Date Received:** 8/28/2018  
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Lab Sample No.	ME180828-18(7)		
Sample ID	1941-07		
Sample Location	Environmental Outside N-15 Post 1545		
Sample Type (Device)	Air (Air-O-Cell)		
Air Volume	150 L		
Total Concentration (counts/m³)**	15,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	159	1,100	7
Basidiospores (O,I)	1,835	12,000	81
Cladosporium, Group HM (O)			
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™			
Cladosporium, Group C (O,I)	211	1,400	9
Cladosporium, Group S (I)			
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)			
Chaetomium (I)			
Ulocladium (I)			
Memmoniella (I)			
Trichoderma (I)			
Scopulariopsis (I)			
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	3	20	<1
Alternaria (O,I)	3	20	<1
Cercospora (O)	11	73	<1
Curvularia (O,I)			
Drechslera/Bipolaris-like (O)	1	7	<1
Epicoccum (O)	1	7	<1
Fusarium (O,I)			
Myxomycetes/Smuts/Periconia (O,I)	3	20	<1
Nigrospora (O)	2	13	<1
Pithomyces (O)	8	53	<1
Rusts (O)			
Unknown (O,I)	4	27	<1
<b>Skin Cells Rating</b>	None		
<b>Debris Rating</b>	2 (6 - 25%)		
<b>Note</b>			

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





RUSH!

Chain of Custody

RUSH!

256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)
Tel: 856-489-0011 www.QLabUSA.com

Form with fields: Lab Job No.: ME180829-15, Telephone No.: 845-298-6031, Company Contact: Lewis Johnson, Company Name: QUEST, Project ID: Q18-1941, Company Address: 1376 Route 9, Wappingers Falls, NY 12590, Fax No., Date/Time sampled: 8/29/18, Email address: kock@qualityenv.com

Table with columns: Sample ID, Sample Location, Analysis Code, Turnaround Time (Std, Day, Hr), Sample Type, Volume (L) or Area (in^2), Note. Contains 8 rows of sample data.

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP.

Fungi Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) [Signature] (print) Lewis Johnson Date submitted: 8/29/18
Received by: (sign) [Signature] (print) Whitney Wang Date and time received: 8/29/18 6:51 PM
Page 1 of 1 QLAB\_C-O-C\_V4.01





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-15  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180829-15(1)			ME180829-15(2)			ME180829-15(3)		
Sample ID	1941-09			1941-10			1941-11		
Sample Location	Environmental O/S E17 Pre 1555-1605			Classroom # E12 1606-1611			Classroom # E13 1607-1612		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	11,000 cts/m³			3,900 cts/m³			6,600 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	166	1,100	10						
Basidiospores (O,I)	1,231	8,200	72				38	510	8
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	287	1,900	17				43	570	9
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				296	3,900	100	412	5,500	83
Cluster(s)				0% - 30% - 70%			0% - 24% - 76%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)				1	13	<1			
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	2	13	<1						
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	8	53	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	2	13	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	3	20	<1						
Nigrospora (O)	1	7	<1						
Pithomyces (O)	6	40	<1				1	13	<1
Rusts (O)	2	13	<1						
Unknown (O,I)	12	80	<1						
<b>Skin Cells Rating</b>	None			Trace			Low		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-15  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Lab Sample No.	ME180829-15(4)			ME180829-15(5)			ME180829-15(6)		
Sample ID	1941-12			1941-13			1941-14		
Sample Location	Classroom # E16 1608-1613			East Wing Hallway 1609-1614			Environmental O/S E17 Post 1615-1625		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			150 L		
Total Concentration (counts/m³)**	17,000 cts/m³			2,500 cts/m³			15,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 67; LQL = 1300 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)							45	300	2
Basidiospores (O,I)				11	150	6	1,314	8,800	58
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)							891	5,900	39
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	1,259	17,000	100	171	2,300	93			
Cluster(s)	Too numerous to categorize			0% - 5% - 95%					
	Too numerous to categorize								
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)	1	13	<1						
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)				1	13	<1	5	33	<1
Alternaria (O,I)							2	13	<1
Cercospora (O)							5	33	<1
Curvularia (O,I)							4	27	<1
Drechslera/Bipolaris-like (O)							1	7	<1
Epicoccum (O)							1	7	<1
Fusarium (O,I)							3	20	<1
Myxomycetes/Smuts/Periconia (O,I)	1	13	<1				1	7	<1
Nigrospora (O)							9	60	<1
Pithomyces (O)									
Rusts (O)							5	33	<1
Unknown (O,I)									
<b>Skin Cells Rating</b>	Medium			Low			None		
<b>Debris Rating</b>	3 (26 - 75%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-15  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Lab Sample No.	ME180829-15(7)			ME180829-15(8)		
Sample ID	1941-15			1941-16		
Sample Location	Field Blank			Batch Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m³)**	< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™						
Cluster(s)						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memnoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected) No fungal structure observed			0 (None detected) No fungal structure observed		
<b>Note</b>						

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)
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Lab Job No.: ME18082916
Telephone No.: 845-298-6031
Company Contact: Louis Johnson III
Company Name: QUEST
Please select: Fax Report ( ) or Email Report (X)
Project ID: Q18-1941
Company Address: 1376 Route 9, Wappingers Falls, NY 12591
Fax No.:
Date/Time sampled: 8/29/18
Email address: kecc@qualityenv.com, ljohnson@qualityenv.com, ljohnson@quest.com

Table with 7 columns: Sample ID, Sample Location, Analysis Code, Turnaround Time (Std, Day, Hr), Sample Type, Volume (L) or Area (in^2), Note. Rows include samples 1941-01 through 1941-08 with locations like Environmental O/S NIS, Classroom N21, Environmental O/S NIS post, Environmental O/S SR post, Main office Vestibule/Hallway, Environmental O/S SR post, Field Blank, and Batch Blank.

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP.

Fungal Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) [Signature] (print) Louis N. Johnson III Date submitted: 8/29/18
Received by: (sign) [Signature] (print) WAYNE WANG Date and time received: 8/29/18 6:51 PM





# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
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AIHA EMPAT Lab ID: 178794

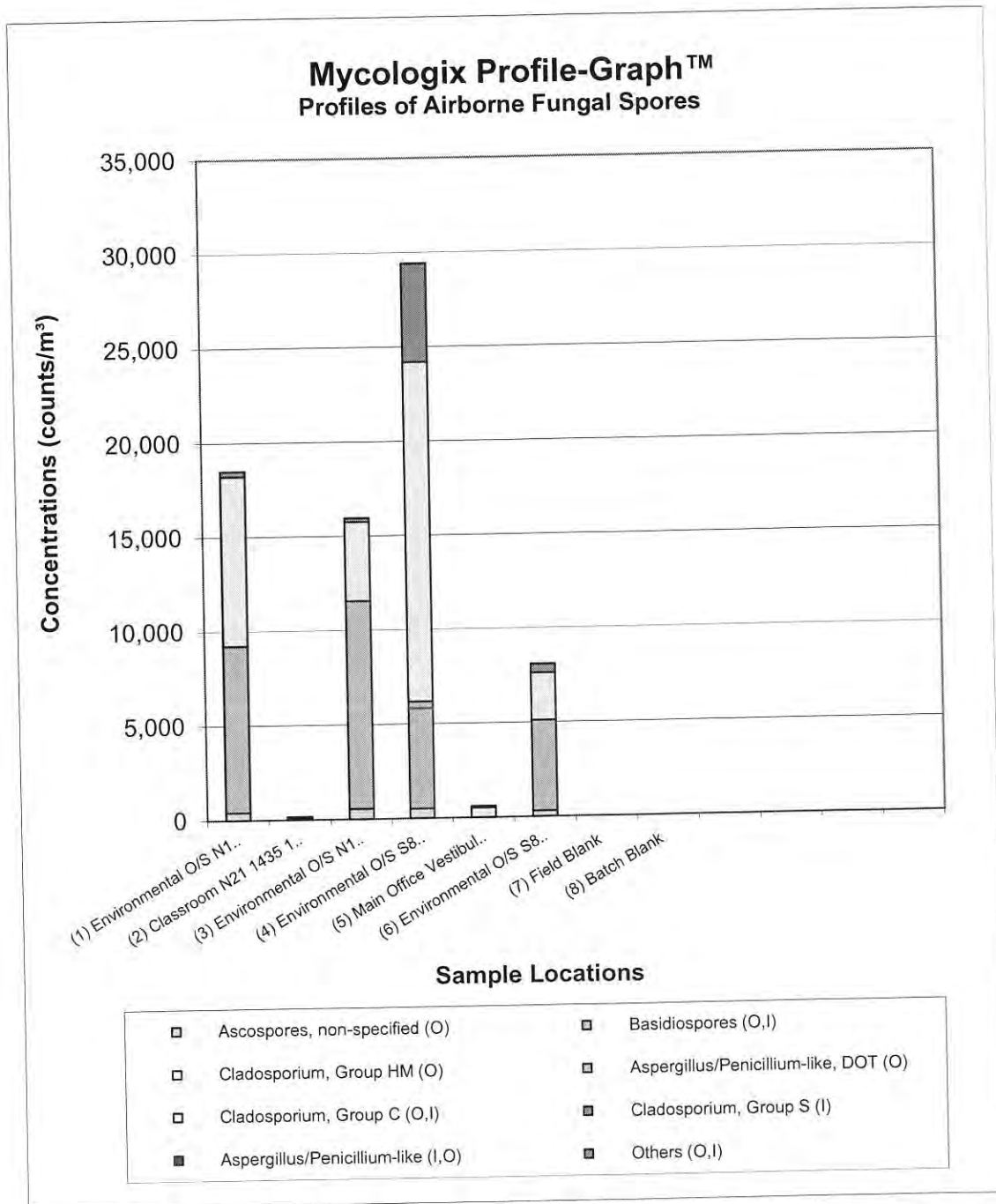
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-16  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





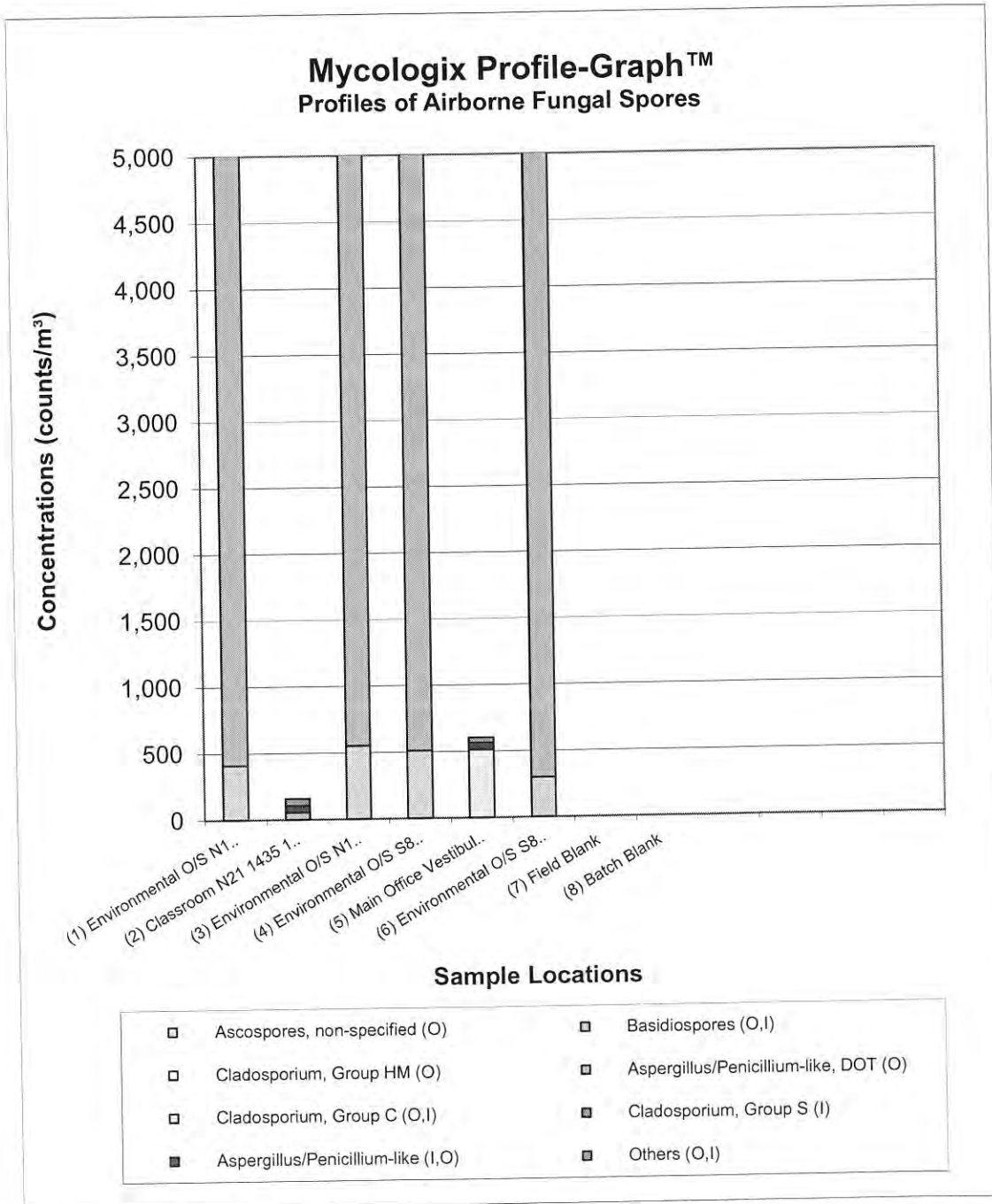
# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-16  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Please see original data for complete interpretation.





AccuScience™  
Analysis Report

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**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-16  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180829-16(1)			ME180829-16(2)			ME180829-16(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Environmental O/S N15 Pre 1423 1433			Classroom N21 1435 1440			Environmental O/S N15 Post 1442 1452		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			150 L		
Total Concentration (counts/m³)**	18,000 cts/m³			160 cts/m³			16,000 cts/m³		
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 67; LQL = 1300 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	61	410	2				83	550	3
Basidiospores (O,I)	1,323	8,800	48	4	53	33	1,616	11,000	69
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	1,348	9,000	49				627	4,200	26
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				4	53	33			
Cluster(s)						0% - 0% - 100%			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	2	13	<1				2	13	<1
Alternaria (O,I)	1	7	<1				1	7	<1
Cercospora (O)	2	13	<1				2	13	<1
Curvularia (O,I)	5	33	<1				3	20	<1
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	1	7	<1				3	20	<1
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	3	20	<1	3	40	25	9	60	<1
Nigrospora (O)	2	13	<1						
Pithomyces (O)	19	130	<1	1	13	8	7	47	<1
Rusts (O)	5	33	<1				1	7	<1
Unknown (O,I)	2	13	<1				2	13	<1
<b>Skin Cells Rating</b>	None			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			3 (26 - 75%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-16  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Lab Sample No.	ME180829-16(4)			ME180829-16(5)			ME180829-16(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Environmental O/S S8 Pre 1450-1500			Main Office Vestibule/Hallway 1502-1507			Environmental O/S S8 Post 1508-1518		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			150 L		
Total Concentration (counts/m³)**	29,000 cts/m³			600 cts/m³			8,100 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 33; LQL = 670 cts/m³		
Ascospores, non-specified (O)	76	510	2				45	300	4
Basidiospores (O,I)	800	5,300	18				719	4,800	59
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O)	56	370	1						
#Cluster-Chain-Loose Spore Profile™	100% - 0% - 0%								
Cladosporium, Group C (O,I)	2,727	18,000	61	38	510	85	382	2,500	31
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O)				4	53	9			
### Cluster-Chain-Loose Spore Profile™				0% - 0% - 100%					
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)	8	53	<1				14	93	1
Alternaria (O,I)	2	13	<1				2	13	<1
Cercospora (O)	1	7	<1				5	33	<1
Curvularia (O,I)	196	1,300	4				16	110	1
Drechslera/Bipolaris-like (O)	1	7	<1				2	13	<1
Epicoccum (O)	5	33	<1				3	20	<1
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	2	13	<1						
Nigrospora (O)	2	13	<1				1	7	<1
Pithomyces (O)	544	3,600	12	3	40	7	22	150	2
Rusts (O)									
Unknown (O,I)	29	190	<1				5	33	<1
<b>Skin Cells Rating</b>	None			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Johnson, Louis, III  
**Project ID:** Q18-1941  
**Date Sampled:** 8/29/2018

**QLab Job No.:** ME180829-16  
**Date Received:** 8/29/2018  
**Date Analyzed:** 8/29/2018  
**Date Reported:** 8/29/2018

Lab Sample No.	ME180829-16(7)			ME180829-16(8)		
Sample ID	1941-07			1941-08		
Sample Location	Field Blank			Batch Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m <sup>3</sup> )**	< DL cts/smp			< DL cts/smp		
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memmoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected)			0 (None detected)		
<b>Note</b>	No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



256 Bridge Street, Metuchen, NJ 08840, USA

**EXPEDITE**

Chain of Custody

**EXPEDITE**

Tel: 856-489-0011 Fax: 888-QLab-Wei (888-752-2934) www.QLabUSA.com

Lab Job No.: (lab use only) <b>ME180830-17</b>	Telephone No.: <b>845-559-8537</b>	Company Contact: <b>Taney Ranaidive</b>
Company Name: <b>QuES&amp;T</b>	Please select: Fax Report ( ) or Email Report ( <input checked="" type="checkbox"/> )	Project ID: <b>Q18-1941</b>
Company Address: <b>1376 Route 9, Wappingers Falls, NY 12590</b>	Fax No.:	Date/Time sampled: <b>08/30/18 15:40</b>
	Email address: <b>tranaidive@qualityenv.com</b>	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Outside Pre-sample	1459-1504 FD-01HP			3 Hr	Air-O-cell	150L	2657 7322
1941-02	Rm. N16	1512-1517					75L	2657 5880
1941-03	Rm. N13	1514-1519					75L	2657 7451
1941-04	Hallway	1516-1521					75L	2657 5863
1941-05	Rm. N12	1517-1522					75L	2657 7329
1941-06	outside Post-sample	1523-1533					150L	2657 7328
1941-07	Batch Blank						-	2657-7319
1941-08	Field Blanks						-	2657-7340

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP.

Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) Taney Ranaidive (print) Taney Ranaidive

Date submitted: 08/30/18

Received by: (sign) Mindy Wang (print) Mindy Wang

Date and time received: 08/30/18 6:16PM





# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™

**Client:** QuES&T  
Wappingers Falls, NY

**Contact:** Ranadive, Tanay

**Project ID:** Q18-1941

**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17

**Date Received:** 8/30/2018

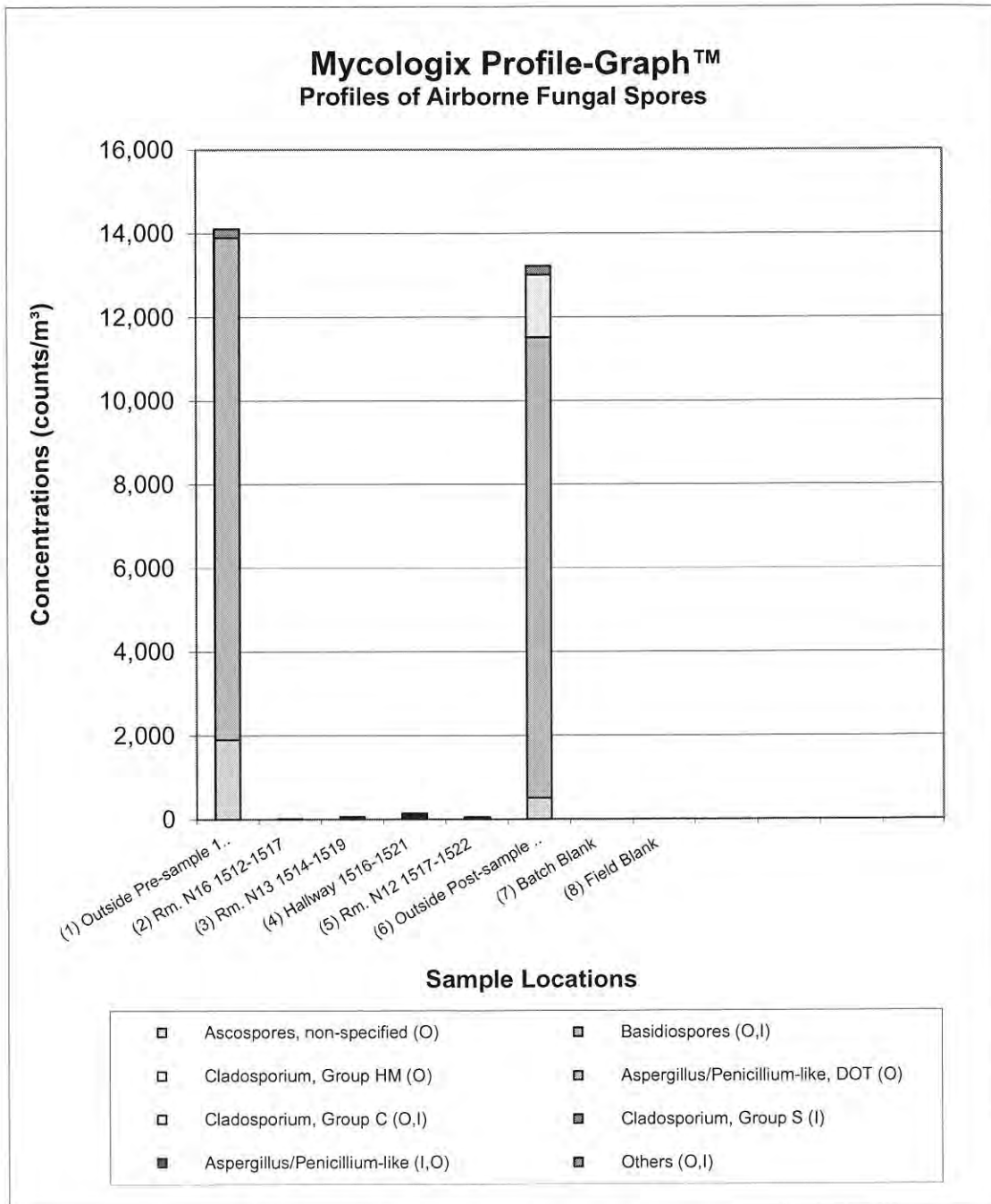
**Date Analyzed:** 8/30/2018

**Date Reported:** 8/30/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180830-17(1)			ME180830-17(2)			ME180830-17(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-sample 1459-1509			Rm. N16 1512-1517			Rm. N13 1514-1519		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	14,000 cts/m³			13 cts/m³			66 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	279	1,900	13				4	53	80
Basidiospores (O,I)	1,767	12,000	85						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™									
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	4	27	<1						
Alternaria (O,I)									
Cercospora (O)	2	13	<1						
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	2	13	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	11	73	<1						
Nigrospora (O)	2	13	<1						
Pithomyces (O)	1	7	<1	1	13	100			
Rusts (O)	2	13	<1						
Unknown (O,I)	8	53	<1				1	13	20
<b>Skin Cells Rating</b>	None			Low			Low		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

Lab Sample No.	ME180830-17(4)			ME180830-17(5)			ME180830-17(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Hallway 1516-1521			Rm. N12 1517-1522			Outside Post-sample 1523-1533		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			150 L		
Total Concentration (counts/m³)**	150 cts/m³			53 cts/m³			13,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)							76	510	4
Basidiospores (O,I)	4	53	36				1,699	11,000	83
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	4	53	36				227	1,500	11
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				4	53	100			
Cluster(s)						0% - 0% - 100%			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)							1	7	<1
Alternaria (O,I)							1	7	<1
Cercospora (O)									
Curvularia (O,I)							3	20	<1
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)							1	7	<1
Myxomycetes/Smuts/Periconia (O,I)	1	13	9				1	7	<1
Nigrospora (O)							2	13	<1
Pithomyces (O)	2	27	18				17	110	<1
Rusts (O)									
Unknown (O,I)							6	40	<1
<b>Skin Cells Rating</b>	Trace			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

Lab Sample No.	ME180830-17(7)			ME180830-17(8)		
Sample ID	1941-07			1941-08		
Sample Location	Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m <sup>3</sup> )**	< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) ### Cluster-Chain-Loose Spore Profile™ Cluster(s)						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memnoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected)			0 (None detected)		
<b>Note</b>	No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



256 Bridge Street, Metuchen, NJ 08840, USA

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Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: (lab use only) <b>ME180830-17</b>	Telephone No.: <b>845-559-8537</b>	Company Contact: <b>Tanay Ranadive</b>
Company Name: <b>QuES&amp;T</b>	Please select: Fax Report ( ) or Email Report (✓)	Project ID: <b>Q18-1941</b>
Company Address: <b>1376 Route 9, Wappingers Falls, NY 12590</b>	Fax No.:	Date/Time sampled: <b>08/30/18 15:40</b>
	Email address: <b>tranaadive@qualityenv.com</b>	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Outside Pre-sample <sup>1459-1509</sup>	FD-01HP			3 Hr	Air-O-cell	150L	2657 7322
1941-02	Rm. N16 <sup>1512-1517</sup>	"			"	"	75L	2657 5880
1941-03	Rm. N13 <sup>1514-1519</sup>	"			"	"	75L	2657 7451
1941-04	Hallway <sup>1516-1521</sup>	"			"	"	75L	2657 5863
1941-05	Rm. N12 <sup>1517-1522</sup>	"			"	"	75L	2657 7329
1941-06	outside Post-Sample <sup>1523-1533</sup>	"			"	"	150L	2657 7328
1941-07	Batch Blank					"	-	2657-7319
1941-08	Field Blanks					"	-	2657-7340

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP.

Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive

Date submitted: 08/30/18

Received by: (sign) Nancy Wang (print) Nancy Wang

Date and time received: 08/30/18 6:16 PM



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™

**Client:** QuES&T  
Wappingers Falls, NY

**Contact:** Ranadive, Tanay

**Project ID:** Q18-1941

**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17

**Date Received:** 8/30/2018

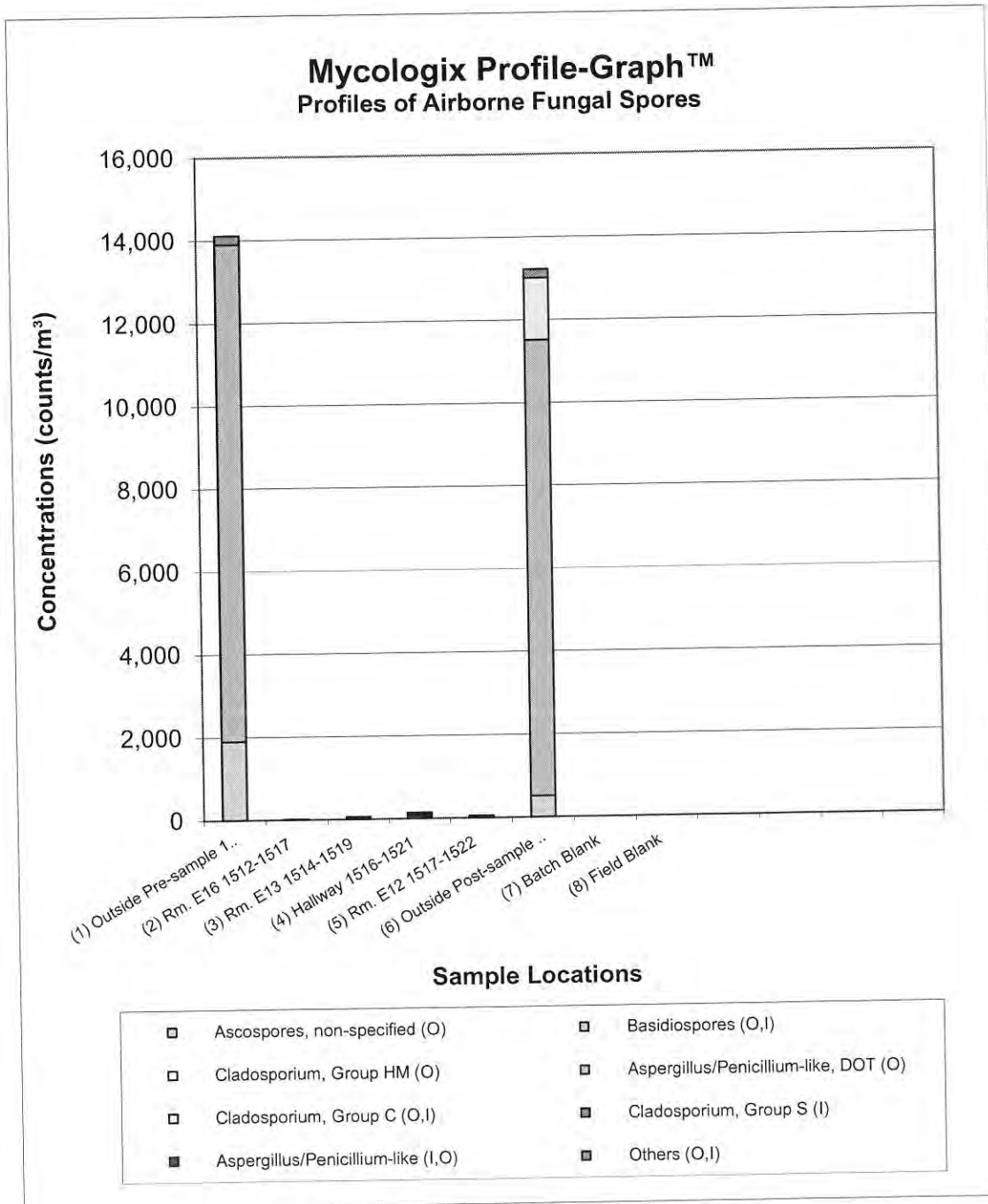
**Date Analyzed:** 8/30/2018

**Date Reported:** 8/30/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180830-17(1)			ME180830-17(2)			ME180830-17(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-sample 1459-1509			Rm. E16 1512-1517			Rm. E13 1514-1519		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	14,000 cts/m³			13 cts/m³			66 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	279	1,900	13						
Basidiospores (O,I)	1,767	12,000	85				4	53	80
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	4	27	<1						
Alternaria (O,I)									
Cercospora (O)	2	13	<1						
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	2	13	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	11	73	<1						
Nigrospora (O)	2	13	<1						
Pithomyces (O)	1	7	<1	1	13	100			
Rusts (O)	2	13	<1						
Unknown (O,I)	8	53	<1				1	13	20
<b>Skin Cells Rating</b>	None			Low			Low		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

Lab Sample No.	ME180830-17(4)			ME180830-17(5)			ME180830-17(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Hallway 1516-1521			Rm. E12 1517-1522			Outside Post-sample 1523-1533		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			150 L		
Total Concentration (counts/m³)**	150 cts/m³			53 cts/m³			13,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)							76	510	4
Basidiospores (O,I)	4	53	36				1,699	11,000	83
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	4	53	36				227	1,500	11
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				4	53	100			
Cluster(s)						0% - 0% - 100%			
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)							1	7	<1
Alternaria (O,I)							1	7	<1
Cercospora (O)									
Curvularia (O,I)							3	20	<1
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)							1	7	<1
Myxomycetes/Smuts/Periconia (O,I)	1	13	9				1	7	<1
Nigrospora (O)							2	13	<1
Pithomyces (O)	2	27	18				17	110	<1
Rusts (O)									
Unknown (O,I)							6	40	<1
<b>Skin Cells Rating</b>	Trace			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 8/30/2018

**QLab Job No.:** ME180830-17  
**Date Received:** 8/30/2018  
**Date Analyzed:** 8/30/2018  
**Date Reported:** 8/30/2018

Lab Sample No.	ME180830-17(7)			ME180830-17(8)			
Sample ID	1941-07			1941-08			
Sample Location	Batch Blank			Field Blank			
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			
Air Volume	1 smp			1 smp			
Total Concentration (counts/m <sup>3</sup> )**	< DL cts/smp			< DL cts/smp			
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%	
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp			
Ascospores, non-specified (O)							
Basidiospores (O,I)							
Cladosporium, Group HM (O)							
Aspergillus/Penicillium-like, DOT (O)							
#Cluster-Chain-Loose Spore Profile™							
Cladosporium, Group C (O,I)							
Cladosporium, Group S (I)							
Aspergillus/Penicillium-like (I,O)							
## Cluster-Chain-Loose Spore Profile™							
Cluster(s)							
<b>2. Indoor Hydrophilic Fungi<sup>‡</sup></b>	DL = 1 cts/smp			DL = 1 cts/smp			
Stachybotrys (I)							
Chaetomium (I)							
Ulocladium (I)							
Memmoniella (I)							
Trichoderma (I)							
Scopulariopsis (I)							
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp			
Hyphal fragment (O,I)							
Alternaria (O,I)							
Cercospora (O)							
Curvularia (O,I)							
Drechslera/Bipolaris-like (O)							
Epicoccum (O)							
Fusarium (O,I)							
Myxomycetes/Smuts/Periconia (O,I)							
Nigrospora (O)							
Pithomyces (O)							
Rusts (O)							
Unknown (O,I)							
<b>Skin Cells Rating</b>	None			None			
<b>Debris Rating</b>	0 (None detected)			0 (None detected)			
<b>Note</b>	No fungal structure observed			No fungal structure observed			

\*: cts/smp; counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





**EXPEDITE**

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**Chain of Custody**

256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: (lab use only) <b>ME180-901-06</b>	Telephone No.: <b>845-559-8537</b>	Company Contact: <b>Tanay Ranadive</b>
Company Name: <b>QUEST</b>	Please select: Fax Report ( ) or Email Report ( <input checked="" type="checkbox"/> )	Project ID: <b>Q18-1941 Library Wing</b>
Company Address: <b>1376 Route 9 Wappingers Falls, NY 12590</b>	Fax No.:	Date/Time sampled: <b>09/01/18 15:30</b>
	Email address: <b>tranadive@qualityson.com</b>	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	outside Pre-Sample 1445-1455	FD-01HP			3hr	Air-o-cell	150 L 2657 7347	2657 5886
1941-02	Kitchen Pantry closet 1457-1502	FD-01HP					75	2657 7330
1941-03	Speech Room Hall 1505-1510						75	2657 7320
1941-04	Cl in Hall 1506-1511						75	2657 7327
1941-05	Mech room side Library 1507-1512						75	2657 7310
1941-06	Library-Center 1508-1513						75	2657-7493
1941-07	Library wood floor 1509-1514						75	2657-7434
1941-08	Cl in Hallway 1516-1521						75	2657 5861
1941-09	outside first sample 1523-1533						150	2657 7338
1941-10	Batch Blank							2657 5703
1941-11	Field Blank							

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.

**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive Date submitted: 09/01/18  
 Received by: (sign) Wei Tang (print) Wei Tang Date and time received: 09/01/18 6:12 PM



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

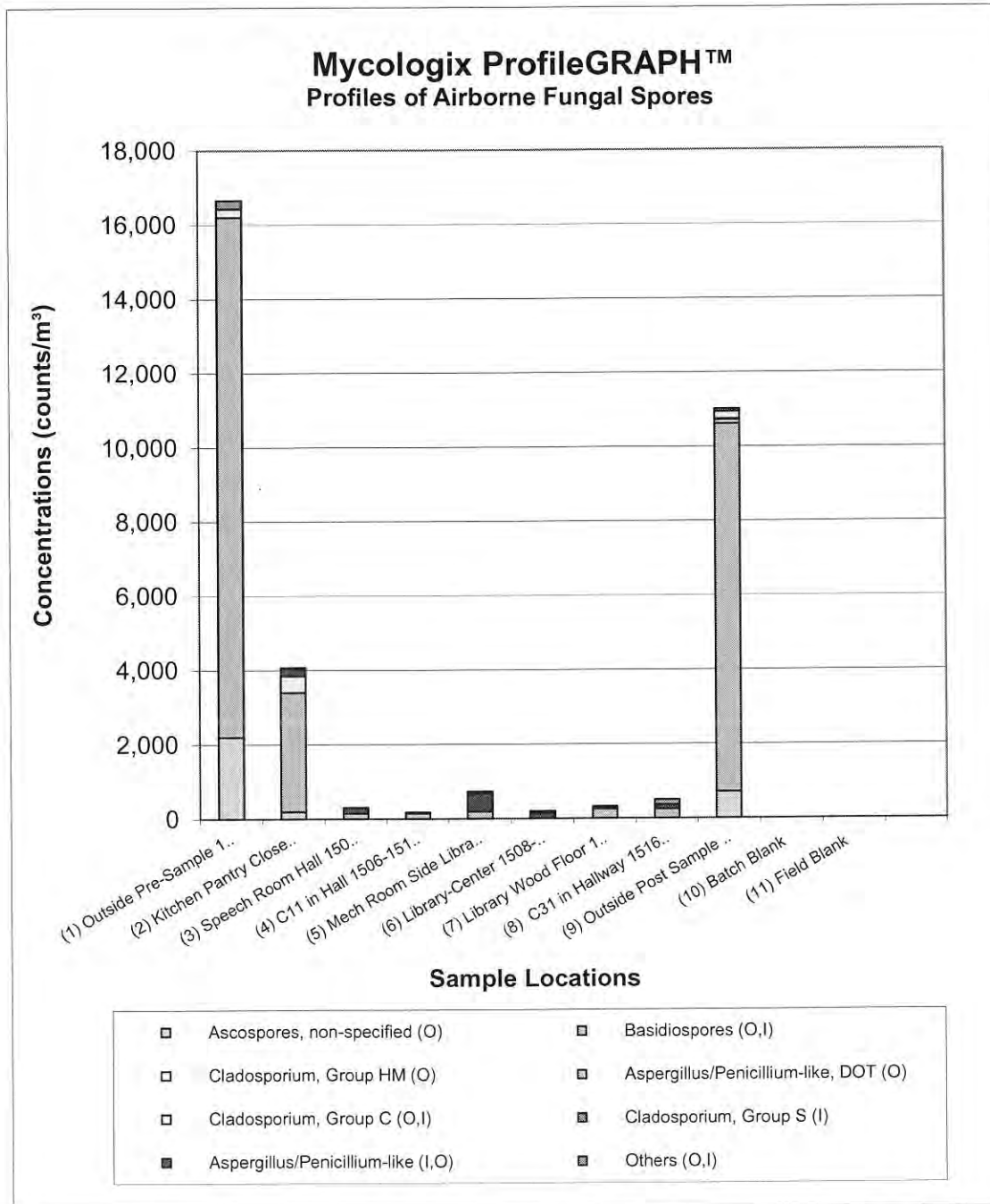
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Library Wing  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-06  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





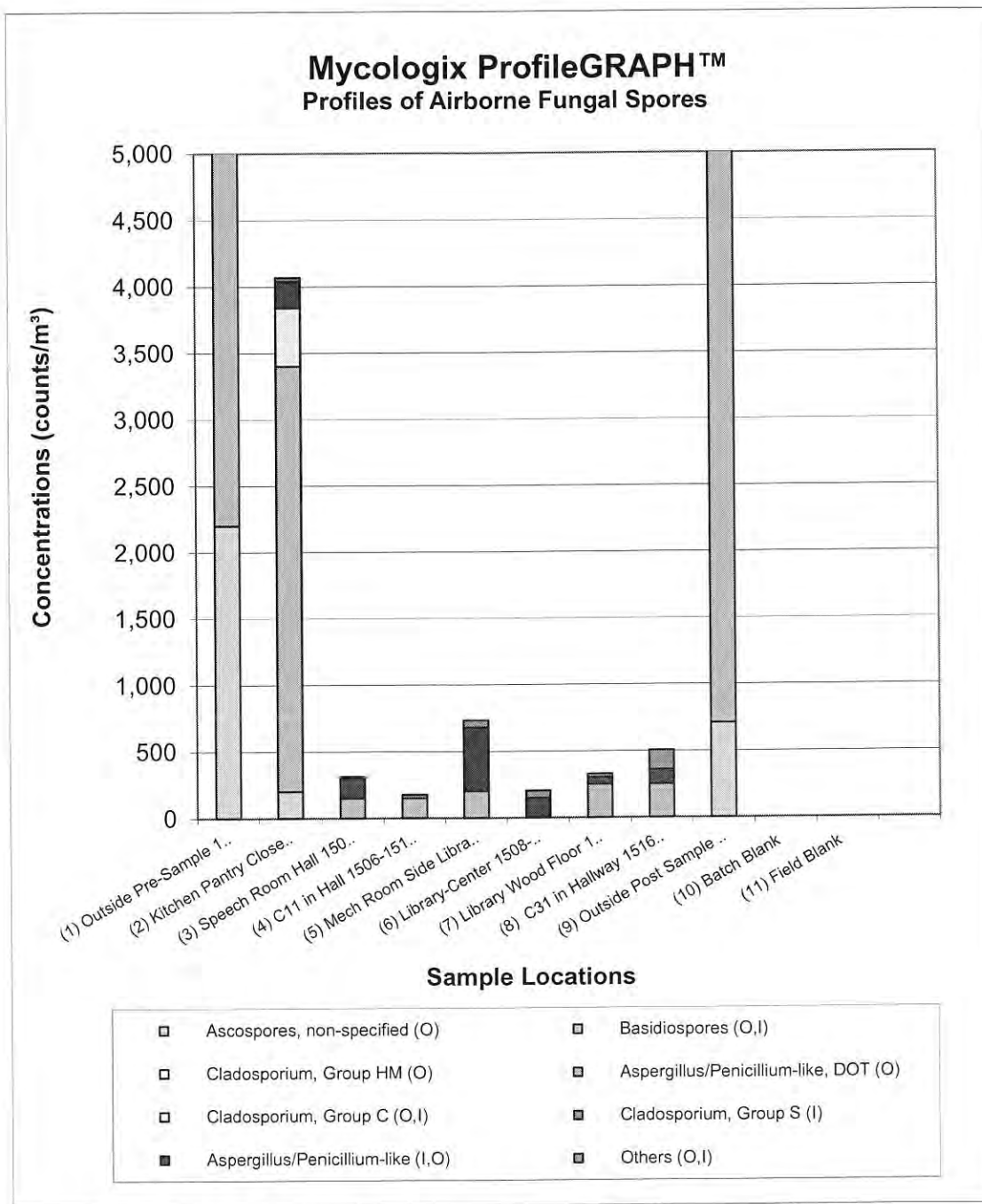
# AccuScience™ Analysis Report

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**Date Sampled:** 9/1/2018

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**Date Reported:** 9/1/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180901-06(1)			ME180901-06(2)			ME180901-06(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-Sample 1445-1455			Kitchen Pantry Closet 1457-1502			Speech Room Hall 1505-1510		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	17,000 cts/m³			4,100 cts/m³			310 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 67; LQL = 1300 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	333	2,200	13	15	200	5			
Basidiospores (O,I)	2,111	14,000	84	238	3,200	79	11	150	48
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	34	230	1	33	440	11			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				15	200	5	11	150	48
Cluster(s)				0% - 0% - 100%			0% - 64% - 36%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	1	7	<1						
Alternaria (O,I)	1	7	<1						
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	2	13	<1				1	13	4
Nigrospora (O)	1	7	<1						
Pithomyces (O)	26	170	1	2	27	<1			
Rusts (O)									
Unknown (O,I)	4	27	<1						
<b>Skin Cells Rating</b>	Trace			Low			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Library Wing  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-06  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-06(4)			ME180901-06(5)			ME180901-06(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	C11 in Hall 1506-1511			Mech Room Side Library 1507-1512			Library-Center 1508-1513		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	180 cts/m³			730 cts/m³			200 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)	11	150	85	15	200	27			
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™				36	480	65	11	150	74
Cluster(s)				58% - 0% - 42%			0% - 0% - 100%		
				1 cluster(s) of 21 spores					
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)				1	13	2	2	27	13
Nigrospora (O)									
Pithomyces (O)	1	13	7	2	27	4	2	27	13
Rusts (O)									
Unknown (O,I)	1	13	7	1	13	2			
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	1 (≤ 5%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Library Wing  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-06  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-06(7)			ME180901-06(8)			ME180901-06(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	Library Wood Floor 1509-1514			C31 in Hallway 1516-1521			Outside Post Sample 1523-1533		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			150 L		
Total Concentration (counts/m³)**	330 cts/m³			510 cts/m³			11,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)							106	710	6
Basidiospores (O,I)	19	250	76	19	250	49	1,487	9,900	90
Cladosporium, Group HM (O)							15	100	<1
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™							4	27	<1
Cladosporium, Group C (O,I)							30	200	2
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	4	53	16	8	110	22			
Cluster(s)	0% - 0% - 100%			0% - 0% - 100%			0% - 100% - 0%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 7; LQL = 130 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)							1	7	<1
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	2	27	8	1	13	3	2	13	<1
Nigrospora (O)									
Pithomyces (O)				6	80	16	6	40	<1
Rusts (O)							2	13	<1
Unknown (O,I)				4	53	10			
<b>Skin Cells Rating</b>	Trace			Low			Low		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			3 (26 - 75%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Library Wing  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-06  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-06(10)			ME180901-06(11)		
Sample ID	1941-10			1941-11		
Sample Location	Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m³)**	< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) <small>#Cluster-Chain-Loose Spore Profile™</small>						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) <small>## Cluster-Chain-Loose Spore Profile™</small> <small>Cluster(s)</small>						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memnoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected)			0 (None detected)		
<b>Note</b>	No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934) Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: <small>(lab use only)</small> ME180901-07	Telephone No.: 845-559-8537	Company Contact: Tanay Ranadive
Company Name: QUEST	Please select: Fax Report ( ) or Email Report (✓)	Project ID: Q18-1941 Gym
Company Address: 1376 Route 9 Wappingers Falls, NY 12590	Fax No.:	Date/Time sampled: 09/01/18 16:30
	Email address: tranadive@qualityenv.com	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	OUTSIDE PRE-SAMPLE <sup>1548</sup> <del>1558</del>	FD-01HP			3HR	AIR-O-CELL	150	2657-7363
1941-02	GYM LOBBY BY WOMENS RESTROOM <sup>1600-1605</sup>						75	2657-7343
1941-03	GYM LOBBY BY MENS RESTROOM <sup>1601-</sup> 1606						75	2657-7341
1941-04	BY GYM <sup>MENS</sup> LOCKER ROOM <sup>1605-</sup> 1608						75	2657-7371
1941-05	BY GYM WOMENS LOCKER ROOM <sup>1604</sup> 1609						75	2657-7359
1941-06	CENTER OF GYMNASIUM <sup>1606-</sup> 1610						75	2657-7361
1941-07	HALLWAY BY GYM - BOTTOM <sup>1609</sup> 1614						75	2657-7342
1941-08	HALLWAY BY GYM - MIDDLE <sup>1610</sup> 1615						75	2657-7362
1941-09	HALLWAY BY GYM - TOP <sup>1611</sup> 1616						75	2657-7503
1941-10	OUTSIDE POST SAMPLE <sup>1617</sup> 1627						150	2657-7397
1941-11	BACK BLANK						●	2657-7350
1941-12	FIELD BLANK							2657-7357

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP. Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive Date submitted: 9/1/18

Received by: (sign) [Signature] (print) Wu Tang Date and time received: 9/1/18 6:12PM

Page 1 of 1



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Analysis Report

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AIHA EMPAT Lab ID: 178794

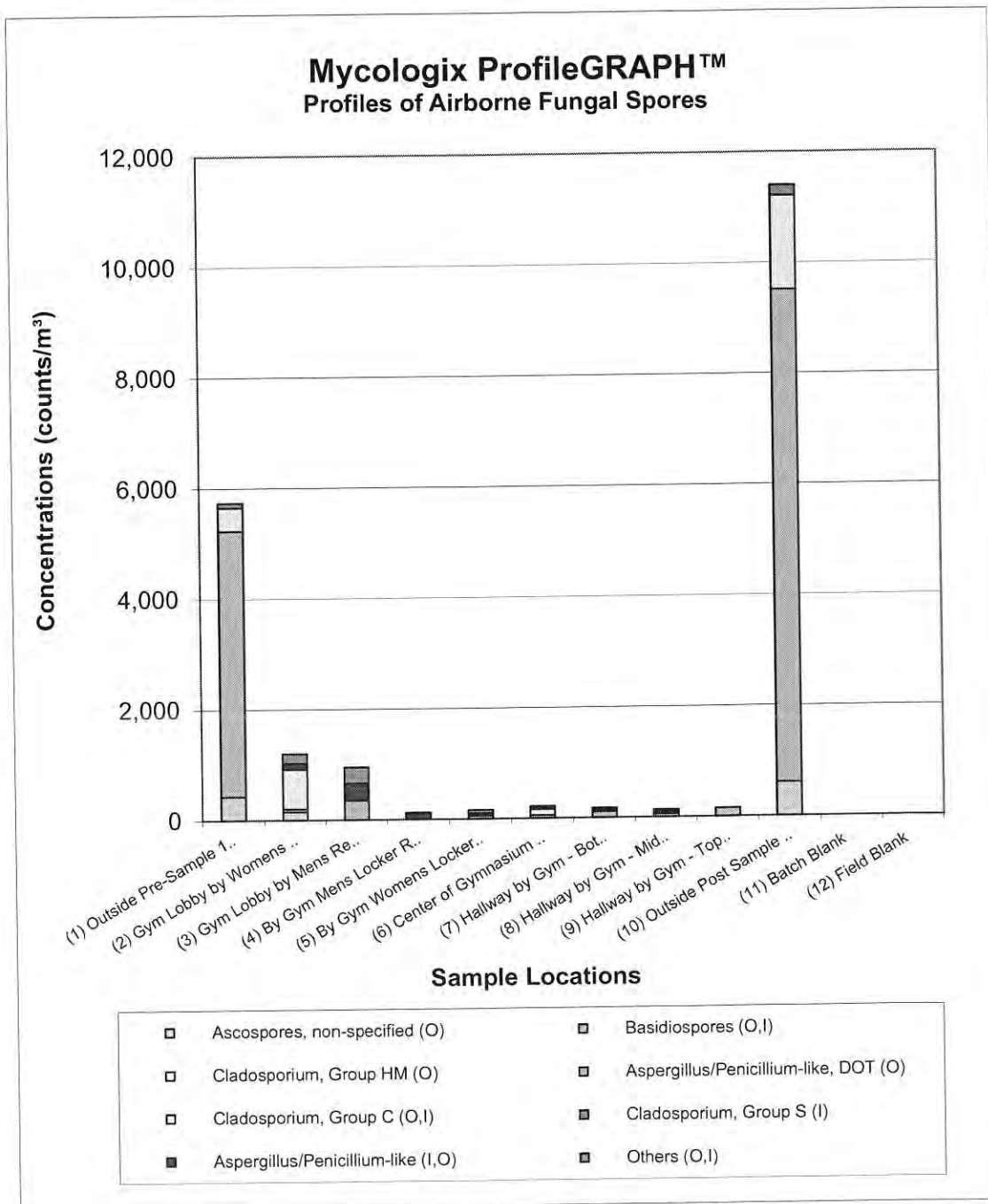
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Gym  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-07  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Reviewed by: WT

Approved by: Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







AccuScience™  
Analysis Report

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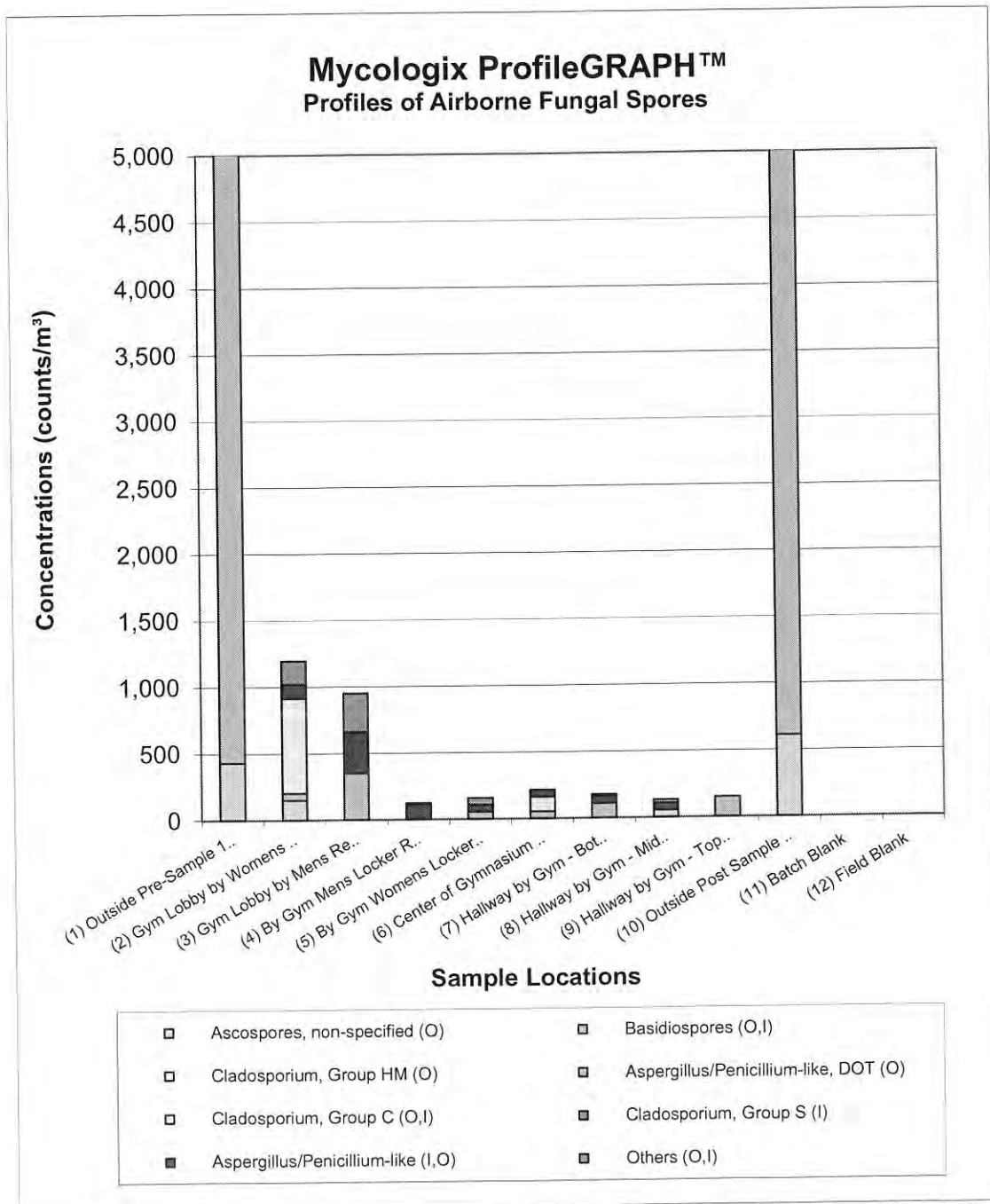
info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
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Please see original data for complete interpretation.





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**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
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**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Gym  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-07  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180901-07(1)			ME180901-07(2)			ME180901-07(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-Sample 1548-1558			Gym Lobby by Womens Restroom 1600-1605			Gym Lobby by Mens Restroom 1601-1606		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	5,700 cts/m³			1,200 cts/m³			950 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	64	430	7	11	150	13			
Basidiospores (O,I)	726	4,800	84	4	53	4	26	350	37
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) <i>#Cluster-Chain-Loose Spore Profile™</i>									
Cladosporium, Group C (O,I)	63	420	7	53	710	59			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) <i>### Cluster-Chain-Loose Spore Profile™</i>				8	110	9	23	310	33
<i>Cluster(s)</i>					0% - 0% - 100%			0% - 49% - 51%	
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)	1	7	<1	2	27	2			
Alternaria (O,I)	2	13	<1						
Cercospora (O)									
Curvularia (O,I)	2	13	<1	1	13	1	1	13	1
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)				2	27	2	10	130	14
Nigrospora (O)									
Pithomyces (O)	4	27	<1	7	93	8	7	93	10
Rusts (O)									
Unknown (O,I)	4	27	<1	1	13	1	4	53	6
<b>Skin Cells Rating</b>	None			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥ 0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Gym  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-07  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-07(4)			ME180901-07(5)			ME180901-07(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	By Gym Mens Locker Room 1603-1608			By Gym Womens Locker Room 1604-1609			Center of Gymnasium 1605-1610		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	120 cts/m³			160 cts/m³			220 cts/m³		
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)				4	53	33	4	53	25
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)							8	110	51
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	8	110	89	4	53	33	4	53	25
Cluster(s)	0% - 0% - 100%			0% - 0% - 100%			0% - 0% - 100%		
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	1	13	11	1	13	8			
Nigrospora (O)									
Pithomyces (O)				3	40	25			
Rusts (O)									
Unknown (O,I)									
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	1 (≤ 5%)			1 (≤ 5%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941 Gym  
**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-07  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-07(7)			ME180901-07(8)			ME180901-07(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	Hallway by Gym - Bottom 1609-1614			Hallway by Gym - Middle 1610-1615			Hallway by Gym - Top 1611-1616		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	180 cts/m³			130 cts/m³			150 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)				4	53	40			
Basidiospores (O,I)	8	110	63				11	150	100
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) ##Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)									
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™	4	53	30	4	53	40			
Cluster(s)	0% - 0% - 100%			0% - 0% - 100%					
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	1	13	7						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)									
Nigrospora (O)									
Pithomyces (O)									
Rusts (O)									
Unknown (O,I)				2	27	20			
<b>Skin Cells Rating</b>	Trace			Trace			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			2 (6 - 25%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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**Client:** QuES&T  
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**Contact:** Ranadive, Tanay  
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**Date Sampled:** 9/1/2018

**QLab Job No.:** ME180901-07  
**Date Received:** 9/1/2018  
**Date Analyzed:** 9/1/2018  
**Date Reported:** 9/1/2018

Lab Sample No.	ME180901-07(10)			ME180901-07(11)			ME180901-07(12)		
Sample ID	1941-10			1941-11			1941-12		
Sample Location	Outside Post Sample 1617-1627			Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			1 smp			1 smp		
Total Concentration (counts/m³)**	11,000 cts/m³			< DL cts/smp			< DL cts/smp		
MycoLogix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)	91	610	5						
Basidiospores (O,I)	1,329	8,900	78						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) <small>#Cluster-Chain-Loose Spore Profile™</small>									
Cladosporium, Group C (O,I)	249	1,700	15						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) <small>###Cluster-Chain-Loose Spore Profile™</small> <small>Cluster(s)</small>									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)	1	7	<1						
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	6	40	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)	5	33	<1						
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	2	13	<1						
Nigrospora (O)									
Pithomyces (O)	11	73	<1						
Rusts (O)									
Unknown (O,I)	3	20	<1						
<b>Skin Cells Rating</b>	Trace			None			None		
<b>Debris Rating</b>	2 (6 - 25%)			0 (None detected)			0 (None detected)		
<b>Note</b>				No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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256 Bridge Street, Metuchen, NJ 08840, USA

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)
Tel: 856-489-0011 www.QLabUSA.com

Lab Job No.: ME180902-01
Telephone No.: 845-559-8537
Company Contact: Taryn Ranaivive
Company Name: QUES&T
Please select: Fax Report ( ) or Email Report (X)
Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY 12590
Fax No.:
Date/Time sampled: 09/02/18 14:30
Email address: taryn@qualityenv.com
P.O. No.:

Table with 7 columns: Sample ID, Sample Location, Analysis Code, Turnaround Time (Std, Day, Hr), Sample Type, Volume (L) or Area (in²), Note. Rows include samples 1941-01 to 1941-09 with locations like Outside Pre-Sample, Kitchen Pantry, Mech. Rm. Side Library, etc.

Sample Types: Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. Material Types: wood, paper, etc.

Common Analysis Codes: Fungi, Direct Exam: (1) Spore Trap: FD-01HP; (2) Tape-lift: FD-02HP; (3) Swab, Bulk, Dust: FD-04HP. Fungi, Culture: (1) Andersen/plate: FC-11; (2) Swab, Bulk, Dust: FC-12

Submitted by: (sign) Taryn Ranaivive (print) Taryn Ranaivive Date submitted: 09/02/18
Received by: (sign) [Signature] (print) [Signature] Date and time received: 09/02/18 4:28 PM





# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™

**Client:** QuES&T  
Wappingers Falls, NY

**Contact:** Ranadive, Tanay

**Project ID:** Q18-1941

**Date Sampled:** 9/2/2018

**QLab Job No.:** ME180902-01

**Date Received:** 9/2/2018

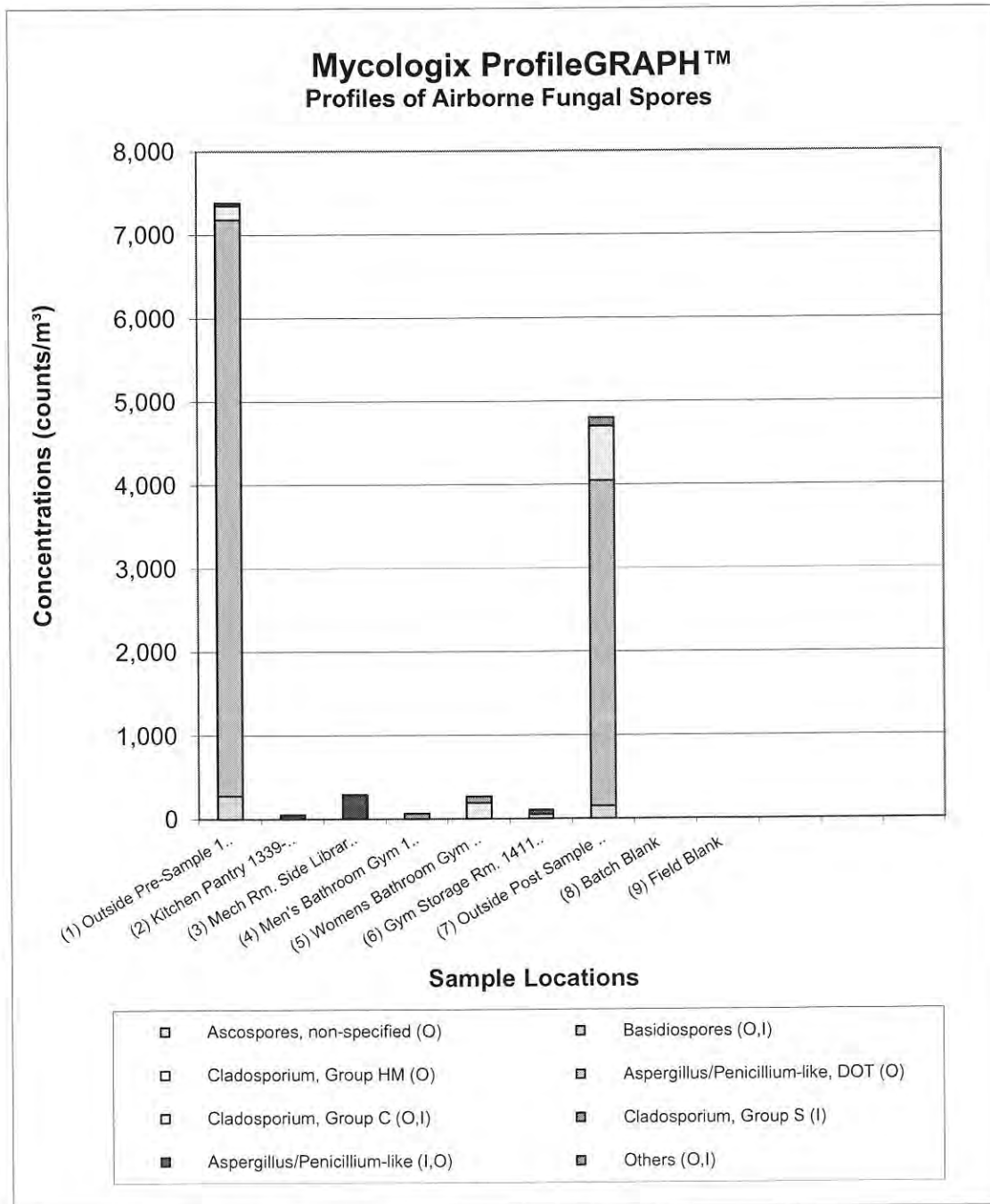
**Date Analyzed:** 9/2/2018

**Date Reported:** 9/2/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 9/2/2018

**QLab Job No.:** ME180902-01  
**Date Received:** 9/2/2018  
**Date Analyzed:** 9/2/2018  
**Date Reported:** 9/2/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180902-01(1)			ME180902-01(2)			ME180902-01(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-Sample 1327-1337			Kitchen Pantry 1339-1344			Mech Rm. Side Library 1347-1352		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			75 L		
Total Concentration (counts/m³)**	7,400 cts/m³			53 cts/m³			290 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)	42	280	4						
Basidiospores (O,I)	1,028	6,900	93						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) ##Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	26	170	2						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)				4	53	100	22	290	100
					0% - 0% - 100%			0% - 82% - 18%	
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	1	7	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	3	20	<1						
Nigrospora (O)									
Pithomyces (O)	1	7	<1						
Rusts (O)									
Unknown (O,I)									
<b>Skin Cells Rating</b>	Trace			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 9/2/2018

**QLab Job No.:** ME180902-01  
**Date Received:** 9/2/2018  
**Date Analyzed:** 9/2/2018  
**Date Reported:** 9/2/2018

Lab Sample No.	ME180902-01(4)			ME180902-01(5)			ME180902-01(6)		
Sample ID	1941-04			1941-05			1941-06		
Sample Location	Men's Bathroom Gym 1356-1401			Womens Bathroom Gym 1403-1408			Gym Storage Rm. 1411-1416		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	75 L			75 L			75 L		
Total Concentration (counts/m³)**	66 cts/m³			270 cts/m³			110 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 53; LQL = 1100 cts/m³		
Ascospores, non-specified (O)									
Basidiospores (O,I)							4	53	50
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) <small>#Cluster-Chain-Loose Spore Profile™</small>									
Cladosporium, Group C (O,I)				14	190	71			
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) <small>### Cluster-Chain-Loose Spore Profile™</small>							4	53	50
<small>Cluster(s)</small>									0% - 100% - 0%
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 13; LQL = 270 cts/m³		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)				1	13	5			
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)									
Nigrospora (O)									
Pithomyces (O)	4	53	80	4	53	20			
Rusts (O)									
Unknown (O,I)	1	13	20	1	13	5			
<b>Skin Cells Rating</b>	Low			Low			Trace		
<b>Debris Rating</b>	2 (6 - 25%)			2 (6 - 25%)			1 (≤ 5%)		
<b>Note</b>									

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.





AccuScience™  
Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840

info@qlabusa.com www.QLABusa.com

AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™

**Client:** QuES&T  
Wappingers Falls, NY

**Contact:** Ranadive, Tanay

**Project ID:** Q18-1941

**Date Sampled:** 9/2/2018

**QLab Job No.:** ME180902-01

**Date Received:** 9/2/2018

**Date Analyzed:** 9/2/2018

**Date Reported:** 9/2/2018

Lab Sample No.	ME180902-01(7)			ME180902-01(8)			ME180902-01(9)		
Sample ID	1941-07			1941-08			1941-09		
Sample Location	Outside Post Sample 1418-1428			Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			1 smp			1 smp		
Total Concentration (counts/m <sup>3</sup> )**	4,800 cts/m <sup>3</sup>			< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m <sup>3</sup>	%	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 27; LQL = 530 cts/m <sup>3</sup>			DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)	23	150	3						
Basidiospores (O,I)	586	3,900	81						
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	98	650	14						
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™									
Cluster(s)									
<b>2. Indoor Hydrophilic Fungi<sup>#</sup></b>	DL = 7; LQL = 130 cts/m <sup>3</sup>			DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memmoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m <sup>3</sup>			DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)									
Alternaria (O,I)									
Cercospora (O)									
Curvularia (O,I)	4	27	<1						
Drechslera/Bipolaris-like (O)									
Epicoccum (O)									
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	5	33	<1						
Nigrospora (O)									
Pithomyces (O)	3	20	<1						
Rusts (O)									
Unknown (O,I)	3	20	<1						
<b>Skin Cells Rating</b>	Trace			None			None		
<b>Debris Rating</b>	2 (6 - 25%)			0 (None detected)			0 (None detected)		
<b>Note</b>				No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



256 Bridge Street, Metuchen, NJ 08840, USA

RUSH!

### Chain of Custody

Toll Free Tel/Fax: 888-QLab-Wei (888-752-2934)  
Tel: 856-489-0011 www.QLabUSA.com

RUSH!

Lab Job No.: <small>(lab use only)</small> ME180904-13	Telephone No.: 845-559-8537	Company Contact: Tanay Ranadive
Company Name: QUEST	Please select: Fax Report ( ) or Email Report (✓)	Project ID: Q18-1941
Company Address: 1376 Route 9 Wappingers Falls, NY 12590	Fax No.:	Date/Time sampled: 09 '04 '18 12:40
	Email address: tranadive@qualityenv.com	P.O. No.:

Sample ID	Sample Location	Analysis Code	Turnaround Time (Std, 1-2 Day, 3-6 Hr)			Sample Type (see below)	Volume (L) or Area (in <sup>2</sup> )	Note (e.g.: material type, weather, etc.)
			Std	Day	3 Hr			
1941-01	Outside Pre-Sample <sup>1156</sup> -1206	FD-01HP			3Hr	Air-O-Cell	150L	2657-7345
1941-02	Mech. Rm Side of Library <sup>1213</sup> -1258	↓			↓	↓	75L	2657-7335
1941-03	Outside Post-Sample <sup>1220</sup> -1236	↓			↓	↓	240L	2657-5949
1941-04	Batch Blank	↓			↓	↓		2657-5937
1941-05	Field Blank	↓			↓	↓		2657-7337

**Sample Types:** Air-O-Cell, Bio-Tape, swab, Andersen, bulk, dust, filter cassette, potable water, non-potable water, etc. **Material Types:** wood, paper, etc.  
**Common Analysis Codes:** Fungi, Direct Exam: (1) Spore Trap: **FD-01HP**; (2) Tape-lift: **FD-02HP**; (3) Swab, Bulk, Dust: **FD-04HP**.  
 Fungi, Culture: (1) Andersen/plate: **FC-11**; (2) Swab, Bulk, Dust: **FC-12**

Submitted by: (sign) Tanay Ranadive (print) Tanay Ranadive Date submitted: 09/04/18  
 Received by: (sign) [Signature] (print) Mindy Wang Date and time received: 09/04/18 3:10 PM  
 Page 1 of 2 QLAB\_C-O-C\_V4.01



# AccuScience™ Analysis Report

QLab, 256 Bridge St, Metuchen, NJ 08840  
info@qlabusa.com www.QLABusa.com  
AIHA EMPAT Lab ID: 178794

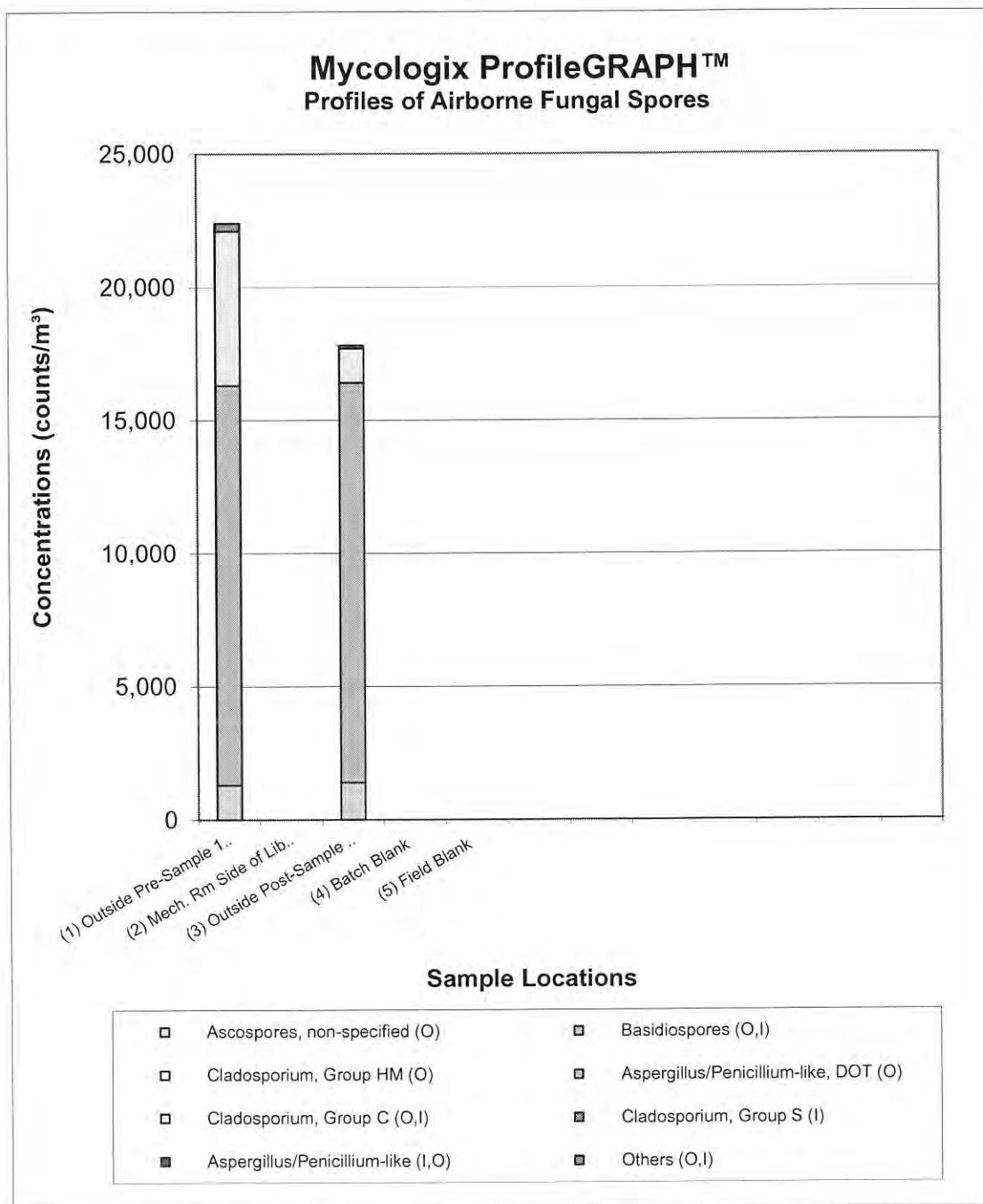
**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 9/4/2018

**QLab Job No.:** ME180904-13  
**Date Received:** 9/4/2018  
**Date Analyzed:** 9/4/2018  
**Date Reported:** 9/4/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Please see original data for complete interpretation.







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Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 9/4/2018

**QLab Job No.:** ME180904-13  
**Date Received:** 9/4/2018  
**Date Analyzed:** 9/4/2018  
**Date Reported:** 9/4/2018

**Reviewed by:** WT

**Approved by:** Wei-Chih Tang, Ph.D., Lab Director

Lab Sample No.	ME180904-13(1)			ME180904-13(2)			ME180904-13(3)		
Sample ID	1941-01			1941-02			1941-03		
Sample Location	Outside Pre-Sample 1156-1206			Mech. Rm Side of Library 1213-1218			Outside Post-Sample 1220-1236		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	150 L			75 L			240 L		
Total Concentration (counts/m³)**	22,000 cts/m³			< DL cts/m³			18,000 cts/m³		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%	cts/smp*	counts/m³	%
<b>1. Common Dominant Spores</b>	DL = 100; LQL = 2000 cts/m³			DL = 53; LQL = 1100 cts/m³			DL = 63; LQL = 1300 cts/m³		
Ascospores, non-specified (O)	196	1,300	6				332	1,400	8
Basidiospores (O,I)	2,190	15,000	67				3,488	15,000	84
Cladosporium, Group HM (O)									
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™									
Cladosporium, Group C (O,I)	876	5,800	26				300	1,300	7
Cladosporium, Group S (I)									
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)									
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 4; LQL = 83 cts/m³		
Stachybotrys (I)									
Chaetomium (I)									
Ulocladium (I)									
Memnoniella (I)									
Trichoderma (I)									
Scopulariopsis (I)									
<b>3. Others</b>	DL = 7; LQL = 130 cts/m³			DL = 13; LQL = 270 cts/m³			DL = 4; LQL = 83 cts/m³		
Hyphal fragment (O,I)	8	53	<1						
Alternaria (O,I)	2	13	<1				7	29	<1
Cercospora (O)	3	20	<1				1	4	<1
Curvularia (O,I)									
Drechslera/Bipolaris-like (O)	2	13	<1						
Epicoccum (O)							1	4	<1
Fusarium (O,I)									
Myxomycetes/Smuts/Periconia (O,I)	3	20	<1				7	29	<1
Nigrospora (O)									
Pithomyces (O)	8	53	<1				7	29	<1
Rusts (O)	6	40	<1				4	17	<1
Unknown (O,I)	12	80	<1						
<b>Skin Cells Rating</b>	None			Trace			None		
<b>Debris Rating</b>	2 (6 - 25%)			1 (≤ 5%)			2 (6 - 25%)		
<b>Note</b>				No fungal structure observed					

\*; cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #. Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.



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Analysis Report

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AIHA EMPAT Lab ID: 178794

**Analysis:** AccuScience Premium Level 3 Fungal Spore Count™  
**Client:** QuES&T  
 Wappingers Falls, NY  
**Contact:** Ranadive, Tanay  
**Project ID:** Q18-1941  
**Date Sampled:** 9/4/2018

**QLab Job No.:** ME180904-13  
**Date Received:** 9/4/2018  
**Date Analyzed:** 9/4/2018  
**Date Reported:** 9/4/2018

Lab Sample No.	ME180904-13(4)			ME180904-13(5)		
Sample ID	1941-04			1941-05		
Sample Location	Batch Blank			Field Blank		
Sample Type (Device)	Air (Air-O-Cell)			Air (Air-O-Cell)		
Air Volume	1 smp			1 smp		
Total Concentration (counts/m³)**	< DL cts/smp			< DL cts/smp		
Mycologix Profile Group 1, 2 & 3	cts/smp*	counts/smp	%	cts/smp*	counts/smp	%
<b>1. Common Dominant Spores</b>	DL = 4 cts/smp			DL = 4 cts/smp		
Ascospores, non-specified (O)						
Basidiospores (O,I)						
Cladosporium, Group HM (O)						
Aspergillus/Penicillium-like, DOT (O) #Cluster-Chain-Loose Spore Profile™						
Cladosporium, Group C (O,I)						
Cladosporium, Group S (I)						
Aspergillus/Penicillium-like (I,O) ## Cluster-Chain-Loose Spore Profile™ Cluster(s)						
<b>2. Indoor Hydrophilic Fungi#</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Stachybotrys (I)						
Chaetomium (I)						
Ulocladium (I)						
Memnoniella (I)						
Trichoderma (I)						
Scopulariopsis (I)						
<b>3. Others</b>	DL = 1 cts/smp			DL = 1 cts/smp		
Hyphal fragment (O,I)						
Alternaria (O,I)						
Cercospora (O)						
Curvularia (O,I)						
Drechslera/Bipolaris-like (O)						
Epicoccum (O)						
Fusarium (O,I)						
Myxomycetes/Smuts/Periconia (O,I)						
Nigrospora (O)						
Pithomyces (O)						
Rusts (O)						
Unknown (O,I)						
<b>Skin Cells Rating</b>	None			None		
<b>Debris Rating</b>	0 (None detected)			0 (None detected)		
<b>Note</b>	No fungal structure observed			No fungal structure observed		

\*: cts/smp: counts per sample. \*\*: All concentrations are rounded to two digits of significant figures. Total concentrations/percentages may not be equal to the sum of individual concentrations/percentages due to rounding. #: Water-loving indoor fungi (min Aw ≥0.89). Absence of hydrophilic fungi does not exclude the possibility of a water damage history. DL: detection limit (analytical sensitivity). LQL: Lower quantitation limit = 20 x DL. Upper quantitation limit depends on sample conditions. ## Asp/Pen-like spores: Loose: 1 to 2 spores; Chain: 3 to 9 spores; Cluster: 10 spores or more. O: Mostly outdoor origin with rare exceptions; I: Mostly indoor origin with rare exceptions. Distinct Outdoor Type (DOT): Distinct outdoor Asp/Pen spores that can be easily differentiated from indoor Asp/Pen spores. DOT is specific to the batch of samples collected at the same time and cannot be used for other batches.

## **Appendix B.1**

### **Chapter 13. Section 2 Mold Assessments**



## CHAPTER 13, SECTION 2

### MOLD ASSESSMENTS: INVESTIGATING, SAMPLING, AND INTERPRETING RESULTS

#### INTRODUCTION

Microbes ... microbiologicals ... bioaerosols ... biological contaminants - all terms for the broad category of airborne particles that are living or have biological origins. This includes fungi, bacteria, viruses, protozoans, pollen, animal dander, insect parts and feces, and human skin scales. While these are all important in indoor environmental quality (IEQ) assessments, by far the most widely suspected, sampled, and publicized members of this group are the fungi.

The number of requests for mold investigations has been steadily climbing, not only in the Navy but nationwide. There are no regulations or standards for mold, so it is often difficult for occupational health professionals to interpret data. Further, most of the underlying reasons for mold contamination originate with building construction or maintenance problems, so it may be difficult for the industrial hygienist to effect the changes needed to resolve occupant complaints.

#### “RULES” FOR MOLD ASSESSMENTS

The following tenets are the foundation for all microbial contamination investigations:

1. **Prevention** is the best way to keep biological contamination from becoming an issue. The key element of prevention is **timely maintenance** and **prompt repair** of facilities.
2. Investigations are a **team effort**, requiring the assistance and cooperation of industrial hygiene, occupational medicine, preventive medicine, safety, occupants, labor representatives (if applicable), facilities and maintenance personnel, housing or office managers, and command public affairs officers.
3. Open, honest **communication** is vital between the personnel conducting the investigation, occupants, and management. At least one team member should be trained in risk communication.
4. If mold is found or suspected, **immediate action** is required to identify and fix the water intrusion source, dry the area, and clean or discard contaminated items. The goal is to minimize the health risk for occupants. While healthy individuals are seldom at risk from mold exposures, there is increased concern for those who are very young, old, debilitated or immunocompromised by other diseases.

## **INVESTIGATOR PROTECTION**

1. Do not disturb contaminated areas or aerosolize biological material.
2. Do not touch visibly contaminated areas with your bare hands. If you do, wash thoroughly with soap and water as soon as possible.
3. If you have to perform destructive sampling in an area (e.g., remove a section of wallboard to access the wall cavity) or disturb a substrate that you suspect is contaminated, use appropriate personal protective equipment (PPE) and lightly spray surfaces with amended water (contains a surfactant) to minimize the possibility of aerosolizing spores.
4. Recommended PPE for those assessing and/or sampling contaminated areas includes: disposable gloves; disposable coveralls; goggles; NIOSH approved half face N-95 respirator (disposable is OK). PPE for remediation projects is discussed in the section 13.3.

## **COMMUNICATION**

The principles and techniques learned in risk communication training are essential for mold contamination projects. Residents in Navy housing and office employees are sure to have seen some of the nationally televised programs about grossly contaminated houses and buildings with occupants who claim they can no longer function normally. There are hundreds of web sites about mold contamination, and most people have read articles about schools or homes contaminated with “toxic molds.”

The more informed employees are about what is happening, the less likely they are to be fearful. Tell them in simple terms what has been found and what you will do to correct the situation. If remediation is required, tell employees what will be done, give them the remediation schedule, and explain how they may be affected (e.g., temporary relocation; control measures; testing). Provide medical support from the cognizant clinic for those with medical concerns or those who develop symptoms they believe to be associated with the contamination. Answer questions honestly and calmly, provide facts sheets tailored to the situation, and provide a contact list for medical and IH issues. Involving employees in the process gives them a stake in the successful outcome.

When mold is found, it is important to make sure that occupants are fully informed about what will be done to correct the building problems. Make sure that points of contact are identified by name so occupants can call if they have IH, medical, or remediation concerns. Section 13.5 contains a detailed discussion of risk communication.

## **ASSESSMENT STRATEGY**

1. **Visual Inspection.** The goals of the investigation are to locate and fix the water intrusion source and to find and remove any associated contamination.

a. Always conduct a thorough visual inspection first, evaluating the building with a critical eye toward potential problem sources. Look for signs of water damage on the ceiling, walls, and floors. Inspect the ventilation system (air handling unit, ducts, fresh air intake location, dampers). Locate odor sources, and look for possible chemical and biological contaminant sources or reservoirs.

b. Likely sources or areas to check for water leaks include the roof; loose or damaged soffits and gutters; chimneys; through-roof pipes or vents; improperly sloped drains; improperly vented appliances, uncontrolled humidity (e.g., moisture condensing on surfaces); improperly installed vapor/moisture barriers or surface finishes (e.g., exterior insulation and finish system [EIFS] or unsealed stucco).

c. Simple tests may be helpful to determine the extent of damage or contamination. For example, a boroscope can be used to check the condition of ventilation ducts. A moisture meter can quickly identify wet building materials. Assessing indoor thermal conditions (temperature and relative humidity [see ASHRAE 55-1992 or Section 13.1 for acceptable ranges] can also help identify areas where mold reservoirs are likely.

d. If mold is found, locate the source of water and repair to prevent additional water damage. Proceed with cleanup and remediation procedures in Section 13.3.

If mold is not found during the visual inspection, but the team believes there is contamination in the building (because of odors, visible water damage, employee illnesses, etc.), take additional investigative steps.

## 2. **Additional Investigation**

a. Review building plans and check maintenance and preventive schedules for possible relations between mechanical component locations, maintenance procedures, and complaints.

b. Talk with employees about their complaints and symptoms, especially anything that they may have noticed different or unusual in the building or whether they detect any pattern in their symptoms or with problems in the building.

c. Check the building's relation to nearby industrial operations for potential pathways that might introduce contaminants.

d. Investigate possible hidden mold reservoirs. This may require destructive procedures, such as removing wall coverings, wall board, carpet or floor covering. Consider that there might be concealed growth behind walls, paneling or wallpaper, under floors, in electrical or plumbing chases, or in ducts.

e. Collecting screening air samples can help locate the general area of unseen mold reservoirs. Consider collecting fungi (spores), total microbial volatile organic compounds (MVOCs), mycotoxins, or glucans.



f. If mold is found, proceed with remediation per Section 13.3. Locate and fix the water source to prevent further intrusion.

If mold is not found, further investigative techniques might include investigating/testing for non-microbial causative agents. Examples include mites, allergens, or neurosensory factors (e.g., visual or perception disruptors).

## **SAMPLING STRATEGY**

DO NOT collect samples without a sampling plan that details how and when samples will be taken, collection requirements for each type of sample, what criteria will be used to interpret results, and what benefits you expect from sampling, i.e., what question(s) will be answered and what actions will result.

ALWAYS consult the analytical laboratory before sampling to ensure sample collection and shipping are done per the lab's requirements and that results will meet your expectations.

### **1. When to Sample**

a. **The rule of thumb in microbial investigations is Do Not Sample when visible mold is present. Regardless of the mold identified or the number of spores, it does not change the requirement to stop the water intrusion and clean up the contamination.**

This is probably one of the biggest challenges during the investigation, since sampling is a natural action for industrial hygienists and a normal expectation from occupants.

b. If you cannot collect a sufficient number of samples to fully characterize the site (i.e., because of funding constraints or insufficient sampling media), it is probably best not to collect any samples. Inadequate sample data usually lead to misleading or confusing results.

c. The investigation team should be guided by their collective expertise in deciding whether or not sampling is indicated. The following are some situations in which bioaerosol sampling is indicated:

- If an occupant has been diagnosed with a disease that is caused by a specific mold or the physician suspects an association between symptoms and mold in the workplace, the physician may request confirmation of the presence of the causative agent.
- If remediation is required, pre- and post-remediation sampling can be used to verify success of the decontamination. Surface samples are especially useful.
- If the investigation team suspects biological contamination but cannot find visible evidence, air sampling may help to verify or locate the reservoir. In

such cases, air sampling could include testing for microbiologicals (viable and/or non-viable), mycotoxins, and/or microbiological volatile organic compounds (MVOCs).

- If litigation is underway or anticipated.
- If the ventilation system was cleaned/ remediated because of microbial contamination (verified by visual or bulk/swab samples), use air sampling to determine if the areas supplied by the system are ready to reoccupy, that is, the ventilation system is not distributing bioaerosols.

d. Because you are sampling bioaerosols whose presence depends on environmental conditions (heat, light, water availability), carefully consider ambient weather conditions. For example, rain can “wash” the air clean of many spore types, such that sampling on rainy, foggy, or very humid days can result in low outdoor counts or species distributions that are significantly different from those on warm, sunny days. In general, levels of ascospores and basidiospores will be higher during rainy weather.

Sampling when there are strong winds can result in outside counts that are significantly higher than on non-windy days. In addition, high outdoor counts may mask small to moderate indoor mold problems since interpretation is dependent in part on ratios of indoor to outdoor spore counts.

Compensate for ambient conditions by adjust your sampling schedule if possible. At least be aware that outside samples may not represent normal conditions so that you do not misinterpret results.

## 2. Where To Sample

a. Complaint/problem area – Use complaint patterns, symptom descriptions, and visual indications to guide you in choosing sample locations. You may need a sampling array within a single office, on an entire floor of the building, or throughout the building to get results that are representative. Preliminary or screening samples may help target the areas that require further characterization.

b. Non-complaint area – Results serve as controls to compare with complaint area results.

c. Outside – Outside samples must be taken at the same time as indoor samples so that the types and quantities of ambient flora can be compared with those in the building. Ideally, at least one outside sample is collected at the fresh air intake that supplies the inside area being sampled.

3. Number of Samples. There is no formula to determine how many samples you need to adequately characterize a complaint area. Further, statistical validity considerations cannot be used because of the difficulty in predicting the environmental variability. The AIHA *Field*

*Guide for the Determination of Biological Contaminants in Environmental Samples* gives this guidance:

- a. The number of samples depends on the size and organization of the space being investigated.
- b. Sample as many locations within the area of study, control locations, and outdoors as is practically and economically feasible.
- c. When possible, take duplicate side-by-side samples. According to Chapter 3 of the *AIHA Field Guide*, “duplicate side-by-side sampling is considered adequate to define the mean and the random sampling and analysis error given the high temporal and spatial variability of bioaerosol concentrations in air.” .... “Acceptability of the agreement between side-by-side duplicate samples must be determined by the investigator based on the intended use of the data.”
- d. Investigate temporal variations by sampling at least two time periods during the day, preferably separated by a long interval, e.g., morning and late afternoon. Sample on different days or during different seasons if daily/seasonal variations appear to influence conditions.

#### 4. **Choosing the Appropriate Type of Sample – Bulk, Surface, Air**

Before taking a sample, think about why you need the result and what you want the results to tell you. If you’re trying to determine if an area is contaminated or if what you see is really mold, a swab or bulk sample is sufficient. If you are trying to support a medical diagnosis, a viable sample is needed for the lab to identify and speciate the mold.

A brief discussion of the methods to collect fungal samples for identification and/or quantification follows. [Appendix 13.2-A](#) summarizes sampling methods, their strengths and limitations, and provides resources for more information.

- a. **Bulk samples** – Used to identify contaminants, especially when trying to locate or confirm the presence of a mold species as a causative agent for medical diagnosis. Examples of materials that might be collected include carpet, insulation, duct lining, wallpaper, or wallboard (sheetrock).

Collect samples from visibly contaminated surfaces by scraping or cutting with clean tools (e.g., wall board). Place sample in a clean, plastic bag and label for transport.

**Bulk water samples** can be collected from condensate drain pans, cooling towers (i.e., for *Legionella*), or other water reservoirs suspected of being a contaminant source. Collect in a sterile container, seal tightly, and transport in a secondary container such as a ziplock bag to contain the sample in case of breakage or leaks.

Another type of bulk sample can be taken using a **microvacuum**. This is basically a cassette attached to a pump that is used to vacuum carpets, furniture, or other substrates to collect the



particulate matter. Though the sample can be randomly vacuumed into the cassette, using a specific grid collection area will allow quantitative results.

b. Surface samples – Surfaces can be sampled by swabbing or using clear cellophane tape (also called a “sticky tape” sample). The sample is analyzed by direct microscopic examination to determine if there is microbial contamination. Sterile swab collections can be cultured for identification. Surface sampling is limited to identifying settled fungi or spores and may not be related to airborne results.

c. Settling plates/ gravity plates – Open nutrient agar petri dishes are placed on a flat surface to collect anything that settles out of the air. Results are not particularly meaningful, since what grows depends on random settling of airborne particulates onto a non-specific growth medium. Navy personnel will not use this method.

d. Air samples. Air sampling is the most common collection method for bioaerosols. A pump is used to draw in air and deposit the particulate onto a collection medium. Most air sampling methods can be used for microscopic analysis, a few for culturing techniques, and others for specialized testing. Each kind of air sample has its benefits and disadvantages, depending on the media used and the collection and analytical method chosen. Regardless, air sample results for molds are subject to false negative results. That is, there may be contamination present even when results indicate otherwise. Consider:

(1) Most samples are very short, and therefore capture only 5 - 10 minute snapshots of what is actually happening at the sample location.

(2) You have to decide whether you want viable or non-viable results (culturable or non-culturable, respectively) before you sample.

(3) If the sampling method does not have sufficient collection efficiency in the size range(s) of the mold present, you will not collect the spores.

(4) If collecting samples directly onto agar (viable sample), you are likely to miss molds that have special growth requirements.

5. **Choosing the Sampling Method.** [Appendix 13.2-A](#) summarizes sampling methods, advantages/ disadvantages, and provides information resources. You may want to refer to the chart during discussion.

a. Viable (culturable) vs. Non-viable (non-culturable) samples

(1) Viable (culturable) samples are collected on nutrient agar initially, or can be collected in/on inert media and prepared for culture at the laboratory. Samples are incubated for several days to allow cell growth and replication into visible colonies. The entire colony, not just the spore, is used for the identification, allowing the lab to make a more exacting identification of certain mold types.

Culturable samples tend to underestimate the number of total spores present, since only viable organisms will grow. Of the viable fungi that impact onto the agar during sampling, only a percentage of those will actually grow during incubation.

Further, remember that some molds require specific nutrients or growing conditions. If these are not present, the organism will grow very slowly or not at all. For example, *Stachybotrys* requires cellulose. If you use Malt Extract Agar (MEA) for sampling and the report shows no *Stachybotrys*, this means that (1) there really was no *Stachybotrys* in the sampled area; or (2) *Stachybotrys* was present but MEA did not support its growth. If you suspect *Stachybotrys*, contact the laboratory to determine the nutrient agar of choice (usually Rose Bengal or cellulose agar) for collecting viable samples. A better alternative is to collect a non-viable sample since the spores are very distinctive and can easily be identified by direct microscopic examination.

You can also get relatively good recovery of fungi (about 60%) if you collect the sample using a button sampler and filter media. Note that if you use this method to collect a bacteria sample, recovery is <20% due to cell dessication. You can increase bacteria recovery to around 60% by using a gelatin filter.

(2) Non-viable samples are examined directly under a microscope to identify and count spores and other particulates (e.g., fibers, skin cells, mycelial fragments) based on morphological features. Some molds, such as *Aspergillus* and *Penicillium*, cannot be distinguished by their spores alone, so they are reported as a group, e.g., *Aspergillus/ Penicillium* group; *Drechslera/Bipolaris* group; or Smuts/*Periconia/Myxomycetes*.

b. Bioaerosol collection principles. In general, collecting bioaerosols involves either filtration or impaction. Figure 13.2-1 shows the collection methods and the possible analyses that can be performed using each.

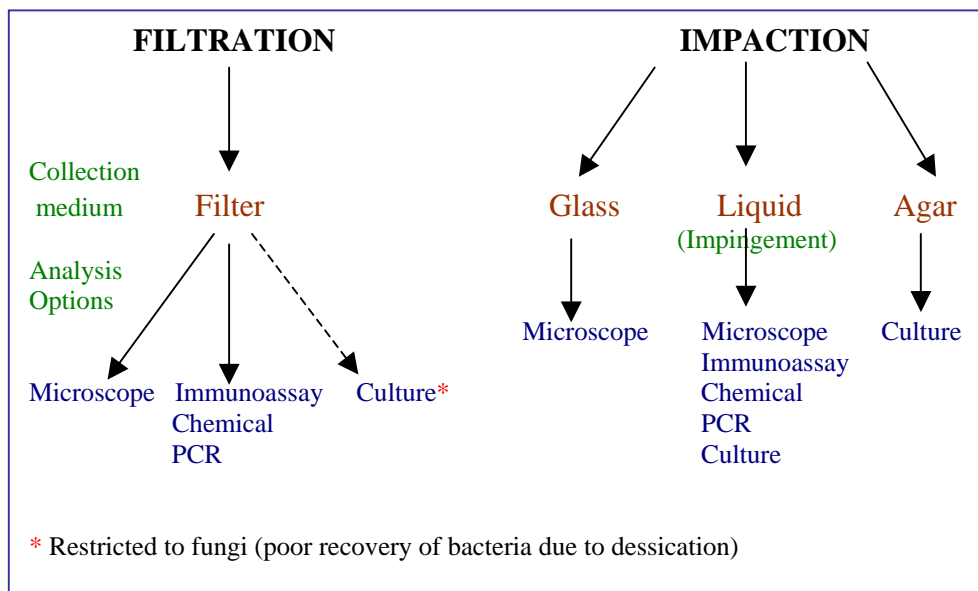


Figure 13.2-1. Bioaerosol collection methods.

(1) Filtration - Bioaerosol is collected on a filter as air passes through it. Filter media can have different diameters, pore sizes, and composition, so consult the laboratory before sampling.

(2) Impaction – Bioaerosol is impacted onto a collection media such as glass (may contain collection strip of agar, grease, adhesive, or tape), agar plates, or liquid. Impaction into a liquid medium is also called impingement.

c. Why mold size is important

Particle collection efficiency is driven by the size of the particle you want to collect. With spores, this can make the difference in whether a negative result means there really is no mold present – or that the mold is there but you didn't collect it.

[Appendix 13.2-A](#) lists collection efficiencies of some of the more commonly used methods. For example, if you suspect that you have *Cladosporium cladosporoides* contamination and you sample using a Burkard spore trap (impaction onto slide), you'll probably get negative results. Notice that the 50% cut size of the Burkard sampler is around 2.5  $\mu\text{m}$ . Since *C. cladosporoides* is around 2  $\mu\text{m}$ , you will miss most – if not all – of the spores simply because the collection device is inefficient (about 10%) at 2  $\mu\text{m}$ .

## 6. Sampling Tips

a. Before collecting any samples, select the analytical lab you will use. Call the lab to ensure that you sample according to their requirements, especially if you are taking viable culture samples.

b. In some cases, the laboratory may provide the sample collection equipment. For example, most labs will loan you an Andersen N6 and provide the correct agar for the targeted biological population. For non-viable sampling, you will have to purchase the sampling media (e.g., Air-O-Cell cassettes), but the lab may loan you the high volume pump.

c. Sampling conditions should be reflective of “normal” building conditions. The ventilation system should be on the usual daily setting (i.e., temperature, damper opening(s), setbacks, auxiliary/booster fan operation, fresh air intake settings, etc.) and employees should work as they typically do. **DO NOT** intentionally alter the area to be sampled.

d. Sample on different days and at different times of the day to get samples that represent conditions over time. Replicate samples are a good idea to increase confidence in your results. Remember that results tend to be less reliable or repeatable when sample times are very short!

e. Aggressive sampling is not recommend for investigational studies. While aggressive techniques will disturb accessible mold reservoirs, it complicates result interpretation because it is not representative of normal building conditions.

f. Record ambient conditions during sample collection, such as temperature and relative humidity. Also make notations of conditions inside that may impact results, such as obvious water damage or contamination in relation to the sample location; potential microbial reservoirs, like fish tanks, plants or trash; condition of HVAC system components; presence of pets; or open/ leaky doors and windows. Outside sample notes should include weather conditions (cloud cover, recent precipitation, wind) and locations of land features (ditches or standing water, landfills, playgrounds, construction areas).

g. Chain of Custody (COC). It is prudent to use a COC form with your bioaerosol samples. The COC is particularly important should you become involved in litigation, but should be used anyway to track the samples' journey from collection to analysis. If you don't have a COC form, most labs will supply you with one. You can view examples at the following sites:

<http://www.aerobiology.net/COC.pdf>

<http://www.aerotechlabs.com/InfoBase/cocs.aspx>

<http://www.emlab.com/media/resources/submit.pdf>

[http://www.emsl.com/new\\_chain.pdf](http://www.emsl.com/new_chain.pdf)

<http://www.stl-inc.com> (Go to Our Labs > P&K Microbiological > Chain of Custody)

## 7. **Sampling for Fungal Metabolites and Cell Components**

### a. Microbial Volatile Organic Compounds (MVOCs)

(1) MVOCs are produced by molds that are metabolically active. They are also responsible for many of the musty odors associated with molds. If you 'smell mold' but cannot see it, MVOC sampling may help to locate the fungal reservoir.

(2) Collect samples using low flow pumps and sorbent tubes as specified by the lab. Ship samples on ice and protect from heat and light.

### b. Mycotoxins

(1) Fungi are primarily saprophytic, that is, they use nonliving organic material as the nutrient source for growth and reproduction. During the digestion process, fungi secrete enzymes to help break down complex compounds into simpler ones that can be taken up and digested. The by-products of digestion are classified as primary or secondary metabolites.

(2) Primary metabolites are produced from cellulose and other compounds that are used by the fungus for energy, growth, and reproduction. Secondary metabolites, called mycotoxins, are natural by-products that are not necessary for growth and are usually derived from precursors formed during primary metabolism. They are thought to give the fungi a competitive edge against other microorganisms, including other fungi.

Whether a toxigenic fungus actually produces mycotoxins appears to depend on environmental conditions, including temperature, growth substrate, and pH. Some of the mycotoxins most



commonly associated with mold contamination in buildings are briefly described in [Appendix 13.2-B](#).

(3) Exposure. Mycotoxins accumulate in spores, mycelium, and growth substrates. Consequently, they can be inhaled (when spores or substrates are disturbed and aerosolized), ingested (consuming toxin-containing spores when eating, drinking or smoking in a contaminated area), or absorbed through the skin (e.g., when handling contaminated materials).

(4) Health Effects. Symptoms associated with exposure to mycotoxins include dermatitis, cold and flu symptoms, sore throat, headache, fatigue, diarrhea, inflammatory reactions, and impaired or altered immune function (which can lead to opportunistic infection). Many toxin-producing fungi, such as *Stachybotrys*, *Penicillium*, *Aspergillus* and *Fusarium* species, have been linked to illnesses resulting from exposure to fungi growing in water-damaged buildings. Other mycotoxins have been associated with cancer (e.g., aflatoxins from *Aspergillus*), cardiovascular effects (ergot alkaloids), and neurological symptoms (*Aspergillus fumigatus*).

(5) Sampling and Analysis. Contact the laboratory before collection for specific lab instructions. Because air sampling for mycotoxins has limitations, bulk, surface, or dust samples are usually best.

- Air - Collect on filter cassette. Store samples at ambient temperature under desiccated conditions.
- Dust/Bulk - Select an area with visible contamination, and collect 25-50 grams of material in a suitable container. Microvacuuming and surface swab (methanol swab) techniques can also be used.
- Water - Collect 5 ml of water. Seal. Refrigerate and ship to the laboratory via overnight courier under refrigerated conditions.

### c. Glucans and Ergosterol

(1) Both (1→3)- $\beta$ -D-glucan (glucans) and ergosterol are fungal cell wall components of filamentous fungi, which includes most saprophytes. These compounds have been sampled successfully as chemical markers to show that such fungi are present. Sampling will detect ergosterol in both living and dead spores (ergosterol is fairly stable in spores).

(2) Collect glucans on a membrane filter, extract, and analyze using a *Limulus* amoebocyte lysate (LAL).

(3) Ergosterol sampling is also done on a filter that is extracted to remove the ergosterol. Analysis can be done using high performance liquid chromatography (HPLC), gas chromatography (GC), or GC with mass spectrophotometry (MS).

(4) There is little data comparing the number or mass of spores to chemical marker concentrations.

d. PCR (polymerase chain reaction) analysis provides genetic confirmation of certain fungal species using species-specific DNA probes or primers. PCR is quick and specific, but the technology is limited to the species probes available for fungal confirmation.

Consult the individual laboratories that offer PCR to determine what fungi are in their detection panels. Some of the available probes/primers of interest developed to date include: *Alternaria alternata*; *Aspergillus flavus*, *fumigatus*, *niger*, *sydowii*, *versicolor*; *Chaetomium globosum*; *Cladosporium cladosporoides*; *Penicillium aurantiogriseum*, *brevicompactum*, *chrysogenum expansu*, *griseofulvum*, *purpurogenum*, *viridicatum*; *Stachybotrys chartarum*; and *Ulocladium botrytis*.

## 8. Sampling for Bacterial Cell Components: Endotoxins

a. Endotoxins are found in the cell walls of gram negative bacteria. Made of lipopolysaccharides, they can elicit health effects in susceptible individuals whether the bacteria is viable or not. The most common exposure routes are inhalation and ingestion. Gram negative bacteria are most often associated with water, sewage, humidifiers, and gray/black water contamination.

b. Air samples are collected using endotoxins-free polystyrene cassettes. The samples must be collected carefully to ensure there is no human contamination.

Bulk water samples can be taken in endo-free vials, again using sterile techniques, and must be kept on ice for shipment to the lab.

## SAMPLE ANALYSIS

1. Use only analytical laboratories that are proficient in the Environmental Microbiology Proficiency Analytical Testing (EMPAT) program. The EMPAT evaluates the lab's ability to correctly identify cultured fungi and bacteria that might be found in mold contamination investigations. Under the current program, labs must correctly identify the genus, and they receive bonus points for correctly speciating the organism. The EMPAT certificate states whether the proficiency is for identification of bacteria, fungi, or both.

At this time, proficiency testing does not involve counting (of spores or colonies) or identifying organisms from mixed cultures.

Beware of laboratories that advertise that they *participate* in the EMPAT rather than that they are *proficient* in the EMPAT.

The American Industrial Hygiene Association (AIHA) administers the EMPAT. Consult their web site at <http://www.aiha.org/LaboratoryServices/html/micro.htm> for the most current proficiency testing results and accreditation category.

2. While not required at this time, it is recommended that analytical laboratories also be accredited through the Environmental Microbiology Laboratory Accreditation Program (EMLAP). This program assesses and rates various lab parameters, such as: personnel qualifications, EMPAT scores (performance), facilities, quality assurance programs, record-keeping, analytical methods, and operating procedures. EMLAP also includes triennial site visits to the laboratory. Details are at <http://www.aiha.org/LaboratoryServices/html/micro.htm>.
3. Consult with the laboratory to define sample collections methods, turnaround time, costs, shipping requirements, and exactly what the analysis report includes. For example, some labs clearly report genus and spore count, e.g., *Cladosporium* 450 spores/m<sup>3</sup>, while others may report results as *Cladosporium*-like (not definitive for *Cladosporium* but spores look similar) or *Cladosporium* 42 spores (you have to calculate concentration).
4. [Appendix 13.2-C](#) provides a consolidated list of environmental microbiology laboratories, bioaerosol services available, accreditation status, and contact information. Please provide changes or additions to [IH-Director@nehc.mar.med.navy.mil](mailto:IH-Director@nehc.mar.med.navy.mil).

## **INTERPRETING RESULTS**

The presence of mold does not mean that occupants will have adverse health effects or that they will even be exposed. Like any other stressor, you must have a completed exposure pathway to the biocontaminant. The mold or mold fragments, spores, or metabolites must be produced, released, reach the occupants, then be inhaled, physically contacted, or ingested. Even after contact, human response will depend on individual susceptibility (e.g., genetic predispositions to allergens, age, health status) and type of exposure (allergen, toxin, infectious agent).

There are no standards for biological sample results. The American Industrial Hygiene Association, American Conference of Governmental Industrial Hygienists, Environmental Protection Agency, and numerous other resources agree that the best criteria for interpreting results is to compare inside samples with outside and/or contaminated areas with uncontaminated areas, along with consideration of both the kinds of mold present (genus/species) and the numbers (spore or colony counts).

### **Interpretation Criteria**

- a. Compare Indoor and Outdoor Results. An effective interpretation is based on comparing inside and outside sample results. In general, inside counts should be around 30-80% of outside and have the same general distribution of genera.
  - Rank order the genera/species results. The relative order inside should be similar to outside. If the dominant types of mold in indoor samples are not the same as those in

outdoor samples, it indicates an indoor mold source.

- The concentration of each genus/species identified inside should be less than outside. Higher inside levels indicate there is fungal amplification indoors.
- The presence or absence of a few genera in small numbers should not be considered abnormal.
- Normal outside fungi typically include *Cladosporium*, *Alternaria*, *Epicoccum*, and Basidiomycetes, so it is common to see these identified in indoor samples.
- The presence of certain fungi indoors should prompt immediate risk management decisions. Examples of fungi of concern include *Aspergillus versicolor*, *Aspergillus flavus*, *Aspergillus fumigatus*, *Stachybotrys chartarum*, *Fusarium moniliforme*, *Histoplasma*, and *Cryptococcus*.
- Numerical guidelines can be useful as a secondary interpretive resource when evaluating viable sample results (i.e., reported in colony forming units per cubic meter of air (CFU/m<sup>3</sup>)). Fungi levels in excess of these numbers do not mean that the conditions are unsafe or hazardous. Do not use these guidelines for non-viable sampling results.

< 150 CFU/m<sup>3</sup> total fungi is acceptable if the reported genera are reflective of normal outdoor flora (e.g., *Cladosporium* and other leaf and tree fungi).

< 500 CFU/ m<sup>3</sup> total fungi is acceptable in summer if the reported genera are reflective of normal outdoor flora.

> 50 CFU/m<sup>3</sup> of a single species other than *Cladosporium* or *Alternaria* should prompt further investigation.

>1000 CFU/m<sup>3</sup> total fungi indicates potential building related problems and requires further investigation.

b. Consider Outside Air Entry.

- Filtered or conditioned air will affect the relative numbers of genera. In an office building with little fresh outside air or poor air exchange rates, ‘normal’ inside counts may be very low, i.e., 2-5% of outside. The rank order of genera should be similar.
- If sampling in a building or residence when doors and/or windows are open, expect ‘normal’ inside counts to be very similar to outside – as high as 95%. The rank order of genera should be similar.



c. Put Results in Context With Other Facts.

- On microscopic examination, morphologically similar spores cannot be differentiated. The most common example of this is with *Aspergillus*, *Penicillium*, *Gliocladium*, *Trichoderma*, and other small, round, colorless spores. Non-culture results will report all such spores as *Aspergillus/Penicillium* group.

If results show high indoor counts of *Aspergillus/Penicillium*, you may want to collect samples for culture to separate the genera and determine which species of *Aspergillus* is present, since several produce mycotoxins and are infectious.

- Myxomycetes/Rust/Smut or Smuts/Periconia/Myxomycetes will also be reported together on non-cultured sample reports. These are morphologically similar round, brown spores that are common outdoor plant molds.
- The presence of fungal fragments such as hyphae or conidiophores suggests colonization, growth, or accumulation of fungi in the sampling location.
- The presence of yeast suggests wet conditions.
- Be sure you know the ambient sampling conditions before using outside results:

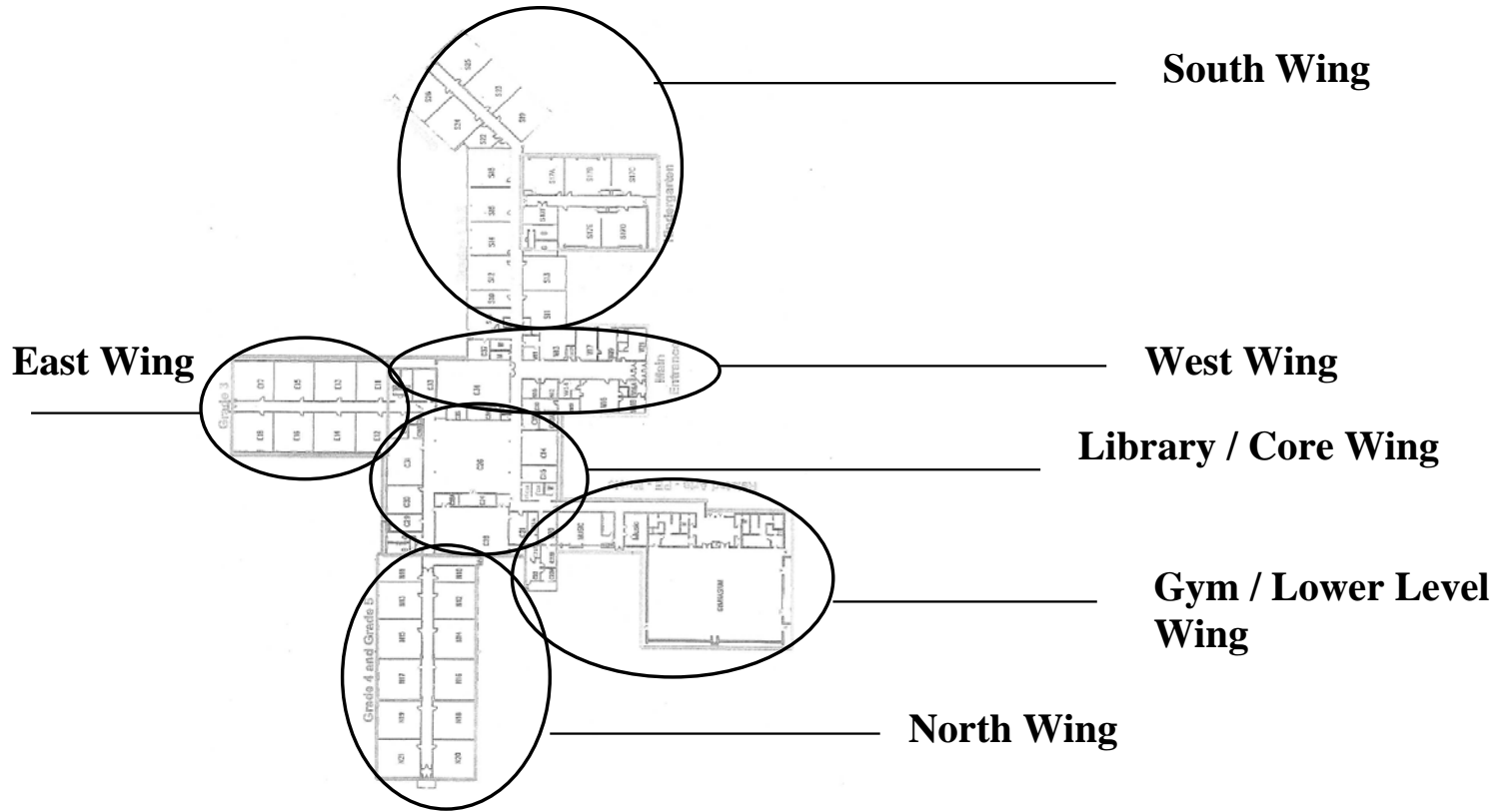
Outside samples collected during or soon after rain will usually have lower total spore counts but higher relative concentrations of ascospores and basidiospores.

Expect higher concentrations of fungi in warmer weather, lower total counts in cooler weather.

# Appendix C

## Map

# Pequenakonck Elementary School



School Floor Key Plan

\*\*Drawing Not to Scale\*\*

This Drawing is not intended to be used as the sole basis for soliciting pricing for asbestos abatement. An abatement plan, specification, drawing and/or variances should be developed to identify scope, timing, phasing and remediation means & methods for any asbestos project.

**North Salem CSD**  
230 June Road  
North Salem, NY 10560

**Pequenakonck Elementary School**  
230 June Road  
North Salem, NY 10560



Quality Environmental  
Solutions & Technologies, Inc.  
1376 Route 9  
Wappingers Falls, NY 12590  
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**Date:** 9-12-2018  
**Version #** 1

**Issued For:**  
Post-Remediation Assessment

**Project NO:**  
Q18-1941

**Project Manager:**  
Larry Holzaphel

**Drawing Prepared By:**  
Tanay Ramadive

**01**

**Appendix D**  
**Mold Assessment Documentation**  
**South Wing**



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	S-8	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/24/2018											
Time:	1100											
Assessor:	Tanay Ranadive											
											Response Action	
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate
		Yes	No		Yes	No	Yes	No	Yes	No		
Walls	North	X			X			X	X		<b>Cove Base on Wall</b>	<b>#2 Clean, #3 Encapsulate</b>
	East		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	South		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	West		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Ceiling	Tiles		X		X			X	X			<b>#2 Clean, #3 Encapsulate</b>
	Above Ceiling		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Pipes/Insulation/Etc.		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Drip Pans		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Lighting		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Insulation		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Other		X			X		X	X			
Floor	Carpet Front		X			X		X	X			<b>#1 Dispose of Carpet</b>
	Carpet Back	X				X		X	X			<b>#1 Dispose of Carpet</b>
	Tiles		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Doors	Classroom Door		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Closet Door		X			X		X	X			
	Bathroom Door		X			X		X	X			
Door Frames	Classroom		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
	Bathroom		X			X		X	X			
	Closet Door		X			X		X	X			
	Other (Describe)		X			X		X	X			
Windows	Frame/Sills/Sash/Curtains		X		X			X	X		<b>Window Components to be Cleaned</b>	<b>#2 Clean, #3 Encapsulate</b>
Bookcases	<i>all sides, top, bottom</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
File Cabinets			X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Inside Closets			X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Bulletin Boards	<i>Check Behind</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Chalkboards	<i>Check Behind</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
White Boards	<i>Check Behind</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Wallpaper	<i>Check Behind</i>		X			X		X	X			
Wall Artwork	<i>Check Behind</i>		X			X		X	X			
Books/Magazines Etc.			X			X		X	X			<b>#1 Dispose of Books, #2 Clean, #3 Encapsulate</b>
Room Contents	<i>Games Easels, etc.</i>		X			X		X	X			
Desks	<i>all sides, top, bottom</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Chairs	<i>all sides, top, bottom</i>	X			X			X	X		<b>Tennis Balls on Chairs</b>	<b>#1 Dispose of Tennis Balls, #2 Clean Chairs, #3 Encapsulate</b>
HVAC system	<i>Supply/Return/Filters/Ducts</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Unit Ventilators	<i>Filter/Cage/Cover</i>		X			X		X	X			<b>#2 Clean, #3 Encapsulate</b>
Equipment	<i>all sides, top, bottom</i>		X			X		X	X			
Clothes			X			X		X	X			
Boxes			X			X		X	X			
Backpacks, shoes,			X			X		X	X			
Leather goods			X			X		X	X			
Shelves			X			X		X	X			
Under Sinks/Cabinets			X			X		X	X			

# Mold Assessment Field Documentation Sheet

<b>School Name:</b>	Pequanock Elementary	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.
<b>Room #:</b>	S-Wing 2nd Grade	
<b>Room Type:</b>	Hallway	
<b>Date:</b>	8/23/2018	
<b>Time:</b>	930	
<b>Assessor:</b>	Frank Manna & Michael Smith	

		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action
Room Component		Yes	No		Yes	No	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		x			x	X				
	East	x		20 SF		x	X			Adjacent to S10 & Bathroom	#3 on Sheetrock
	South		x			x	X				
	West	x		14 SF		x	X			Between S13 - S11	#1 Dispose of 14 SF of Sheetrock
	Ceiling	Tiles	x		50 SF		x	X			Multiple Locations 7 - Ceiling Tiles
	Above Ceiling		x			x	X				
	Pipes/Insulation/Etc.		x			x	X				
	Drip Pans		x			x	X				
	Lighting		x			x	X				
	Insulation		x			x	X				
	Other		x			x	X				
Floor	Carpet Front		x			x	X				
	Carpet Back		x			x	X				
	Tiles		x			x	X			Clean Hallway Floors	#2 Clean All Floors
	Doors	Classroom Door		x			x	X		Clean Multiple Classroom Doors	#2 Clean Classroom Doors
	Closet Door		x			x	X				
	Bathroom Door		x			x	X				
Door Frames	Classroom		x			x	X			Clean Classroom Doorframes	#2 Clean Door Frames
	Bathroom		x			x	X				
	Closet Door		x			x	X				
	Other (Describe)		x			x	X				
Windows	Frame/Sills/Sash/Curtains		x			x	X			Clean Window System	#2 Clean all Window Systems Within Hallway
Bookcases	<i>all sides, top, bottom</i>		x			x	X				
File Cabinets			x			x	X				
Inside Closets			x			x	X				
Bulletin Boards	<i>Check Behind</i>		x			x	X			Clean All Bulletin Boards in Hallway	#2 Clean Bulletin Bords
Chalkboards	<i>Check Behind</i>		x			x	X				
White Boards	<i>Check Behind</i>		x			x	X				
Wallpaper	<i>Check Behind</i>		x			x	X				
Wall Artwork	<i>Check Behind</i>		x			x	X				
Books/Magazines Etc.			x			x	X				
Room Contents	<i>Games Easels, etc.</i>		x			x	X				
Desks	<i>all sides, top, bottom</i>		x			x	X				
Chairs	<i>all sides, top, bottom</i>		x			x	X				
HVAC system	<i>Supply/Return/Filters/Ducts</i>		x			x	X				
Unit Ventilators	<i>Filter/Cage/Cover</i>		x			x	X				
Equipment	<i>all sides, top, bottom</i>		x			x	X				
Clothes			x			x	X				
Boxes			x			x	X				
Backpacks, shoes,			x			x	X				
Leather goods			x			x	X				
Shelves			x			x	X				
Under Sinks/Cabinets			x			x	X				







# Mold Assessment Field Documentation Sheet

School Name:	Pequanock Elementary	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.									
Room #:	Classroom S11										
Room Type:	2nd Grade Classroom										
Date:	8/23/2018										
Time:											
Assessor:	Frank Manna & Michael Smith										
										Response Action	
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate
		Yes	No		Yes	No		No	Yes		
Walls	North		x			X		X			
	East		x			X		X			
	South	x		8 SF	x			X		<b>Under Whiteboard-Walls to be Cleaned</b>	<b>#3 Encapsulate</b>
	West	x		6SF	x			X		<b>Walls to be Cleaned</b>	<b>#2 Clean, #3 Encapsulate</b>
Ceiling	Tiles	x			X			X		<b>3 - 2' x 4' Ceiling Tiles</b>	<b>#3 Encapsulate</b>
	Above Ceiling		x			x		X			
	Pipes/Insulation/Etc.		x			x		X			
	Drip Pans		x			x		X			
	Lighting		x			x		X			
	Insulation		x			x		X			
	Other		x			x		X			
Floor	Carpet Front		x			x		X			
	Carpet Back		x			x		X			
	Tiles		x			x		X			
Doors	Classroom Door	x				x		X		<b>Door to be Cleaned</b>	<b>#2 Door Cleaned</b>
	Closet Door	x				x		X		<b>Doors to be Cleaned</b>	<b>#2 Doors Cleaned</b>
	Bathroom Door		x			x		X			
Door Frames	Classroom		x			x		X			
	Bathroom		x			x		X			
	Closet Door		x			x		X			
	Other (Describe)		x			x		X			
Windows	Frame/Sills/Sash/Curtains		x		x			X			
Bookcases	<i>all sides, top, bottom</i>	x			x			X		<b>Bookcase to be cleaned</b>	<b>#2 Bookcases Cleaned</b>
File Cabinets		x			x			X		<b>File Cabinets to be Cleaned</b>	<b>#2 File Cabinet Cleaned</b>
Inside Closets			x		x			X			
Bulletin Boards	<i>Check Behind</i>		X			x		X			
Chalkboards	<i>Check Behind</i>		x			x		X			
White Boards	<i>Check Behind</i>		X			x		X			
Wallpaper	<i>Check Behind</i>		x			x		X			
Wall Artwork	<i>Check Behind</i>		x			x		X			
Books/Magazines Etc.			x			x		X			
Room Contents	<i>Games Easels, etc.</i>		x		x			X			
Desks	<i>all sides, top, bottom</i>		x			x		X			
Chairs	<i>all sides, top, bottom</i>		x			x		X			
HVAC system	<i>Supply/Return/Filters/Ducts</i>	x			x			X		<b>Filters/Ducts to be Cleaned</b>	<b>#2 Vents/Ducts Cleaned</b>
Unit Ventilators	<i>Filter/Cage/Cover</i>	x			x			X		<b>Filters/Covers to be Cleaned</b>	<b>#2 Vents/Covers Cleaned</b>
Equipment	<i>all sides, top, bottom</i>		x			x		X			
Clothes			x			x		X			
Boxes			x			x		X			
Backpacks, shoes,			x			x		X			
Leather goods			x			x		X			
Shelves			X			x		X			
Under Sinks/Cabinets			x			x		X			



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Classroom S25	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	1st Grade Classroom											
Date:	8/22/2018											
Time:	1450											
Assessor:	L. Johnson III & T. Ranadive											
Room Component		Fungal Growth		Qty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	East		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	South		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	West		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
Ceiling	Tiles		X	24 SF	X			X	X		3 - 2' x 4' Ceiling Tiles	#1 Disposal of (3) Ceiling Tiles
	Above Ceiling	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Pipes/Insulation/Etc.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Drip Pans	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Lighting		X	16 SF	N/A	X		X		X	Cleaning of Light Fixtures Near Removed Ceiling Tiles	#2 Light Fixtures Cleaned
	Insulation	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Floor	Carpet Front	X		100 SF		X		X	X		1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Carpet Back	X		100 SF		X		X		X	1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Tiles		X	900 SF		X		X		X	Floors to be Cleaned Throughout	#2 Floor Cleaned
Doors	Classroom Door		X	32 SF		X		X	X		Entry Door to be Cleaned	#2 Door Cleaned
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Bathroom Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Door Frames	Classroom		X	12 SF		X		X	X		Entry Door Frame to be Cleaned	#2 Door Frame Cleaned
	Bathroom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other (Describe)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Windows	Frame/Sills/Sash/Curtains		X	24 SF		X		X	X		Frames/Sills to be Cleaned	#2 All Window Frames/Sills Cleaned
Bookcases	all sides, top, bottom		X	120 SF		X		X	X		Bookcase to be cleaned	#2 Bookcases Cleaned
File Cabinets		X		100 SF		X		X	X		File Cabinets to be Cleaned	#2 File Cabinet Cleaned
Inside Closets			X	60 SF		X		X	X		Closet Interiors to be Cleaned	#2 Closet Interiors Cleaned
Bulletin Boards	Check Behind		X	120 SF		X		X	X		Bulletin Boards to be Cleaned or Disposed	#2 Bulletin Board Cleaned
Chalkboards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
White Boards	Check Behind		X	60 SF		X		X	X		White Boards to be Cleaned	#2 White Boards Cleaned
Wallpaper	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wall Artwork	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Books/Magazines Etc.		X				X		X	X		Books Etc. to be Cleaned or Disposed	#1 & #2 Books Etc. Cleaned/Disposed
Room Contents	Games Easels, etc.	X				X		X	X		Games, Books Misc. items (Tennis Balls)	#1 & #2 Items Cleaned/Disposed & Disposed of Tennis Balls & Books
Desks	all sides, top, bottom	X				X		X	X		Multiple Desks Tops/Bottoms	#1 & #2 Tops/Bottoms of Desks Cleaned/Disposed
Chairs	all sides, top, bottom	X				X		X	X		Multiple Chairs	#1 & #2 Chairs Cleaned/Disposed - 1 Chair Disposed of
HVAC system	Supply/Return/Filters/Ducts	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Unit Ventilators	Filter/Cage/Cover		X			X		X		X	Filters/Covers to be Cleaned	#2 Vents/Covers Cleaned
Equipment	all sides, top, bottom		X			X		X		X	Equipment to be cleaned	#2 Equipment Cleaned
Clothes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Boxes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Backpacks, shoes,		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Leather goods		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Shelves			X	160 SF		X		X	X		Shelves to be Cleaned	#2 Shelves Cleaned
Under Sinks/Cabinets			X	30 SF		X		X		X	Sink & Counters/Components to be Cleaned	#2 Sink & Counters/Components to be Cleaned

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	S 13	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/23/2018											
Time:	930											
Assessor:	Frank Manna & Michael Smith											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No		
Walls	North	x		4 SF		x		X			<b>On Wall</b>	<b>#1 Dispose of 4 SF of Sheetrock</b>
	East		x			x		X				
	South	x		20 SF		x		X			<b>Under Smart Board</b>	<b>#2 Clean &amp; #3 Encapsulate Wall</b>
	West		x			x		X				
Ceiling	Tiles		x		x			X			<b>(4) 2' x 4' Ceiling Tiles</b>	<b>#1 Dispose of Ceiling Tiles</b>
	Above Ceiling		x			x		X				
	Pipes/Insulation/Etc.		x			x		X				
	Drip Pans		x			x		X				
	Lighting		x			x		X				
	Insulation		x			x		X				
	Other		x			x		X				
Floor	Carpet Front		x			x		X				
	Carpet Back		x			x		X				
	Tiles		x			x		X				
Doors	Classroom Door		x			x		X				
	Closet Door		x			x		X				
	Bathroom Door		x			x		X				
Door Frames	Classroom		x			x		X				
	Bathroom		x			x		X				
	Closet Door		x			x		X				
	Other (Describe)		x			x		X				
Windows	Frame/Sills/Sash/Curtains	x			x			X			<b>Window System</b>	<b>#2 Clean Complete Window System</b>
Bookcases	<i>all sides, top, bottom</i>		x		x			X				
File Cabinets			x		x			X			<b>Top of File Cabinet</b>	<b>#2 Clean File Cabinet</b>
Inside Closets			x			x		X				
Bulletin Boards	<i>Check Behind</i>	x				x		X			<b>On &amp; Behind Bulletin Board</b>	<b>#2 Clean Bulletin Board</b>
Chalkboards	<i>Check Behind</i>		x			x		X				
White Boards	<i>Check Behind</i>		x			x		X				
Wallpaper	<i>Check Behind</i>		x			x		X				
Wall Artwork	<i>Check Behind</i>		x			x		X				
Books/Magazines Etc.			x			x		X				
Room Contents	<i>Games Easels, etc.</i>		x			x		X				
Desks	<i>all sides, top, bottom</i>		x			x		X				
Chairs	<i>all sides, top, bottom</i>		x			x		X			<b>Tennis Balls on Chairs</b>	<b>#1 Dispose of Tennis Balls</b>
HVAC system	<i>Supply/Return/Filters/Ducts</i>		x			x		X				
Unit Ventilators	<i>Filter/Cage/Cover</i>		x			x		X				
Equipment	<i>all sides, top, bottom</i>		x			x		X				
Clothes			x			x		X				
Boxes			x			x		X				
Backpacks, shoes,			x			x		X				
Leather goods			x			x		X				
Shelves			x			x		X				
Under Sinks/Cabinets			x			x		X				





























School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Classroom S23	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	1st Grade Classroom											
Date:	8/22/2018											
Time:	1540											
Assessor:	L. Johnson III & T. Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	East		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	South		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	West		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
Ceiling	Tiles	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Above Ceiling	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Pipes/Insulation/Etc.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Drip Pans	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Lighting	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Insulation	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Floor	Carpet Front	X		100 SF		X		X	X		1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Carpet Back	X		100 SF		X		X		X	1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Tiles		X	900 SF		X		X		X	Floors to be Cleaned Throughout	#2 Floor Cleaned
Doors	Classroom Door		X	32 SF		X		X	X		Entry Door to be Cleaned	#2 Door Cleaned
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Bathroom Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Door Frames	Classroom		X	12 SF		X		X	X		Entry Door Frame to be Cleaned	#2 Door Frame Cleaned
	Bathroom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other (Describe)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Windows	Frame/Sills/Sash/Curtains		X	24 SF		X		X	X		Frames/Sills to be Cleaned	#2 All Window Frames/Sills Cleaned
Bookcases	all sides, top, bottom		X	120 SF		X		X	X		Bookcase to be cleaned	#2 Bookcases Cleaned
File Cabinets		X		100 SF		X		X		X	File Cabinets to be Cleaned	#2 File Cabinet Cleaned
Inside Closets			X	60 SF		X		X	X		Closet Interiors to be Cleaned	#2 Closet Interiors Cleaned
Bulletin Boards	Check Behind		X	120 SF		X		X	X		Bulletin Boards to be Cleaned or Disposed	#2 Bulletin Board Cleaned
Chalkboards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
White Boards	Check Behind		X	60 SF		X		X	X		White Boards to be Cleaned	#2 White Boards Cleaned
Wallpaper	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wall Artwork	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Books/Magazines Etc.		X				X		X	X		Books Etc. to be Cleaned or Disposed	#1 & #2 Books Etc. Cleaned/Disposed
Room Contents	Games Easels, etc.	X				X		X	X		Games, Books Misc. items (Tennis Balls)	#1 & #2 Items Cleaned/Disposed & Disposed of Tennis Balls & Books
Desks	all sides, top, bottom	X				X		X	X		Multiple Desks Tops/Bottoms	#1 & #2 Tops/Bottoms of Desks Cleaned/Disposed
Chairs	all sides, top, bottom	X				X		X	X		Multiple Chairs	#1 & #2 Chairs Cleaned/Disposed - 1 Chair Disposed of
HVAC system	Supply/Return/Filters/Ducts	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Unit Ventilators	Filter/Cage/Cover		X			X		X		X	Filters/Covers to be Cleaned	#2 Vents/Covers Cleaned
Equipment	all sides, top, bottom		X			X		X		X	Equipment to be cleaned	#2 Equipment Cleaned
Clothes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Boxes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Backpacks, shoes,		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Leather goods		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Shelves			X	120 SF		X		X	X		Shelves to be Cleaned	#2 Shelves Cleaned
Under Sinks/Cabinets			X	30 SF		X		X		X	Sink & Counters/Components to be Cleaned	#2 Sink & Counters/Components to be Cleaned



School Name:	Pequanock Elementary	<b>Mold Assessment Field Documentation Sheet</b>										
Room #:	Classroom S24	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	1st Grade Classroom											
Date:	8/22/2018											
Time:	1622											
Assessor:	L. Johnson III & T. Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	East		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	South		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	West		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
Ceiling	Tiles	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Above Ceiling	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Pipes/Insulation/Etc.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Drip Pans	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Lighting	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Insulation	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Floor	Carpet Front	X		100 SF		X		X	X		1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Carpet Back	X		100 SF		X		X		X	1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Tiles		X	900 SF		X		X		X	Floors to be Cleaned Throughout	#2 Floor Cleaned
Doors	Classroom Door		X	32 SF		X		X	X		Entry Door to be Cleaned	#2 Door Cleaned
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Bathroom Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Door Frames	Classroom		X	12 SF		X		X	X		Entry Door Frame to be Cleaned	#2 Door Frame Cleaned
	Bathroom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other (Describe)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Windows	Frame/Sills/Sash/Curtains		X	24 SF		X		X	X		Frames/Sills to be Cleaned	#2 All Window Frames/Sills Cleaned
Bookcases	all sides, top, bottom		X	120 SF		X		X	X		Bookcase to be cleaned	#2 Bookcases Cleaned
File Cabinets		X		100 SF		X		X		X	File Cabinets to be Cleaned	#2 File Cabinet Cleaned
Inside Closets			X	60 SF		X		X	X		Closet Interiors to be Cleaned	#2 Closet Interiors Cleaned
Bulletin Boards	Check Behind		X	120 SF		X		X	X		Bulletin Boards to be Cleaned or Disposed	#2 Bulletin Board Cleaned
Chalkboards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
White Boards	Check Behind		X	60 SF		X		X	X		White Boards to be Cleaned	#2 White Boards Cleaned
Wallpaper	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wall Artwork	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Books/Magazines Etc.		X				X		X	X		Books Etc. to be Cleaned or Disposed	#1 & #2 Books Etc. Cleaned/Disposed
Room Contents	Games Easels, etc.	X				X		X	X		Games, Books Misc. items (Tennis Balls)	#1 & #2 Items Cleaned/Disposed & Disposed of Tennis Balls & Books
Desks	all sides, top, bottom	X				X		X	X		Multiple Desks Tops/Bottoms	#1 & #2 Tops/Bottoms of Desks Cleaned/Disposed of 4- Desks
Chairs	all sides, top, bottom	X				X		X	X		Multiple Chairs	#1 & #2 Chairs Cleaned/Disposed
HVAC system	Supply/Return/Filters/Ducts	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Unit Ventilators	Filter/Cage/Cover		X			X		X		X	Filters/Covers to be Cleaned	#2 Vents/Covers Cleaned
Equipment	all sides, top, bottom		X			X		X		X	Equipment to be Cleaned	#2 Equipment Cleaned
Clothes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Boxes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Backpacks, shoes,		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Leather goods		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Shelves			X	120 SF		X		X	X		Shelves to be Cleaned	#2 Shelves Cleaned
Under Sinks/Cabinets			X	30 SF		X		X		X	Sink & Counters/Components to be Cleaned	#2 Sink & Counters/Components to be Cleaned

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Classroom S26	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	1st Grade Classroom											
Date:	8/24/2018											
Time:	1540											
Assessor:	L. Johnson III & T. Ranadive											
Room Component		Fungal Growth		Qty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
	East	X		30 SF		X		X	X		Behind Covebase Molding	#1 Disposal of 30 SF of Sheetrock
	South	X		20 SF		X		X	X		Behind Covebase Molding	#1 Disposal of 20 SF of Sheetrock
	West		X			X		X	X		Walls to be Cleaned	#2 Walls Cleaned
Ceiling	Tiles		X	16 SF	X			X	X		2 - 2' x 4' Ceiling Tiles	#1 Disposal of (2) Ceiling Tiles
	Above Ceiling	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Pipes/Insulation/Etc.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Drip Pans	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Lighting		X	8 SF	N/A	X		X		X	Cleaning of Light Fixtures Near Removed Ceiling Tiles	#2 Light Fixtures Cleaned
	Insulation	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Floor	Carpet Front	X		100 SF		X		X	X		1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Carpet Back	X		100 SF		X		X		X	1 Carpet	#1 & #2 Carpet Sent to be Cleaned/Disposed
	Tiles		X	900 SF		X		X		X	Floors to be Cleaned Throughout	#2 Floor Cleaned
Doors	Classroom Door		X	32 SF		X		X	X		Entry Door to be Cleaned	#2 Door Cleaned
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Bathroom Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Door Frames	Classroom		X	12 SF		X		X	X		Entry Door Frame to be Cleaned	#2 Door Frame Cleaned
	Bathroom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other (Describe)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Windows	Frame/Sills/Sash/Curtains		X	24 SF		X		X	X		Frames/Sills to be Cleaned	#2 All Window Frames/Sills Cleaned
Bookcases	all sides, top, bottom		X	120 SF		X		X	X		Bookcase to be cleaned	#2 Bookcases Cleaned
File Cabinets		X		100 SF		X		X		X	File Cabinets to be Cleaned	#2 File Cabinet Cleaned
Inside Closets			X	60 SF		X		X	X		Closet Interiors to be Cleaned	#2 Closet Interiors Cleaned
Bulletin Boards	Check Behind		X	120 SF		X		X	X		Bulletin Boards to be Cleaned or Disposed	#2 Bulletin Board Cleaned
Chalkboards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
White Boards	Check Behind		X	60 SF		X		X	X		White Boards to be Cleaned	#2 White Boards Cleaned
Wallpaper	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wall Artwork	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Books/Magazines Etc.		X				X		X	X		Books Etc. to be Cleaned or Disposed	#1 & #2 Books Etc. Cleaned/Disposed
Room Contents	Games Easels, etc.	X				X		X	X		Games, Books Misc. items (Tennis Balls)	#1 & #2 Items Cleaned/Disposed & Disposed of Tennis Balls & Books
Desks	all sides, top, bottom	X				X		X	X		Multiple Desks Tops/Bottoms	#1 & #2 Tops/Bottoms of Desks Cleaned/Disposed
Chairs	all sides, top, bottom	X				X		X	X		Multiple Chairs	#1 & #2 Chairs Cleaned/Disposed - 1 Chair Disposed of
HVAC system	Supply/Return/Filters/Ducts	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Unit Ventilators	Filter/Cage/Cover		X			X		X		X	Filters/Covers to be Cleaned	#2 Vents/Covers Cleaned
Equipment	all sides, top, bottom		X			X		X		X	Equipment to be cleaned	#2 Equipment Cleaned
Clothes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Boxes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Backpacks, shoes,		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Leather goods		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Shelves			X	120 SF		X		X	X		Shelves to be Cleaned	#2 Shelves Cleaned
Under Sinks/Cabinets			X	30 SF		X		X		X	Sink & Counters/Components to be Cleaned	#2 Sink & Counters/Components to be Cleaned

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Custodial Closet	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Closet Door											
Date:	8/22/2018											
Time:	1520											
Assessor:	Louis N. Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		4SF		X		X	X		Behind Covebase Molding	#1 Disposal of 4 SF of Sheetrock Wall
	East	X		10 SF		X		X	X		Behind Covebase Molding	#1 Disposal of 10 SF of Sheetrock Wall
	South	X		4 SF		X		X	X		Behind Covebase Molding	#1 Disposal of 4 SF of Sheetrock Wall
	West	X		10 SF		X		X	X		Behind Covebase Molding	#1 Disposal of 10 SF of Sheetrock Wall
Ceiling	Tiles	X		8 SF		X		X	X		1 - 2' x 4' Ceiling Tile	#1 Disposal of (1) Ceiling Tile
	Above Ceiling	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Pipes/Insulation/Etc.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Drip Pans	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Lighting	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Insulation	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Floor	Carpet Front	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Carpet Back	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Tiles		X	30 SF		X		X		X	Floors to be Cleaned Throughout	#2 Floor Cleaned
Doors	Door	X		32 SF		X		X	X		Door to be Cleaned	#2 Door Cleaned
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Bathroom Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Door Frames	Door		X	10 SF		X		X	X		Door Frames to be Cleaned	#2 Door Frames Cleaned
	Bathroom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Closet Door	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
	Other (Describe)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Windows	Frame/Sills/Sash/Curtains	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Bookcases	all sides, top, bottom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
File Cabinets		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Inside Closets		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Bulletin Boards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Chalkboards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
White Boards	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wallpaper	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Wall Artwork	Check Behind	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Books/Magazines Etc.		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Room Contents	Games Easels, etc.	X				X		X	X		Misc. items to be Cleaned	#1 & #2 Items Cleaned/Disposed
Desks	all sides, top, bottom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Chairs	all sides, top, bottom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
HVAC system	Supply/Return/Filters/Ducts	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Unit Ventilators	Filter/Cage/Cover	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Equipment	all sides, top, bottom	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Clothes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Boxes		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Backpacks, shoes,		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Leather goods		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Shelves			X	100 SF		X		X	X		Shelves to be Cleaned	#2 Shelves Cleaned
Under Sinks/Cabinets			X	6 SF		X		X		X	Slop Sink to be Cleaned	#2 Slop Sink Cleaned









**Appendix D(1)**  
**Mold Assessment Documentation**  
**West Wing**

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	W-13	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Faculty											
Date:	8/23/2018											
Time:	1430											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X			#2 Clean, #3 Encapsulate
	East		X			X		X	X			#2 Clean, #3 Encapsulate
	South		X			X		X	X			#2 Clean, #3 Encapsulate
	West		X			X		X	X			#2 Clean, #3 Encapsulate
Ceiling	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate
	Above Ceiling		X			X		X		X		#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean, #3 Encapsulate
	Drip Pans		X			X		X		X		#2 Clean, #3 Encapsulate
	Lighting		X			X		X		X		#2 Clean, #3 Encapsulate
	Insulation		X			X		X		X		#2 Clean, #3 Encapsulate
	Other		X			X		X		X		#2 Clean, #3 Encapsulate
Floor	Carpet Front		X	30 SF	X			X	X			#1 Dispose of Carpet
	Carpet Back		X			X		X	X			#1 Dispose of Carpet
	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate
Doors	Classroom Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X		X		#2 Clean, #3 Encapsulate
	Bathroom		X			X		X		X		
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X		X		X		X	Window Components to be Cleaned		#2 Clean, #3 Encapsulate
Bookcases	all sides, top, bottom		X			X		X				#2 Clean, #3 Encapsulate
File Cabinets			X			X		X				#2 Clean, #3 Encapsulate
Inside Closets			X			X		X				#2 Clean, #3 Encapsulate
Bulletin Boards	Check Behind		X			X		X		X		
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X		X		
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		
Books/Magazines Etc.			X			X		X		X		#2 Clean, #3 Encapsulate
Room Contents	Games Easels, etc.		X			X		X		X		
Desks	all sides, top, bottom		X			X		X		X		#2 Clean
Chairs	all sides, top, bottom	X				X		X	X		Tennis Balls on Chairs	#1 Dispose of Tennis Balls, #2 Clean Chairs, #3 Encapsulate
HVAC system	Supply/Return/Filters/Ducts		X		X			X		X		#2 Clean
Unit Ventilators	Filter/Cage/Cover		X		X			X		X		#2 Clean
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean, #3 Encapsulate
Clothes			X			X		X		X		#2 Clean, #3 Encapsulate
Boxes			X			X		X		X		#2 Clean, #3 Encapsulate
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		
Shelves			X			X		X	X			
Under Sinks/Cabinets			X			X		X		X		























School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	W19	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Asst Principals Office											
Date:	8/23/2018											
Time:	1300											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No	#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North	x				x		X			By Entry Door/Endcap	#3 Encapsulate on Sheetrock
	East	x		20 SF		x		X			Entry Dividing Wall	#1 Dispose of 20 SF of Sheetrock
	South		x			x		X				
	West		x	10 SF	x			X			Near Entry Door	#3 Encapsulate 10 SF of Sheetrock Wall
Ceiling	Tiles		x			x		X				
	Above Ceiling		x			x		X				
	Pipes/Insulation/Etc.		x			x		X				
	Drip Pans		x			x		X				
	Lighting		x			x		X				
	Insulation		x			x		X				
	Other		x			x		X				
Floor	Carpet Front		x			x		X				
	Carpet Back		x			x		X				
	Tiles		x			x		X			Throughout Room	#2 Clean Floor Tiles
Doors	Classroom Door		x			x		X			Multiple Locations	#2 Clean Door
	Closet Door		x			x		X				
	Bathroom Door		x			x		X				
Door Frames	Classroom		x			x		X			Entire Frame	#2 Clean Door Frame
	Bathroom		x			x		X				
	Closet Door		x			x		X				
Back Cabinets	Other (Describe)		x			x		X				
Windows	Frame/Sills/Sash/Curtains		x		x		x		x		Window Sills	#2 Clean Window Sills
Bookcases	all sides, top, bottom		x			x		X			Throughout Bookshelf	#2 Clean Top, Side and Shelves of Bookcase
File Cabinets			x			x		X			Throughout File Cabinet	#2 Clean Filing Cabinet
Inside Closets			x			x		X				
Bulletin Boards	Check Behind		x			x		X				
Chalkboards	Check Behind		x			x		X				
White Boards	Check Behind		x			x		X				
Wallpaper	Check Behind		x			x		X				
Wall Artwork	Check Behind		x			x		X			2 Picture Frames	#2 Clean Picture Frames
Books/Magazines Etc.			x			x		X			Multiple Books & Magazines	#2 Clean Books & Magazines
Room Contents	Games Easels, etc.		x			x		X				
Desks	all sides, top, bottom		x			x		X			Multiple Locations	#2 Clean All Desks
Chairs	all sides, top, bottom		x			x		X			Multiple Locations	#2 Clean All Chairs
HVAC system	Supply/Return/Filters/Ducts		x		x		x		x			
Unit Ventilators	Filter/Cage/Cover		x		x		x		x		Multiple Locations	#2 Clean All Univentilator System
Equipment	all sides, top, bottom		x			x		X			All Sides	#2 Clean All Equipment
Clothes			x			x		X				
Boxes			x			x		X				
Backpacks, shoes,			x			x		X				
Leather goods			x			x		X				
Shelves			x			x		X			Multiple Locations	#2 Clean All Shelving
Under Sinks/Cabinets			x			x		X				

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	W21	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Nurses Office											
Date:	8/27/2018											
Time:	1300											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No	#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North	x		4 SF		x		X			Dispose of Sheetrock Wall	#1 Dispose of 4 SF of Sheetrock Wall
	East		x			x		X				
	South		x			x		X				
	West	x		15 SF				X			Dispose of Sheetrock Wall	#1 Dispose of 15 SF of Sheetrock
Ceiling	Tiles	x		24 SF	x			X			Near Entry & Center	#1 Dispose of (3) 2' x 4' Ceiling Tiles
	Above Ceiling		x			x		X				
	Pipes/Insulation/Etc.		x			x		X				
	Drip Pans		x			x		X				
	Lighting		x			x		X				
	Insulation		x			x		X				
	Other		x			x		X				
Floor	Carpet Front		x			x		X				
	Carpet Back		x			x		X				
	Tiles		x			x		X			Multiple Locations	# 2 Clean Entire Surface
Doors	Classroom Door		x			x		X			Multiple Locations	# 2 Clean Entire Surface
	Closet Door		x			x		X			Multiple Locations	# 2 Clean Entire Surface
	Bathroom Door		x			x		X			Multiple Locations	# 2 Clean Entire Surface
Door Frames	Classroom		x			x		X			Multiple Locations	# 2 Clean Entire Surface
	Bathroom		x			x		X			Multiple Locations	# 2 Clean Entire Surface
	Closet Door		x			x		X			Multiple Locations	# 2 Clean Entire Surface
Back Cabinets	Other (Describe)		x			x		X			Multiple Locations	# 2 Clean Entire Surface
Windows	Frame/Sills/Sash/Curtains		x			x		X			Multiple Locations	# 2 Clean Entire Surface
Bookcases	all sides, top, bottom		x			x		X			Multiple Locations	# 2 Clean Entire Surface
File Cabinets			x			x		X			Multiple Locations	# 2 Clean Entire Surface
Inside Closets			x			x		X			Multiple Locations	# 2 Clean Entire Surface
Bulletin Boards	Check Behind		x			x		X				
Chalkboards	Check Behind		x			x		X				
White Boards	Check Behind		x			x		X				
Wallpaper	Check Behind		x			x		X				
Wall Artwork	Check Behind		x			x		X			(2) Picture Frames	#2 Clean Picture Frame
Books/Magazines Etc.			x			x		X				
Room Contents	Games Easels, etc.		x			x		X				
Desks	all sides, top, bottom	x				x		X			Back Room, Multiple Desks	#2 Clean All Desks & Chairs
Chairs	all sides, top, bottom	x				x		X			Felt Chair, Refridgerator	#1 Dispose of these materials
HVAC system	Supply/Return/Filters/Ducts		x			x		X				
Unit Ventilators	Filter/Cage/Cover		x			x		X			Multiple Locations	#2 Clean All Unit Ventilators
Equipment	all sides, top, bottom		x			x		X			EMT Bags, Crutches, Disposal Bags, Clipboard	#1 Dispose of These Materials
Clothes			x			x		X				
Boxes			x			x		X				
Backpacks, shoes,			x			x		X				
Leather goods			x			x		X				
Shelves			x			x		X			Multiple Locations	#2 Clean All Shelves
Under Sinks/Cabinets		x			x			X			Back Room	#2 Clean All Cabinets & Under Sink



**Appendix D(2)**  
**Mold Assessment Documentation**  
**Library Core Wing**

















School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet												
Room #:	C-22A	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.												
Room Type:	Head Custodial Office													
Date:	8/31/2018													
Time:	1330													
Assessor:	Tanay Ranadive													
										Response Action				
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate		
		Yes	No		Yes	No	Yes	No	Yes	No				
Walls	North		X			X		X		X		#2 Clean		
	East		X			X		X		X		#2 Clean		
	South		X			X		X		X		#2 Clean		
	West		X			X		X		X		#2 Clean		
Ceiling	Tiles		X			X		X		X		#2 Clean		
	Above Ceiling		X			X		X		X		#2 Clean		
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean		
	Drip Pans		X			X		X		X		#2 Clean		
	Lighting		X			X		X		X		#2 Clean		
	Insulation		X			X		X		X		#2 Clean		
	Other		X			X		X		X		#2 Clean		
				X			X		X		X		#2 Clean	
Floor	Carpet Front		X			X		X		X		#2 Clean		
	Carpet Back		X			X		X		X		#2 Clean		
	Tiles		X			X		X		X		#2 Clean		
Doors	Classroom Door		X			X		X		X		#2 Clean		
	Closet Door		X			X		X		X		#2 Clean		
	Bathroom Door		X			X		X		X		#2 Clean		
Door Frames	Classroom		X			X		X		X		#2 Clean		
	Bathroom		X			X		X		X		#2 Clean		
	Closet Door		X			X		X		X		#2 Clean		
	Other (Describe)		X			X		X		X		#2 Clean		
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		#2 Clean		
Bookcases	all sides, top, bottom		X			X		X		X		#2 Clean		
File Cabinets			X			X		X		X		#2 Clean		
Inside Closets			X			X		X		X		#2 Clean		
Bulletin Boards	Check Behind		X			X		X		X		#2 Clean		
Chalkboards	Check Behind		X			X		X		X		#2 Clean		
White Boards	Check Behind		X			X		X		X		#2 Clean		
Wallpaper	Check Behind		X			X		X		X		#2 Clean		
Wall Artwork	Check Behind		X			X		X		X		#2 Clean		
Books/Magazines Etc.			X			X		X		X		#2 Clean		
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean		
Desks	all sides, top, bottom		X			X		X		X		#2 Clean		
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean		
HVAC system	Supply/Return/Filters/Ducts	X		10 SF	X		X			X	Around Vent Fins	#2 Clean		
Unit Ventilators	Filter/Cage/Cover	X		10 SF	X		X			X	Around Vent Fins and Metal Plate	#2 Clean		
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean		
Clothes			X			X		X		X		#2 Clean		
Boxes			X			X		X		X		#2 Clean		
Backpacks, shoes,			X			X		X		X		#2 Clean		
Leather goods			X			X		X		X		#2 Clean		
Shelves			X			X		X		X		#2 Clean		
Under Sinks/Cabinets			X			X		X		X		#2 Clean		

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C-22B	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Storage Room											
Date:	8/31/2018											
Time:	1400											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X		X		#2 Clean
	East		X			X		X		X		#2 Clean
	South		X			X		X		X		#2 Clean
	West		X			X		X		X		#2 Clean
Ceiling	Tiles		X			X		X		X		#2 Clean
	Above Ceiling		X			X		X		X		#2 Clean
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean
	Drip Pans		X			X		X		X		#2 Clean
	Lighting		X			X		X		X		#2 Clean
	Insulation		X			X		X		X		#2 Clean
	Other		X			X		X		X		#2 Clean
Floor	Carpet Front		X			X		X		X		#2 Clean
	Carpet Back		X			X		X		X		#2 Clean
	Tiles		X			X		X		X		#2 Clean
Doors	Classroom Door		X			X		X		X		#2 Clean
	Closet Door		X			X		X		X		#2 Clean
	Bathroom Door		X			X		X		X		#2 Clean
Door Frames	Classroom		X			X		X		X		#2 Clean
	Bathroom		X			X		X		X		#2 Clean
	Closet Door		X			X		X		X		#2 Clean
	Other (Describe)		X			X		X		X		#2 Clean
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		#2 Clean
Bookcases	all sides, top, bottom		X			X		X		X		#2 Clean
File Cabinets			X			X		X		X		#2 Clean
Inside Closets			X			X		X		X		#2 Clean
Bulletin Boards	Check Behind		X			X		X		X		#2 Clean
Chalkboards	Check Behind		X			X		X		X		#2 Clean
White Boards	Check Behind		X			X		X		X		#2 Clean
Wallpaper	Check Behind		X			X		X		X		#2 Clean
Wall Artwork	Check Behind		X			X		X		X		#2 Clean
Books/Magazines Etc.			X			X		X		X		#2 Clean
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean
Desks	all sides, top, bottom		X			X		X		X		#2 Clean
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		#2 Clean
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		#2 Clean
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean
Clothes			X			X		X		X		#2 Clean
Boxes			X			X		X		X		#2 Clean
Backpacks, shoes,			X			X		X		X		#2 Clean
Leather goods			X			X		X		X		#2 Clean
Shelves			X			X		X		X		#2 Clean
Under Sinks/Cabinets			X			X		X		X		#2 Clean



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C26	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Library Hallway											
Date:	9/1/2018											
Time:	1400											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X		X		#2 Clean, #3 Encapsulate
	East		X			X		X		X		#2 Clean, #3 Encapsulate
	South		X			X		X		X		#2 Clean, #3 Encapsulate
	West		X			X		X		X		#2 Clean, #3 Encapsulate
Ceiling	Tiles		X			X		X		X		#1 Dispose of all Ceiling Tiles
	Above Ceiling		X			X		X		X		#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.	X		60 SF	X			X	X		Pipe Insulation to be Disposed	#1 Pipe Insulation and Duct Insulation Disposed
	Drip Pans		X			X		X		X		#2 Clean, #3 Encapsulate
	Lighting		X			X		X		X		#2 Clean, #3 Encapsulate
	Insulation		X			X		X		X		#2 Clean, #3 Encapsulate
	Other		X			X		X		X		#2 Clean, #3 Encapsulate
Floor	Carpet Front		X			X		X	X			#2 Clean
	Carpet Back		X			X		X		X		
	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate
Doors	Classroom Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Closet Door		X			X		X		X		
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X		X		#2 Clean, #3 Encapsulate
	Bathroom		X			X		X		X		
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains	X				X		X		X	Window Components to be Cleaned	#2 Clean
Bookcases	all sides, top, bottom	X				X		X		X	Bookcase to be cleaned	#2 Bookcases Cleaned
File Cabinets			X			X		X		X	File Cabinets to be Cleaned	#2 File Cabinet Cleaned
Inside Closets			X			X		X		X		
Bulletin Boards	Check Behind		X			X		X		X		
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X		X		
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		
Books/Magazines Etc.		X		30 Books	X			X	X		Books to Be Cleaned and Disposed of	#1 Dispose of Books, #2 Clean Books
Room Contents	Games Easels, etc.	X				X		X		X	Table, American Flag, and Tent to be Disposed of	#1 Dispose Contents
Desks	all sides, top, bottom		X			X		X		X		
Chairs	all sides, top, bottom		X			X		X		X		
HVAC system	Supply/Return/Filters/Ducts		X		X			X		X		#2 Vents/Ducts Cleaned
Unit Ventilators	Filter/Cage/Cover		X		X			X		X		#2 Vents/Covers Cleaned
Equipment	all sides, top, bottom		X			X		X		X		
Clothes			X			X		X		X		
Boxes			X			X		X		X		
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		#1 Dispose Contents
Shelves		X			X			X	X		Cove Base of Bookshelf by Speck Room 10	#1 Dispose Contents, #2 Clean, #3 Encapsulate
Under Sinks/Cabinets			X			X		X		X		

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C-28	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Cafeteria											
Date:	8/31/2018											
Time:	1030											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X			#2 Clean, #3 Encapsulate
	East		X			X		X	X			#2 Clean, #3 Encapsulate
	South		X			X		X	X			#2 Clean, #3 Encapsulate
	West		X			X		X	X			#2 Clean, #3 Encapsulate
Ceiling	Tiles		X			X		X	X			#2 Clean, #3 Encapsulate
	Above Ceiling		X			X		X	X			#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.		X			X		X	X			#2 Clean, #3 Encapsulate
	Drip Pans		X			X		X	X			#2 Clean, #3 Encapsulate
	Lighting		X			X			X			#2 Clean, #3 Encapsulate
	Insulation		X			X			X			
	Other		X			X			X			#2 Clean, #3 Encapsulate
Floor	Carpet Front		X			X			X			#2 Clean, #3 Encapsulate
	Carpet Back		X			X			X			#2 Clean, #3 Encapsulate
	Tiles		X			X			X			#2 Clean, #3 Encapsulate
Doors	Classroom Door		X			X		X	X			#2 Clean, #3 Encapsulate
	Closet Door		X			X		X	X			#2 Clean, #3 Encapsulate
	Bathroom Door		X			X			X			
Door Frames	Classroom		X			X			X			#2 Clean, #3 Encapsulate
	Bathroom		X			X			X			
	Closet Door		X			X			X			#2 Clean, #3 Encapsulate
	Other (Describe)		X			X			X			
Windows	Frame/Sills/Sash/Curtains		X			X			X			
Bookcases	all sides, top, bottom		X			X			X			
File Cabinets			X			X			X			
Inside Closets			X			X			X			
Bulletin Boards	Check Behind		X			X			X			
Chalkboards	Check Behind		X			X			X			
White Boards	Check Behind		X			X			X			
Wallpaper	Check Behind		X			X			X			
Wall Artwork	Check Behind	X				X			X		Fruit Side Frames	#2 Clean Frames
Books/Magazines Etc.			X			X			X			
Room Contents	Games Easels, etc.		X			X			X			
Tables	all sides, top, bottom		X		X				X	X		#2 Clean Tables
Chairs	all sides, top, bottom		X			X			X			
HVAC system	Supply/Return/Filters/Ducts		X		X				X			
Unit Ventilators	Filter/Cage/Cover		X		X				X			
Equipment	all sides, top, bottom		X			X			X			
Clothes			X			X			X			
Boxes			X			X			X			
Backpacks, shoes,			X			X			X			
Leather goods			X			X			X			
Shelves			X			X			X			
Under Sinks/Cabinets			X			X			X			

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C-29	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	ELL Classroom											
Date:	8/31/2018											
Time:	1100											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X			#2 Clean, #3 Encapsulate
	East		X			X		X	X			#2 Clean, #3 Encapsulate
	South		X			X		X	X			#2 Clean, #3 Encapsulate
	West		X			X		X	X			#2 Clean, #3 Encapsulate
Ceiling	Tiles	X		36 SF	X		X		X		Area Underneath HVAC Unit	#1 Dispose of Ceiling Tiles
	Above Ceiling		X			X		X		X		#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.	X		10 LF	X		X		X		HVAC Pipes	#1 Dispose of Pipe Insulation
	Drip Pans		X		X			X		X		#2 Clean, #3 Encapsulate
	Lighting		X		X			X		X		
Floor	Insulation		X		X			X	X			
	Other		X			X		X		X		
	Carpet Front		X			X		X		X		
	Carpet Back		X			X		X		X		
	Tiles		X			X		X		X		
Doors	Classroom Door		X			X		X		X		
	Closet Door		X			X		X		X		
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X		X		
	Bathroom		X			X		X		X		
	Closet Door		X			X		X		X		
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X		X			X		X	Window Sills to be Cleaned	#2 Clean Window Sills
Bookcases	all sides, top, bottom		X			X		X		X		
File Cabinets		X				X		X		X	Top Side of Cabinets to be Cleaned	#2 Clean File Cabinets
Inside Closets			X			X		X		X		
Bulletin Boards	Check Behind	X				X		X		X		#1 Dispose of Bulletin Board
Chalkboards	Check Behind		X			X		X		X		#2 Clean, #3 Encapsulate
White Boards	Check Behind		X			X		X		X		#2 Clean, #3 Encapsulate
Wallpaper	Check Behind		X			X		X		X		#2 Clean, #3 Encapsulate
Wall Artwork	Check Behind		X			X		X		X		#2 Clean, #3 Encapsulate
Books/Magazines Etc.			X			X		X		X		#2 Clean, #3 Encapsulate
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean, #3 Encapsulate
Desks	all sides, top, bottom	X				X		X		X		#2 Clean Desks
Chairs	all sides, top, bottom	X				X		X		X	1 Chair	#1 Dispose of Chair
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		#2 Vents/Ducts Cleaned, #3 Encapsulated
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		#2 Vents/Ducts Cleaned, #3 Encapsulated
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean, #3 Encapsulate
Clothes			X			X		X		X		#2 Clean, #3 Encapsulate
Boxes			X			X		X		X		#2 Clean, #3 Encapsulate
Backpacks, shoes,			X			X		X		X		#2 Clean, #3 Encapsulate
Leather goods			X			X		X		X		#2 Clean, #3 Encapsulate
Shelves			X			X		X		X		#2 Clean, #3 Encapsulate
Under Sinks/Cabinets			X			X		X		X		#2 Clean, #3 Encapsulate





School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet												
Room #:	C-31	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.												
Room Type:	Art Room													
Date:	8/30/2018													
Time:	1300													
Assessor:	Tanay Ranadive													
										Response Action				
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate		
		Yes	No		Yes	No	Yes	No	Yes	No				
Walls	North		X			X		X		X		#2 Clean, #3 Encapsulate		
	East		X			X		X		X		#2 Clean, #3 Encapsulate		
	South		X			X		X		X		#2 Clean, #3 Encapsulate		
	West		X			X		X		X		#2 Clean, #3 Encapsulate		
Ceiling	Tiles		X		X			X		X	2 Above Exhaust Hood	#1 Dispose of Ceiling Tiles		
	Above Ceiling		X			X		X	X			#2 Clean, #3 Encapsulate		
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean, #3 Encapsulate		
	Drip Pans		X			X		X		X		#2 Clean, #3 Encapsulate		
	Lighting		X			X		X		X		#2 Clean, #3 Encapsulate		
	Insulation		X			X		X		X		#2 Clean, #3 Encapsulate		
	Other		X			X		X		X				
Floor	Carpet Front		X			X		X		X				
	Carpet Back		X			X		X		X				
	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate		
Doors	Classroom Door		X		X			X	X			#2 Clean, #3 Encapsulate		
	Closet Door		X			X		X	X			#2 Clean, #3 Encapsulate		
	Bathroom Door		X			X		X		X				
Door Frames	Classroom		X			X		X		X		#2 Clean, #3 Encapsulate		
	Bathroom		X			X		X		X				
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate		
	Other (Describe)		X			X		X		X				
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		#2 Clean, #3 Encapsulate		
Bookcases	all sides, top, bottom	X			X			X		X	Base of Bookcase Along West Wall	#2 Clean Bookcase		
File Cabinets			X		X			X		X	Base of Doors Along West Wall	#2 Clean, #3 Encapsulate		
Inside Closets			X			X		X	X			#2 Clean, #3 Encapsulate		
Bulletin Boards	Check Behind	X		80 SF		X		X		X	Northwest Corner	#1 Dispose of Bulletin Board		
Chalkboards	Check Behind		X			X		X	X			#2 Clean, #3 Encapsulate		
White Boards	Check Behind		X			X		X	X			#2 Clean, #3 Encapsulate		
Wallpaper	Check Behind		X			X		X	X			#2 Clean, #3 Encapsulate		
Wall Artwork	Check Behind		X			X		X	X			#2 Clean, #3 Encapsulate		
Books/Magazines Etc.			X			X		X	X			#2 Clean, #3 Encapsulate		
Room Contents	Games Easels, etc.		X			X		X	X			#2 Clean, #3 Encapsulate		
Desks	all sides, top, bottom	X				X		X	X		Sides	#2 Clean Desks		
Chairs	all sides, top, bottom		X			X		X	X			#2 Clean, #3 Encapsulate		
HVAC system	Supply/Return/Filters/Ducts		X		X			X	X			#2 Clean, #3 Encapsulate		
Unit Ventilators	Filter/Cage/Cover		X			X		X	X			#2 Clean, #3 Encapsulate		
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean, #3 Encapsulate		
Clothes			X			X		X		X		#2 Clean, #3 Encapsulate		
Boxes		X			X			X	X		Cardboard Box in Northeast Corner	#1 Dispose of Boxes		
Backpacks, shoes,			X			X		X		X		#2 Clean, #3 Encapsulate		
Leather goods			X			X		X		X		#2 Clean, #3 Encapsulate		
Shelves			X			X		X		X		#2 Clean, #3 Encapsulate		
Under Sinks/Cabinets			X			X		X		X		#2 Clean, #3 Encapsulate		

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet												
Room #:	C-32	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.												
Room Type:	Storage Room													
Date:	9/1/2018													
Time:	1000													
Assessor:	Tanay Ranadive													
										Response Action				
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate		
		Yes	No		Yes	No	Yes	No	Yes	No				
Walls	North		X			X		X		X		#2 Clean		
	East		X			X		X		X		#2 Clean		
	South		X			X		X		X		#2 Clean		
	West		X			X		X		X		#2 Clean		
Ceiling	Tiles		X			X		X		X		#2 Clean		
	Above Ceiling		X			X		X		X		#2 Clean		
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean		
	Drip Pans		X			X		X		X		#2 Clean		
	Lighting		X			X		X		X		#2 Clean		
	Insulation		X			X		X		X		#2 Clean		
	Other		X			X		X		X		#2 Clean		
Floor	Carpet Front		X			X		X		X		#2 Clean		
	Carpet Back		X			X		X		X		#2 Clean		
	Tiles		X			X		X		X		#2 Clean		
Doors	Classroom Door		X			X		X		X		#2 Clean		
	Closet Door		X			X		X		X		#2 Clean		
	Bathroom Door		X			X		X		X		#2 Clean		
Door Frames	Classroom		X			X		X		X		#2 Clean		
	Bathroom		X			X		X		X		#2 Clean		
	Closet Door		X			X		X		X		#2 Clean		
	Other (Describe)		X			X		X		X		#2 Clean		
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		#2 Clean		
Bookcases	all sides, top, bottom		X			X		X		X		#2 Clean		
File Cabinets			X			X		X		X		#2 Clean		
Inside Closets			X			X		X		X		#2 Clean		
Bulletin Boards	Check Behind		X			X		X		X		#2 Clean		
Chalkboards	Check Behind		X			X		X		X		#2 Clean		
White Boards	Check Behind		X			X		X		X		#2 Clean		
Wallpaper	Check Behind		X			X		X		X		#2 Clean		
Wall Artwork	Check Behind		X			X		X		X		#2 Clean		
Books/Magazines Etc.			X			X		X		X		#2 Clean		
Room Contents	Games Easels, etc.	X		4 SF		X		X		X	Small Wood Board	#1 Dispose of 4 SF of Wood Board		
Desks	all sides, top, bottom		X			X		X		X		#2 Clean		
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean		
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		#2 Clean		
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		#2 Clean		
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean		
Clothes			X			X		X		X		#2 Clean		
Boxes			X			X		X		X		#2 Clean		
Backpacks, shoes,			X			X		X		X		#2 Clean		
Leather goods			X			X		X		X		#2 Clean		
Shelves			X			X		X		X		#2 Clean		
Under Sinks/Cabinets			X			X		X		X		#2 Clean		



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C-35	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	MER											
Date:	8/31/2018											
Time:	1500											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X		X		#2 Clean
	East		X			X		X		X		#2 Clean
	South		X			X		X		X		#2 Clean
	West		X			X		X		X		#2 Clean
Ceiling	Tiles		X			X		X		X		#2 Clean
	Above Ceiling		X			X		X		X		#2 Clean
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean
	Drip Pans		X			X		X		X		#2 Clean
	Lighting		X			X		X		X		#2 Clean
	Insulation		X			X		X		X		#2 Clean
	Other		X			X		X		X		#2 Clean
Floor	Carpet Front		X			X		X		X		#2 Clean
	Carpet Back		X			X		X		X		#2 Clean
	Tiles		X			X		X		X		#2 Clean
Doors	Classroom Door		X			X		X		X		#2 Clean
	Closet Door		X			X		X		X		#2 Clean
	Bathroom Door		X			X		X		X		#2 Clean
Door Frames	Classroom		X			X		X		X		#2 Clean
	Bathroom		X			X		X		X		#2 Clean
	Closet Door		X			X		X		X		#2 Clean
	Other (Describe)		X			X		X		X		#2 Clean
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		#2 Clean
Bookcases	all sides, top, bottom		X			X		X		X		#2 Clean
File Cabinets			X			X		X		X		#2 Clean
Inside Closets			X			X		X		X		#2 Clean
Bulletin Boards	Check Behind		X			X		X		X		#2 Clean
Chalkboards	Check Behind		X			X		X		X		#2 Clean
White Boards	Check Behind		X			X		X		X		#2 Clean
Wallpaper	Check Behind		X			X		X		X		#2 Clean
Wall Artwork	Check Behind		X			X		X		X		#2 Clean
Books/Magazines Etc.			X			X		X		X		#2 Clean
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean
Desks	all sides, top, bottom		X			X		X		X		#2 Clean
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean
HVAC system	Supply/Return/Filters/Ducts	X			X			X		X		#2 Clean
Unit Ventilators	Filter/Cage/Cover	X			X			X		X		#2 Clean
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean
Clothes			X			X		X		X		#2 Clean
Boxes			X			X		X		X		#2 Clean
Backpacks, shoes,			X			X		X		X		#2 Clean
Leather goods			X			X		X		X		#2 Clean
Shelves			X			X		X		X		#2 Clean
Under Sinks/Cabinets			X			X		X		X		#2 Clean



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	N/A	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Bathroom Near C29											
Date:	8/31/2018											
Time:	1034											
Assessor:	Tanay Ranadive											
										Response Action		
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate
		Yes	No		Yes	No	Yes	No	Yes	No		
Walls	North		X			X		X		X		#2 Clean, #3 Encapsulate
	East		X			X		X		X		#2 Clean, #3 Encapsulate
	South		X			X		X		X		#2 Clean, #3 Encapsulate
	West		X			X		X		X		#2 Clean, #3 Encapsulate
Ceiling	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate
	Above Ceiling		X			X		X		X		#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.		X			X		X		X		#2 Clean, #3 Encapsulate
	Drip Pans		X			X		X		X		#2 Clean, #3 Encapsulate
	Lighting		X			X		X		X		#2 Clean, #3 Encapsulate
	Insulation		X			X		X		X		#2 Clean, #3 Encapsulate
	Other		X			X		X		X		#2 Clean, #3 Encapsulate
Floor	Carpet Front		X			X		X		X		#2 Clean, #3 Encapsulate
	Carpet Back		X			X		X		X		#2 Clean, #3 Encapsulate
	Tiles		X		X			X	X		Custodial Closet	#2 Clean
Doors	Classroom Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Bathroom Door		X			X		X		X		#2 Clean, #3 Encapsulate
Door Frames	Classroom		X			X		X		X		#2 Clean, #3 Encapsulate
	Bathroom		X			X		X		X		#2 Clean, #3 Encapsulate
	Closet Door		X			X		X		X		#2 Clean, #3 Encapsulate
	Other (Describe)		X			X		X		X		#2 Clean, #3 Encapsulate
Windows	Frame/Sills/Sash/Curtains		X		X			X	X		Window Sills to Be Cleaned	#2 Clean
Bookcases	all sides, top, bottom		X			X		X		X		
File Cabinets			X			X		X		X		
Inside Closets			X			X		X		X		
Bulletin Boards	Check Behind		X			X		X		X		
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X		X		
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		
Books/Magazines Etc.			X			X		X		X		
Room Contents	Games Easels, etc.		X			X		X		X		
Desks	all sides, top, bottom		X			X		X		X		
Chairs	all sides, top, bottom		X			X		X		X		
HVAC system	Supply/Return/Filters/Ducts	X			X			X	X			#2 Vents/Ducts Cleaned, #3 Encapsulated
Unit Ventilators	Filter/Cage/Cover	X			X			X	X			#2 Vents/Covers Cleaned, #3 Encapsulated
Equipment	all sides, top, bottom		X			X		X	X			#2 Equipment Cleaned, #3 Encapsulated
Clothes			X			X		X	X			
Boxes			X			X		X	X			
Backpacks, shoes,			X			X		X	X			
Leather goods			X			X		X	X			
Shelves			X			X		X	X			
Under Sinks/Cabinets			X			X		X	X			



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C32	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Storage Room											
Date:	8/23/2018											
Time:	1000											
Assessor:	Lou Johnson											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No		
Walls	North	x			x			X			Multiple Locations on Walls	#2 Clean Walls
	East	x			x			X			Multiple Locations on Walls	#2 Clean Walls
	South	x			x			X			Multiple Locations on Walls	#2 Clean Walls
	West	x			x			X			Multiple Locations on Walls	#2 Clean Walls
Ceiling	Tiles		x			x		X				
	Above Ceiling		x			x		X				
	Pipes/Insulation/Etc.		x			x		X				
	Drip Pans		x			x		X				
	Lighting		x			x		X				
	Insulation		x			x		X				
	Other		x			x		X				
Floor	Carpet Front		x			x		X				
	Carpet Back		x			x		X				
	Tiles	x			x			X			Multiple Locations	#2 Clean Floors
Doors	Classroom Door		x			x		X				
	Closet Door		x			x		X				
	Bathroom Door		x			x		X				
Door Frames	Classroom		x			x		X				
	Bathroom		x			x		X				
	Closet Door		x			x		X				
	Other (Describe)		x			x		X				
Windows	Frame/Sills/Sash/Curtains		x			x		X				
Bookcases	all sides, top, bottom		x			x		X				
File Cabinets			x			x		X				
Inside Closets			x			x		X				
Bulletin Boards	Check Behind		x			x		X				
Chalkboards	Check Behind		x			x		X				
White Boards	Check Behind		x			x		X				
Wallpaper	Check Behind		x			x		X				
Wall Artwork	Check Behind		x			x		X				
Books/Magazines Etc.			x			x		X				
Room Contents	Games Easels, etc.		x			x		X				
Desks	all sides, top, bottom		x			x		X				
Chairs	all sides, top, bottom		x			x		X				
HVAC system	Supply/Return/Filters/Ducts		x			x		X				
Unit Ventilators	Filter/Cage/Cover		x			x		X				
Equipment	all sides, top, bottom		x			x		X				
Clothes			x			x		X				
Boxes			x			x		X				
Backpacks, shoes,			x			x		X				
Leather goods			x			x		X				
Shelves		x			x			X			Shelf within Room	#1 Dispose of Shelving
Under Sinks/Cabinets			x			x		X				

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C34	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Multipurpose Room											
Date:	8/23/2018											
Time:	1500											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North	x				x		X		<b>Cove Base Molding</b>	<b>#1 Dispose Molding and Adhesive, #3 Encapsulate</b>	
	East		x			x		X				
	South		x			x		X				
	West		x			x		X				
Ceiling	Tiles	x				x		X		<b>(10) 2' x 4' Ceiling Tiles</b>	<b>#1 Dispose of Ceiling Tiles</b>	
	Above Ceiling		x			x		X				
	Pipes/Insulation/Etc.		x			x		X				
	Drip Pans		x			x		X				
	Lighting		x			x		X				
	Insulation		x			x		X				
	Other		x			x		X				
Floor	Carpet Front		x			x		X				
	Carpet Back		x			x		X				
	Tiles		x			x		X				
Doors	Classroom Door		x			x		X				
	Closet Door		x			x		X				
	Bathroom Door		x			x		X				
Door Frames	Classroom		x			x		X				
	Bathroom		x			x		X				
	Closet Door		x			x		X				
Back Cabinets	Other (Describe)	x				x		X		<b>Top of Cabinets under Clock</b>	<b>#2 Clean Cabinets</b>	
Windows	Frame/Sills/Sash/Curtains		x			x		X				
Bookcases	<i>all sides, top, bottom</i>		x			x		X				
File Cabinets			x			x		X				
Inside Closets			x			x		X				
Bulletin Boards	<i>Check Behind</i>		x			x		X				
Chalkboards	<i>Check Behind</i>		x			x		X				
White Boards	<i>Check Behind</i>		x			x		X				
Wallpaper	<i>Check Behind</i>		x			x		X				
Wall Artwork	<i>Check Behind</i>		x			x		X				
Books/Magazines Etc.			x			x		X				
Room Contents	<i>Games Easels, etc.</i>		x		x			X		<b>Refridgerator - Back Room</b>	<b>#2 Clean Refridgerator</b>	
Desks	<i>all sides, top, bottom</i>		x			x		X				
Chairs	<i>all sides, top, bottom</i>		x			x		X				
HVAC system	<i>Supply/Return/Filters/Ducts</i>	x			x			X		<b>Insulation</b>	<b>#1 Dispose of Insulation</b>	
Unit Ventilators	<i>Filter/Cage/Cover</i>		x			x		X				
Equipment	<i>all sides, top, bottom</i>		x			x		X				
Clothes			x			x		X				
Boxes			x			x		X				
Backpacks, shoes,			x			x		X				
Leather goods			x			x		X				
Shelves			x			x		X				
Under Sinks/Cabinets			x			x		X				



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	C 37	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Electrical Room											
Date:	8/23/2018											
Time:												
Assessor:	Frank Manna & Michael Smith											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No	#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	East		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	South		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	West		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Ceiling	Tiles		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Above Ceiling		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Pipes/Insulation/Etc.		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Drip Pans		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Lighting		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Insulation		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Other		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Floor	Carpet Front		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Carpet Back		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Tiles		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Doors	Classroom Door		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Closet Door		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Bathroom Door		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Door Frames	Classroom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Bathroom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Closet Door		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
	Other (Describe)		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Windows	Frame/Sills/Sash/Curtains		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Bookcases	all sides, top, bottom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
File Cabinets			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Inside Closets			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Bulletin Boards	Check Behind		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Chalkboards	Check Behind		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
White Boards	Check Behind		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Wallpaper	Check Behind		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Wall Artwork	Check Behind		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Books/Magazines Etc.			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Room Contents	Games Easels, etc.		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Desks	all sides, top, bottom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Chairs	all sides, top, bottom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
HVAC system	Supply/Return/Filters/Ducts		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Unit Ventilators	Filter/Cage/Cover		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Equipment	all sides, top, bottom		x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Clothes			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Boxes			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Backpacks, shoes,			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Leather goods			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Shelves			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	
Under Sinks/Cabinets			x			x		X			NO VISIBLE DAMAGE THROUGHOUT	





School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet									
Room #:	Mens Faculty	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.									
Room Type:	Bathroom										
Date:	8/23/2018										
Time:	1500										
Assessor:	Mike Smith										
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		x			x	X			No Visible Damage	
	East		x			x	X			No Visible Damage	
	South		x			x	X			No Visible Damage	
	West		x			x	X			No Visible Damage	
Ceiling	Tiles		x			x	X			No Visible Damage	
	Above Ceiling		x			x	X			No Visible Damage	
	Pipes/Insulation/Etc.		x			x	X			No Visible Damage	
	Drip Pans		x			x	X			No Visible Damage	
	Lighting		x			x	X			No Visible Damage	
	Insulation		x			x	X			No Visible Damage	
	Other		x			x	X			No Visible Damage	
Floor	Carpet Front		x			x	X			No Visible Damage	
	Carpet Back		x			x	X			No Visible Damage	
	Tiles		x			x	X			No Visible Damage	
Doors	Classroom Door		x			x	X			No Visible Damage	
	Closet Door		x			x	X			No Visible Damage	
	Bathroom Door		x			x	X			No Visible Damage	
Door Frames	Classroom		x			x	X			No Visible Damage	
	Bathroom		x			x	X			No Visible Damage	
	Closet Door		x			x	X			No Visible Damage	
	Other (Describe)		x			x	X			No Visible Damage	
Windows	Frame/Sills/Sash/Curtains		x			x	X			No Visible Damage	
Bookcases	all sides, top, bottom		x			x	X			No Visible Damage	
File Cabinets			x			x	X			No Visible Damage	
Inside Closets			x			x	X			No Visible Damage	
Bulletin Boards	Check Behind		x			x	X			No Visible Damage	
Chalkboards	Check Behind		x			x	X			No Visible Damage	
White Boards	Check Behind		x			x	X			No Visible Damage	
Wallpaper	Check Behind		x			x	X			No Visible Damage	
Wall Artwork	Check Behind		x			x	X			No Visible Damage	
Books/Magazines Etc.			x			x	X			No Visible Damage	
Room Contents	Games Easels, etc.		x			x	X			No Visible Damage	
Desks	all sides, top, bottom		x			x	X			No Visible Damage	
Chairs	all sides, top, bottom		x			x	X			No Visible Damage	
HVAC system	Supply/Return/Filters/Ducts		x			x	X			No Visible Damage	
Unit Ventilators	Filter/Cage/Cover		x			x	X			No Visible Damage	
Equipment	all sides, top, bottom		x			x	X			No Visible Damage	
Clothes			x			x	X			No Visible Damage	
Boxes			x			x	X			No Visible Damage	
Backpacks, shoes,			x			x	X			No Visible Damage	
Leather goods			x			x	X			No Visible Damage	
Shelves			x			x	X			No Visible Damage	
Under Sinks/Cabinets			x			x	X			No Visible Damage	





School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Womens Faculty	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Bathroom											
Date:	8/27/2018											
Time:	930											
Assessor:	Mike Smith											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)	Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No		No	Yes	No	#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North		x			x		X			No Visible Damage	
	East		x			x		X			No Visible Damage	
	South		x			x		X			No Visible Damage	
	West		x			x		X			No Visible Damage	
Ceiling	Tiles		x			x		X			No Visible Damage	
	Above Ceiling		x			x		X			No Visible Damage	
	Pipes/Insulation/Etc.		x			x		X			No Visible Damage	
	Drip Pans		x			x		X			No Visible Damage	
	Lighting		x			x		X			No Visible Damage	
	Insulation		x			x		X			No Visible Damage	
	Other		x			x		X			No Visible Damage	
Floor	Carpet Front		x			x		X			No Visible Damage	
	Carpet Back		x			x		X			No Visible Damage	
	Tiles		x			x		X			No Visible Damage	
Doors	Classroom Door		x			x		X			No Visible Damage	
	Closet Door		x			x		X			No Visible Damage	
	Bathroom Door		x			x		X			No Visible Damage	
Door Frames	Classroom		x			x		X			No Visible Damage	
	Bathroom		x			x		X			No Visible Damage	
	Closet Door		x			x		X			No Visible Damage	
	Other (Describe)		x			x		X			No Visible Damage	
Windows	Frame/Sills/Sash/Curtains		x			x		X			No Visible Damage	
Bookcases	all sides, top, bottom		x			x		X			No Visible Damage	
File Cabinets			x			x		X			No Visible Damage	
Inside Closets			x			x		X			No Visible Damage	
Bulletin Boards	Check Behind		x			x		X			No Visible Damage	
Chalkboards	Check Behind		x			x		X			No Visible Damage	
White Boards	Check Behind		x			x		X			No Visible Damage	
Wallpaper	Check Behind		x			x		X			No Visible Damage	
Wall Artwork	Check Behind		x			x		X			No Visible Damage	
Books/Magazines Etc.			x			x		X			No Visible Damage	
Room Contents	Games Easels, etc.		x			x		X			No Visible Damage	
Desks	all sides, top, bottom		x			x		X			No Visible Damage	
Chairs	all sides, top, bottom		x			x		X			No Visible Damage	
HVAC system	Supply/Return/Filters/Ducts		x			x		X			No Visible Damage	
Unit Ventilators	Filter/Cage/Cover		x			x		X			No Visible Damage	
Equipment	all sides, top, bottom		x			x		X			No Visible Damage	
Clothes			x			x		X			No Visible Damage	
Boxes			x			x		X			No Visible Damage	
Backpacks, shoes,			x			x		X			No Visible Damage	
Leather goods			x			x		X			No Visible Damage	
Shelves			x			x		X			No Visible Damage	
Under Sinks/Cabinets			x			x		X			No Visible Damage	

**Appendix D(3)**  
**Mold Assessment Documentation**  
**East Wing**

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-10	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Storage/Classroom											
Date:	8/29/2018											
Time:	1030											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X		X		
	East		X			X		X		X		
	South		X			X		X		X		
	West		X			X		X		X		
Ceiling	Tiles		X			X		X		X		
	Above Ceiling		X			X		X		X		
	Pipes/Insulation/Etc.		X			X		X		X		
	Drip Pans		X			X		X		X		
	Lighting		X			X		X		X		
	Insulation		X			X		X		X		
	Other		X			X		X		X		
Floor	Carpet Front		X			X		X		X		
	Carpet Back		X			X		X		X		
	Tiles		X			X		X		X		
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X		X		
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X	X			
	Bathroom		X			X		X		X		
	Closet Door		X			X		X		X		
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		
Bookcases	all sides, top, bottom	X				X		X	X	Bottoms of Bookcases to be Cleaned	#2 Clean	
File Cabinets			X			X		X	X			
Inside Closets			X			X		X	X			
Bulletin Boards	Check Behind		X			X		X	X			
Chalkboards	Check Behind		X			X		X	X			
White Boards	Check Behind		X			X		X	X			
Wallpaper	Check Behind		X			X		X	X			
Wall Artwork	Check Behind		X			X		X	X			
Books/Magazines Etc.		X				X		X	X			
Room Contents	Games Easels, etc.		X			X		X	X			#1 Dispose of 18 Books, #2 Clean
Desks	all sides, top, bottom	X				X		X	X	Bottoms of Desks to be Cleaned	#2 Clean	
Chairs	all sides, top, bottom	X				X		X	X	Bottoms of Chairs to be Cleaned	#2 Clean	
HVAC system	Supply/Return/Filters/Ducts	X				X		X	X	HVAC Supply to be Cleaned	#2 Clean	
Unit Ventilators	Filter/Cage/Cover		X			X		X	X		#2 Clean	
Equipment	all sides, top, bottom		X			X		X	X		#2 Clean	
Clothes			X			X		X	X			
Boxes			X			X		X	X			
Backpacks, shoes,			X			X		X	X			
Leather goods			X			X		X	X			
Shelves			X			X		X	X		#2 Clean	
Under Sinks/Cabinets			X			X		X	X			



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet												
Room #:	E-11	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.												
Room Type:	Math Classroom													
Date:	8/29/2018													
Time:	1030													
Assessor:	Louis Johnson III													
										Response Action				
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	#1 Dispose/ #2 Clean/ #3 Encapsulate		
		Yes	No		Yes	No	Yes	No	Yes	No				
Walls	North	X		30 SF		X		X	X		On Cementitious Block	#3 Encapsulate		
	East		X			X		X	X					
	South	X		20 SF		X		X	X		Under Whiteboard	#1 Dispose of 20 SF of Sheetrock		
	West	X		20 SF		X		X	X		On Cementitious Block	#3 Encapsulate		
Ceiling	Tiles	X		32 SF		X		X	X		Tiles Next to HVAC System	#1 Dispose of 32 SF of Ceiling Tile		
	Above Ceiling		X			X		X		X		#2 Clean, #3 Encapsulate		
	Pipes/Insulation/Etc.		X			X		X		X				
	Drip Pans		X			X		X		X				
	Lighting		X			X		X		X				
	Insulation		X			X		X		X				
	Other		X			X		X		X				
Floor	Carpet Front		X			X		X		X				
	Carpet Back		X			X		X		X				
	Tiles		X			X		X		X		#2 Clean		
Doors	Classroom Door	X				X		X		X		#2 Clean, #3 Encapsulate		
	Closet Door		X			X		X	X			#2 Clean, #3 Encapsulate		
	Bathroom Door		X			X		X		X				
Door Frames	Classroom		X		X			X	X			#2 Clean, #3 Encapsulate		
	Bathroom		X			X		X		X				
	Closet Door		X		X			X	X			#2 Clean, #3 Encapsulate		
	Other (Describe)		X			X		X		X				
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean, #3 Encapsulate		
Bookcases	all sides, top, bottom	X				X		X	X		Back Side of Bookcases to be Cleaked	#2 Clean		
File Cabinets		X				X		X	X		Top of Black Cabinet to be Cleaned	#2 Clean		
Inside Closets			X			X		X	X			#2 Clean		
Bulletin Boards	Check Behind		X			X		X		X				
Chalkboards	Check Behind		X			X		X	X					
White Boards	Check Behind		X			X		X	X					
Wallpaper	Check Behind		X			X		X		X				
Wall Artwork	Check Behind		X			X		X		X				
Books/Magazines Etc.			X			X		X		X		#2 Clean		
Room Contents	Games Easels, etc.		X			X		X	X			#2 Clean		
Desks	all sides, top, bottom	X				X		X	X		All Sides of Desks to be Cleaned	#2 Clean		
Chairs	all sides, top, bottom	X				X		X		X	Tops of Chairs to be Cleaned	#2 Clean		
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		#2 Clean		
Unit Ventilators	Filter/Cage/Cover	X				X		X		X		#2 Clean		
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean		
Clothes			X			X		X		X				
Boxes			X			X		X		X				
Backpacks, shoes,			X			X		X		X				
Leather goods			X			X		X		X				
Shelves			X			X		X	X			#2 Clean		
Under Sinks/Cabinets			X			X		X		X				

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-12	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	940											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		30 SF		X		X	X		On Cementitious Block	#3 Encapsulate
	East	X		20 SF		X		X		X	On Cementitious Block	#3 Encapsulate
	South	X		45 SF		X		X	X		On Sheetrock by Rear Corner	#1 Dispose of 4 SF of Sheetrock
	West		X			X		X		X		
Ceiling	Tiles	X		32 SF	X			X	X		Center of Room	#1 Dispose of 32 SF of Ceiling Tile
	Above Ceiling		X			X		X		X		
	Pipes/Insulation/Etc.		X			X		X		X		
	Drip Pans		X			X		X		X		
	Lighting		X			X		X		X		
	Insulation		X			X		X		X		
	Other		X			X		X		X		
Floor	Carpet Front		X			X		X		X		
	Carpet Back		X			X		X		X		
	Tiles		X			X		X		X		#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X	X			#2 Clean
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X		X		
	Closet Door		X			X		X	X			#2 Clean
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean
Bookcases	all sides, top, bottom		X			X		X		X	Back Side of Bookcases to be Cleaked	#2 Clean
File Cabinets			X			X		X		X	Top of Black Cabinet to be Cleaned	#2 Clean
Inside Closets			X			X		X		X		#2 Clean
Bulletin Boards	Check Behind		X			X		X		X		
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X		X		
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X	X			
Books/Magazines Etc.			X			X		X	X			#2 Clean
Room Contents	Games Easels, etc.	X				X		X	X			#1 Dispose of 2 Bags of Books
Desks	all sides, top, bottom	X				X		X	X		Bottoms of Desks to be Cleaned	#2 Clean
Chairs	all sides, top, bottom		X			X		X	X			#2 Clean
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		
Equipment	all sides, top, bottom		X			X		X		X		
Clothes			X			X		X		X		
Boxes			X			X		X		X		
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		
Shelves			X			X		X	X			#2 Clean
Under Sinks/Cabinets			X			X		X		X		

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet											
Room #:	E-13	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.											
Room Type:	Computer Lab												
Date:	8/29/2018												
Time:	1444												
Assessor:	Louis Johnson III												
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action	
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate	
Walls	North	X		20 SF		X		X	X		Base Trim	#1 Dispose of 20 SF of Sheetrock	
	East	X		45 SF		X		X	X		South East Corner	#3 Encapsulate, #1 Dispose of 4 SF of Sheetrock	
	South		X	20 SF		X		X	X		Base Trim	#1 Dispose of 4 SF of Sheetrock	
	West	X		30 SF		X		X	X			#1 Dispose of 20 SF of Sheetrock	
Ceiling	Tiles	X		56 SF	X			X	X			#1 Dispose of 56 SF of Ceiling Tile	
	Above Ceiling		X			X		X		X			
	Pipes/Insulation/Etc.		X			X		X		X			
	Drip Pans		X			X		X		X			
	Lighting		X			X		X		X			
	Insulation		X			X		X		X			
	Other		X			X		X		X			
Floor	Carpet Front		X			X		X		X			
	Carpet Back		X			X		X		X			
	Tiles		X			X		X		X		#2 Clean	
Doors	Classroom Door		X			X		X	X			#2 Clean	
	Closet Door		X			X		X		X			
	Bathroom Door		X			X		X		X			
Door Frames	Classroom		X			X		X	X			#2 Clean	
	Bathroom		X			X		X		X			
	Closet Door		X			X		X		X			
	Other (Describe)		X			X		X		X			
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean	
Bookcases	all sides, top, bottom	X		10 SF		X		X	X		Bottom of Bookshelf	#2 Clean	
File Cabinets			X			X		X	X			#2 Clean	
Inside Closets			X			X		X	X				
Bulletin Boards	Check Behind		X			X		X		X		#2 Clean	
Chalkboards	Check Behind		X			X		X		X			
White Boards	Check Behind		X			X		X		X		#2 Clean	
Wallpaper	Check Behind		X			X		X		X			
Wall Artwork	Check Behind		X			X		X		X			
Books/Magazines Etc.			X			X		X		X		#2 Clean	
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean	
Desks	all sides, top, bottom		X			X		X		X		#2 Clean	
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean	
HVAC system	Supply/Return/Filters/Ducts	X				X		X		X		Around Metal Fans	#2 Clean, #3 Encapsulate
Unit Ventilators	Filter/Cage/Cover		X			X		X		X			#2 Clean, #3 Encapsulate
Equipment	all sides, top, bottom		X			X		X		X			
Clothes			X			X		X		X			
Boxes			X			X		X		X			
Backpacks, shoes,			X			X		X		X			
Leather goods			X			X		X		X			
Shelves			X			X		X	X				#2 Clean
Under Sinks/Cabinets			X			X		X		X			

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-14	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	1240											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		10 SF		X		X	X		Outside Whiteboard	#1 Dispose of 10 SF of Sheetrock
	East	X		10 SF		X		X	X		On Cementitious Block	#3 Encapsulate
	South	X		16 SF		X		X	X		Rear Section Base	#1 Dispose of 16 SF of Sheetrock
	West	X		45 SF		X		X	X		By Door	#1 Dispose of 45 SF of Sheetrock
Ceiling	Tiles		X	40 SF	X			X	X			#1 Dispose of 40 SF of Ceiling Tiles
	Above Ceiling		X			X		X		X		
	Pipes/Insulation/Etc.		X			X		X		X		
	Drip Pans		X			X		X		X		
	Lighting		X			X		X		X		
	Insulation		X			X		X		X		
	Other		X			X		X		X		
				X			X		X		X	
Floor	Carpet Front		X			X		X	X			#1 Dispose of Carpet by Whiteboard
	Carpet Back	X				X		X	X			
	Tiles		X			X		X		X		#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X		X		
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X		X		
	Closet Door		X			X		X		X		
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean
Bookcases	all sides, top, bottom	X		85 SF		X		X		X	Bottom of Bookshelf	#2 Clean
File Cabinets			X			X		X		X		#2 Clean
Inside Closets			X			X		X		X		
Bulletin Boards	Check Behind		X			X		X	X			#2 Clean
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X	X			#2 Clean
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		#2 Clean
Books/Magazines Etc.			X			X		X		X		
Room Contents	Games Easels, etc.		X			X		X		X		#2 Clean
Desks	all sides, top, bottom	X				X		X		X	Sides and Bottoms of Desks	#2 Clean
Chairs	all sides, top, bottom		X			X		X		X		#2 Clean
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		#2 Clean
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean
Clothes			X			X		X		X		
Boxes			X			X		X		X		
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		
Shelves			X			X		X	X			#2 Clean
Under Sinks/Cabinets			X			X		X		X		



School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-15	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	1300											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		30 SF		X		X	X		Entry Base Perimeter Wall	#1 Dispose of 30 SF of Sheetrock
	East	X		6 SF		X		X	X		By Door	#1 Dispose of 6 SF of Sheetrock
	South	X		20 SF		X		X	X		Half of Base	#1 Dispose of 20 SF of Sheetrock
	West	X		30 SF		X		X	X		On Cementitious Block	#2 Clean, #3 Encapsulate
Ceiling	Tiles	X		48 SF	X			X	X		4 Ceiling Tiles	#1 Dispose of 48 SF of Ceiling Tiles
	Above Ceiling		X			X		X	X			
	Pipes/Insulation/Etc.		X			X		X	X			
	Drip Pans		X			X		X	X			
	Lighting		X			X		X	X			
	Insulation		X			X		X	X			
	Other		X			X		X	X			
Floor	Carpet Front		X			X		X	X		Multi-Colored USA Map	#1 Dispose of Carpet
	Carpet Back	X				X		X	X		Multi-Colored USA Map	#1 Dispose of Carpet
	Tiles		X			X		X	X			#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X	X			#2 Clean
	Bathroom Door		X			X		X	X			
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X	X			
	Closet Door		X			X		X	X			#2 Clean
	Other (Describe)		X			X		X	X			#2 Clean
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean
Bookcases	all sides, top, bottom		X			X		X	X			#2 Clean
File Cabinets			X			X		X	X			#2 Clean
Inside Closets			X			X		X	X			#2 Clean
Bulletin Boards	Check Behind		X			X		X	X			
Chalkboards	Check Behind		X			X		X	X			
White Boards	Check Behind		X			X		X	X			
Wallpaper	Check Behind		X			X		X	X			
Wall Artwork	Check Behind		X			X		X	X			
Books/Magazines Etc.			X			X		X	X			
Room Contents	Games Easels, etc.		X			X		X	X			#2 Clean
Desks	all sides, top, bottom		X			X		X	X			#2 Clean
Chairs	all sides, top, bottom	X				X		X	X		Tennis Balls on Chairs	#1 Dispose of Tennis Balls on Bottoms of Chairs
HVAC system	Supply/Return/Filters/Ducts		X			X		X	X			
Unit Ventilators	Filter/Cage/Cover		X			X		X	X			#2 Clean
Equipment	all sides, top, bottom		X			X		X	X			#2 Clean
Clothes			X			X		X	X			#2 Clean
Boxes			X			X		X	X			
Backpacks, shoes,			X			X		X	X			
Leather goods			X			X		X	X			
Shelves			X			X		X	X			#2 Clean
Under Sinks/Cabinets			X			X		X	X			

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-16	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	1510											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		30 SF		X		X	X		Perimeter Wall	#3 Encapsulate
	East	X		30 SF		X		X	X		Entry Wall	#1 Dispose of 30 SF of Sheetrock
	South	X		30 SF		X		X	X		Perimeter Wall	#1 Dispose of 30 SF of Sheetrock
	West	X		8 SF		X		X	X		By Door	#1 Dispose of 8 SF of Sheetrock
Ceiling	Tiles		X	8 SF	X			X	X			#1 Dispose of 8 SF of Ceiling Tile
	Above Ceiling		X			X		X		X		
	Pipes/Insulation/Etc.		X			X		X		X		
	Drip Pans		X			X		X		X		
	Lighting		X			X		X		X		
	Insulation		X			X		X		X		
	Other		X			X		X		X		
Floor	Carpet Front		X			X		X	X			
	Carpet Back	X				X		X	X			#1 Dispose of Carpet
	Tiles		X			X		X	X			#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X	X			#2 Clean
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X		X		
	Closet Door		X			X		X	X			#2 Clean
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X			X		X		X		
Bookcases	all sides, top, bottom		X			X		X		X		
File Cabinets			X			X		X		X		
Inside Closets			X			X		X	X			#2 Clean
Bulletin Boards	Check Behind		X			X		X	X			#2 Clean
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X	X			#2 Clean
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		
Books/Magazines Etc.			X			X		X	X			#2 Clean
Room Contents	Games Easels, etc.		X			X		X	X			#2 Clean
Desks	all sides, top, bottom		X			X		X	X			#2 Clean
Chairs	all sides, top, bottom	X				X		X	X		Tennis Balls on Chairs	#1 Dispose of Tennis Balls on Bottoms of Chairs
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		
Unit Ventilators	Filter/Cage/Cover		X			X		X	X			#2 Clean
Equipment	all sides, top, bottom		X			X		X	X			#2 Clean
Clothes			X	40 SF		X		X		X		#1 Dispose of Seat Cushions
Boxes			X			X		X		X		
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		
Shelves			X			X		X	X			#2 Clean
Under Sinks/Cabinets			X			X		X		X		

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-17	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	1148											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X	X			
	East	X				X		X	X		On Cementitious Wall	#3 Encapsulate
	South	X		24 SF		X		X	X		On Cementitious Wall	#3 Encapsulate
	West	X		30 SF		X		X	X		Perimeter Wall	#1 Dispose of 30 SF of Sheetrock
Ceiling	Tiles	X		40 SF	X			X	X		By Front Vent	#1 Dispose of 40 SF of Ceiling Tile
	Above Ceiling		X			X		X		X		
	Pipes/Insulation/Etc.		X			X		X		X		
	Drip Pans		X			X		X		X		
	Lighting		X			X		X		X		
	Insulation		X			X		X		X		
	Other		X			X		X		X		
Floor	Carpet Front		X			X		X	X			
	Carpet Back	X				X		X	X			#1 Dispose of Carpet by Whiteboard
	Tiles		X			X		X		X		#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X	X			#2 Clean
	Bathroom Door		X			X		X		X		
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X		X		
	Closet Door		X			X		X	X			#2 Clean
	Other (Describe)		X			X		X		X		
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean
Bookcases	all sides, top, bottom	X			X			X	X		Bottom of Bookcase on Wood Shelf	#2 Clean
File Cabinets			X			X		X		X		
Inside Closets			X			X		X	X			#2 Clean
Bulletin Boards	Check Behind		X			X		X	X			#2 Clean
Chalkboards	Check Behind		X			X		X		X		
White Boards	Check Behind		X			X		X	X			#2 Clean
Wallpaper	Check Behind		X			X		X		X		
Wall Artwork	Check Behind		X			X		X		X		#2 Clean
Books/Magazines Etc.			X			X		X	X			#2 Clean
Room Contents	Games Easels, etc.		X			X		X	X			#2 Clean
Desks	all sides, top, bottom		X			X		X	X			#2 Clean
Chairs	all sides, top, bottom	X		6 SF		X		X	X		Tennis Balls on Chairs	#1 Dispose of Tennis Balls on Bottoms of Chairs
HVAC system	Supply/Return/Filters/Ducts		X			X		X		X		
Unit Ventilators	Filter/Cage/Cover		X			X		X		X		#2 Clean
Equipment	all sides, top, bottom		X			X		X		X		#2 Clean
Clothes			X			X		X		X		
Boxes			X			X		X		X		
Backpacks, shoes,			X			X		X		X		
Leather goods			X			X		X		X		
Shelves			X			X		X	X		Bottom Metal Shelf & Drawers	#2 Clean
Under Sinks/Cabinets		X				X		X	X		Bottom of Sink & Pipe	#2 Clean

School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	E-18	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Classroom											
Date:	8/29/2018											
Time:	1130											
Assessor:	Louis Johnson III											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North	X		20 SF		X		X	X		Perimeter Wall	#3 Encapsulate
	East	X		20 SF		X		X	X		Perimeter Wall	#3 Encapsulate
	South	X		15 SF		X		X	X		South Wall	#2 Clean, #3 Encapsulate, #1 Dispose of 5 SF of Sheetrock
	West	X		30 SF		X		X	X		Perimeter Wall	#1 Dispose of 30 SF of Sheetrock
Ceiling	Tiles		X	80 SF		X		X	X		Front and Rear	#1 Dispose of 10 SF of Ceiling Tile
	Above Ceiling		X			X		X	X			
	Pipes/Insulation/Etc.		X			X		X	X			
	Drip Pans		X			X		X	X			
	Lighting		X			X		X	X			#2 Clean
	Insulation		X			X		X	X			
	Other		X			X		X	X			
Floor	Carpet Front	X				X		X	X			
	Carpet Back	X				X		X	X			#1 Dispose of Back Carpet
	Tiles		X			X		X	X			#2 Clean
Doors	Classroom Door		X			X		X	X			#2 Clean
	Closet Door		X			X		X	X			#2 Clean
	Bathroom Door		X			X		X	X			
Door Frames	Classroom		X			X		X	X			#2 Clean
	Bathroom		X			X		X	X			
	Closet Door		X			X		X	X			#2 Clean
	Other (Describe)		X			X		X	X			
Windows	Frame/Sills/Sash/Curtains		X			X		X	X			#2 Clean
Bookcases	all sides, top, bottom		X			X		X	X			#2 Clean
File Cabinets		X		20 SF		X		X	X	5 Drawer Cabinet Contents	#1 Dispose of Contents inside File Cabinet Drawers	
Inside Closets			X			X		X	X			#2 Clean
Bulletin Boards	Check Behind		X			X		X	X			#2 Clean
Chalkboards	Check Behind		X			X		X	X			
White Boards	Check Behind		X			X		X	X			#2 Clean
Wallpaper	Check Behind		X			X		X	X			
Wall Artwork	Check Behind		X			X		X	X			
Books/Magazines Etc.			X			X		X	X			#2 Clean
Room Contents	Games Easels, etc.		X			X		X	X			
Desks	all sides, top, bottom		X			X		X	X			#2 Clean
Chairs	all sides, top, bottom	X				X		X	X	Tennis Balls on Chairs	#1 Dispose of Tennis Balls on Bottoms of Chairs	
HVAC system	Supply/Return/Filters/Ducts		X			X		X	X			
Unit Ventilators	Filter/Cage/Cover		X			X		X	X			#2 Clean
Equipment	all sides, top, bottom		X			X		X	X			#2 Clean
Clothes			X			X		X	X			
Boxes			X			X		X	X			
Backpacks, shoes,			X			X		X	X			
Leather goods			X			X		X	X			
Shelves		X		5 SF		X		X	X	Bottom Metal Shelf & Drawers	#2 Clean, #3 Encapsulate	
Under Sinks/Cabinets		X		4 SF		X		X	X	Bottom of Sink & Pipe	#2 Clean, #3 Encapsulate	





**Appendix D(4)**  
**Mold Assessment Documentation**  
**North Wing**





































































**Appendix D(5)**  
**Mold Assessment Documentation**  
**Gym / Lower Level Wing**











School Name:	Pequanock Elementary	Mold Assessment Field Documentation Sheet										
Room #:	Gym	Every space that is assessed must be 1) Visually inspected for the presence of visible suspected microbial (mold) growth. This may appear as dark stains, discolorations, and fuzzy areas. 2) All spaces must be scanned with a thermal imaging camera to properly assess the walls, ceilings and other surfaces for existing moisture. Notes should be made of mold odors, areas of visible water, leaks etc. Do not rush when using a TIC as the camera needs time to evaluate the thermal conditions present. If visible mold is discovered, sufficient documentation of the amount and location must be recorded to ensure that all identified areas are addressed as part of the final remediation plan.										
Room Type:	Gymnasium											
Date:	8/25/2018											
Time:	1030											
Assessor:	Tanay Ranadive											
Room Component		Fungal Growth		Qnty. Ft <sup>2</sup>	Visible Water Damage		Currently Wet (TIC/MM)		Porous		Location/Description/Comments	Response Action
		Yes	No		Yes	No	Yes	No	Yes	No		#1 Dispose/ #2 Clean/ #3 Encapsulate
Walls	North		X			X		X		X		#2 Clean, #3 Encapsulate
	East	X		10 LF		X		X	X		Cove Base Adhesive	#2 Clean, #3 Encapsulate
	South		X			X		X		X		#2 Clean, #3 Encapsulate
	West		X			X		X		X		#2 Clean, #3 Encapsulate
Ceiling	Tiles		X			X		X		X		#2 Clean, #3 Encapsulate
	Above Ceiling		X			X		X				#2 Clean, #3 Encapsulate
	Pipes/Insulation/Etc.		X			X		X				#2 Clean, #3 Encapsulate
	Drip Pans		X			X		X				#2 Clean, #3 Encapsulate
	Lighting		X			X		X				#2 Clean, #3 Encapsulate
	Insulation		X			X		X				#2 Clean, #3 Encapsulate
	Other		X			X		X				#2 Clean, #3 Encapsulate
				X			X		X			
Floor	Carpet Front		X			X		X				#2 Clean, #3 Encapsulate
	Carpet Back		X			X		X				#2 Clean, #3 Encapsulate
	Tiles		X			X		X				#2 Clean, #3 Encapsulate
Doors	Classroom Door		X			X		X				#2 Clean, #3 Encapsulate
	Closet Door		X			X		X				#2 Clean, #3 Encapsulate
	Bathroom Door		X			X		X				#2 Clean, #3 Encapsulate
Door Frames	Classroom		X			X		X				#2 Clean, #3 Encapsulate
	Bathroom		X			X		X				#2 Clean, #3 Encapsulate
	Closet Door		X			X		X				#2 Clean, #3 Encapsulate
	Other (Describe)		X			X		X				#2 Clean, #3 Encapsulate
Windows	Frame/Sills/Sash/Curtains		X			X		X				#2 Clean, #3 Encapsulate
Bookcases	all sides, top, bottom		X			X		X				#2 Clean, #3 Encapsulate
File Cabinets			X			X		X				#2 Clean, #3 Encapsulate
Inside Closets			X			X		X				#2 Clean, #3 Encapsulate
Bulletin Boards	Check Behind		X			X		X				#2 Clean, #3 Encapsulate
Chalkboards	Check Behind		X			X		X				#2 Clean, #3 Encapsulate
White Boards	Check Behind		X			X		X				#2 Clean, #3 Encapsulate
Wallpaper	Check Behind		X			X		X				#2 Clean, #3 Encapsulate
Wall Artwork	Check Behind		X			X		X				#2 Clean, #3 Encapsulate
Books/Magazines Etc.			X			X		X				#2 Clean, #3 Encapsulate
Room Contents	Games Easels, etc.		X			X		X				#2 Clean, #3 Encapsulate
Desks	all sides, top, bottom		X			X		X				#2 Clean, #3 Encapsulate
Chairs	all sides, top, bottom		X			X		X				#2 Clean, #3 Encapsulate
HVAC system	Supply/Return/Filters/Ducts		X			X		X				#2 Clean, #3 Encapsulate
Unit Ventilators	Filter/Cage/Cover		X			X		X				#2 Clean, #3 Encapsulate
Equipment	all sides, top, bottom		X			X		X				#2 Clean, #3 Encapsulate
Clothes			X			X		X				#2 Clean, #3 Encapsulate
Boxes			X			X		X				#2 Clean, #3 Encapsulate
Backpacks, shoes,			X			X		X				#2 Clean, #3 Encapsulate
Leather goods			X			X		X				#2 Clean, #3 Encapsulate
Shelves			X			X		X				#2 Clean, #3 Encapsulate
Under Sinks/Cabinets			X			X		X				#2 Clean, #3 Encapsulate











# **Appendix D(6)**

## **LOGS**



# QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

## DAILY PROJECT LOG

Mold Assessor: Louis N Johnson, Tanay Ranadive  
Date: 08/22/18

Work Site: Pequenakonck. E.S.

Encl: South Wing

Type of Removal: Mold Remediation

Project #: Q18-1941

### Activity:

- 0930 Louis N Johnson of QuES&T arrives on site with All Pro Remediation, goes over the scope of work with All Pro Supervisor. I start conducting initial mold assessments on South Wing hallways, classrooms & all closets.  
All Pro begins setting up negative air machines on scrub mode and dehumidifiers. All Pro workers suit up in full PPE and begin cleaning all 1<sup>st</sup> Grade classrooms and hallway of South Wing.
- 1200 All Pro Crew and I break for 30 minute lunch.
- 1230 All Pro Crew and I return from 30 minute lunch. Crew suits up in full PPE and continues cleaning 1<sup>st</sup> Grade, Kindergarten sections of South Wing.
- 1300 Mold Assessor Tanay Ranadive of QuES&T arrives onsite, and I walk him through the entire project. Kindergarten and 1<sup>st</sup> Grade sections of South Wing are cordoned off with poly and appropriate signage.
- 1500 All Pro crew begins deconning out of PPE. Initial visual inspections of 1<sup>st</sup> Grade South Wing are complete. Half of 1<sup>st</sup> Grade South Wing have been cleaned by All Pro. Initial Mold Assessments of Kindergarten rooms and hallways are continuing.
- 1530 All Pro crew is off site.
- 1600 Louis Johnson and I are off site. Kindergarten and 2<sup>nd</sup> Grade initial mold assessments will continue tomorrow.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Louis N Johnson

Date: 8/23/18

Work Site: Pequenakonck E.S.

Encl: South Wing

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Louis Johnson, Frank Manna and Michael Smith, Mold Assessors of QuES&T on site. All Pro on site and will continue cleaning 1<sup>st</sup> Grade South Wing classroom. While conducting initial visual assessments in 2<sup>nd</sup> Grade South Wing, I noticed microbial growth on lower walls in and behind cove base on sheetrock walls.

1100 I went back to inspect classrooms S-26, S-25, S-24, S-23 to see if microbial growth is behind cove base along with hallway outside classrooms. Going forward with assessments, all cove base molding will be removed to check for microbial growth and All Pro will remove impacted sheetrock and any attached items to impacted walls; ie bookcases, shelves, etc. In room S-16 noted roof leaking, informed custodial staff.

1200 Crew breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch.

1300 Continue initial mold assessments in Kindergarten South Wing and 2<sup>nd</sup> Grade Wing. All Pro continues to clean in 1<sup>st</sup> Grade South Wing and Kindergarten Wing.

1500 Initial mold assessment complete in South Wing Kindergarten and 2<sup>nd</sup> Grade classrooms.

1600 QuES&T and All Pro are off site.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Louis N Johnson  
Date: 8/24/18

Work Site: Pequenakonck E.S.

Encl: South Wing

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Louis Johnson, Frank Manna and Michael Smith, Mold Assessors of QuES&T on site. All Pro on site setting up additional negative air machines, scrubbers and dehumidifiers in each wing and Library Center core section.

1100 All Pro is going to clean and remove impacted materials and sheetrock. After removals are completed All Pro will have cleaning crew go back into areas and re-clean prior to assessments. Initial assessments continue in West Wing, Main Lobby Area and in center core location. Noticed many of the carpets and chairs along with bookcases have microbial growth. Items that cannot be cleaned will be discarded in clear bags.

1200 Crew breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch.

1300 All Pro continues to work in South Wing. Initial assessments completed in West, Center and Gym lower level wings.

1500 Started East and North Wings. Impacted ceiling tiles by microbial growth on water stains have been removed.

1600 All Pro and QuES&T are off site.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Tanay Ranadive  
Date: 8/25/18

Work Site: Pequenaconck E.S.

Encl: South Wing

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 I, Tanay Ranadive, Mold Assessor from QuES&T arrive on site with Technicians: Miguel Lawrence & Justin McFarland. The scope of work is discussed with All Pro supervisor Marco. All Pro suits up in full PPE and continues cleaning in the 2<sup>nd</sup> Grade and Kindergarten section of the South Wing. Technicians Miguel Lawrence and Justin McFarland and I continue the initial mold assessment for North Wing. ALL PRO crews are continuing sheetrock removal in the 2<sup>nd</sup> Grade & Kindergarten South Wing.

1200 ALL PRO crew decons out of PPE. All Pro and QuES&T break for 30 minute lunch.

1230 We return from 30 minute lunch. Initial assessments of North Wing complete, initial assessments of East Wing begin. Some crew starts removing ceiling tiles from North Wing.

1500 Sheetrock removal & cleaning of all of South Wing complete, all ceiling tiles from North Wing classrooms & hallways complete. Crew decons out and is off site. Miguel, Justin and I continue with initial mold assessments for East Wing.

1600 East Wing mold initial assessments are half-way complete. Miguel, Justin, and I are off site.



QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Tanay Ranadive  
Date: 8/26/18

Work Site: Pequenaconck E.S.

Encl: Center Area

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 I, Tanay Ranadive, Mold Assessor of QuES&T onsite with Technician Zach Timpano. All Pro crew suits up in full PPE and begins cleaning the center wing hallway and adjacent rooms. Sheetrock removal continues in these rooms. Center wing is cordoned off with signage and poly.

1200 Crew and I break for 30 minute lunch.

1230 Crew and I return from 30 minute lunch and suit up in PPE and continue cleaning center wing. Zach Timpano and I set up pumps in South wing for mold clearances. Mold cassettes on pumps, 10 minute wait begins.

1300 Mold cassettes collected, Zach is off site to Q Labs. I continue initials of East Wing. Multipurpose & Cafeteria rooms are sampled as well.

1500 Center wing and sheetrock removal complete. All Pro decons out and is off site.

1600 East Wing initial assessments complete. I am off site.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Louis N Johnson, Tanay Ranadive  
Date: 8/27/18

Work Site: Pequenakonck E.S.

Encl: South Wing

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Mold Assessors of QuES&T Louis Johnson and Tanay Ranadive, Technician Zach Timpano, and All Pro Cleaning on site. South wing except for Kindergarten wing has failed and will need to be re-cleaned. Crew suits up in full PPE and re-cleans South wing and takes out all carpets. Crew Stero-mists all of South wing.

1200 Crew breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch. Crew continues stereo-misting South wing and finishing up cleaning center wing.

1500 All Pro crew decons out and is off site. Samples are set in South wing, center wing, multi-purpose & main office.

1600 All samples are collected. QuES&T off site. Tanay Ranadive to Q Labs with samples.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Louis N Johnson & Tanay Ranadive  
Date: 8/28/18

Work Site: Pequenakonck E.S. Encl: North, Wing

Type of Removal: Mold Remediation Project #:Q18-1941

- 0900 Louis Johnson of QuES&T on site as Mold Assessor and All Pro on site. South wing samples have passed. Main lobby and main office have failed. These areas will be re-cleaned.
- 1100 Tanay Ranadive of QuES&T on site and is updated by Louis Johnson. Crew has cleaned failed areas and continues in North wing. Supervisor SteraMists center wing and hallway. Sheetrock removal and carpet removal continues in North wing.
- 1200 Crew breaks for 30 minute lunch.
- 1230 Crew returns from 30 minute lunch. Cafeteria is being re-cleaned.
- 1500 North wing and Cafeteria has been re-cleaned. Mold sampling is completed in these areas. Main Office and entryway is resampled.
- 1600 All Pro and Tanay Ranadive are off site. Louis Johnson is off site to Q Labs with samples.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Louis N Johnson

Date: 8/29/18

Work Site: Pequenakonck E.S.

Encl: East, West Wing

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Mold Assessor Louis Johnson of QuES&T and Technician William Allen of QuES&T on site with All Pro Cleaning. West wing, Cafeteria, and Multi-Purpose room samples passed. North wing classroom N-21 and Main Lobby samples failed.

1100 All Pro re-cleans N-21 and Main Lobby and then moves on to East wing.

1200 All Pro and QuES&T break for 30 minute lunch.

1230 All Pro and QuES&T return from 30 minute lunch. Crew suits up in full PPE to continue cleaning.

1500 QuES&T re-samples areas that failed as well as East wing.

All Pro decons out and is off site.

1600 QuES&T is off site.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Tanay Ranadive  
Date: 8/30/18

Work Site: Pequenakonck E.S.

Encl: Core, East, Gym

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 All Pro and Tanay Ranadive, Mold Assessor of QuES&T and William Allen, Technician of QuES&T arrive on site. Crew suits up in full PPE and begins cleaning all rooms in the core/library wing. East Wing samples have failed and is cordoned off with poly and signage. All Pro supervisor suits up in full PPE and begins to SteraMist East Wing. QuES&T continues initial mold assessments of the library.

1200 Crew breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch and suits up in full PPE to continue cleaning rooms. No sheet rock to be taken out as all walls are cementitious block. Library wing initial assessment is complete, several books will need to be thrown out. Tanay continues initial assessments of Gym, Gym Storage, Boys & Girls Locker Rooms/restrooms and the music rooms.

1500 East Wing has been SteraMisted, library wing classrooms are complete. QuES&T runs mold air samples for East Wing. All Pro decons out and is off site.

1600 Will Allen is off site to Q Labs with samples. Tanay is off site.



**QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES**

**DAILY PROJECT LOG**

Mold Assessor: Tanay Ranadive  
Date: 9/01/18

Work Site: Pequenakonck E.S.

Encl: Core, Gym

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Tanay Ranadive and Jonathan Mages of QuES&T on site. All Pro on site. East Wing and Cafeteria samples have passed. Crew suits up in full PPE and continues cleaning of center library. The tent and American Flag are taken down as it shows mold growth. Several books are thrown out as well. Initial assessments of Gym continue. All Pro splits crew and sends some to clean kitchen pantry and some to Gym Wing.

1200 Crew decons out and breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch and suit up in full PPE. They continue cleaning the Gym Wing and Library.

1500 All Pro decons out and is off site. QuES&T starts mold samples in Library, Gym Wing, and Kitchen Pantry.

1600 All samples are complete. QuES&T is off site and Tanay Ranadive is off site to Q Labs with samples.

QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES

DAILY PROJECT LOG

Mold Assessor: Tanay Ranadive

Date: 9/02/18

Work Site: Pequenaconck E.S.

Encl: Pantry, Gym

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Tanay Ranadive and Justin McFarland of QuES&T are on site.

All Pro is on site. All Gym Wing samples except for Men's & Womens restrooms have passed. Crew suits up in full PPE to re-clean Kitchen Pantry, Gym Storage Room, and both Men's & Women's restrooms. In the Kitchen Pantry, several important documents have mold present, but are set aside to be looked at by state officials.

1200 Crew decons out and breaks for 30 minute lunch.

1230 Crew returns from 30 minute lunch break and suits up in full PPE to SteraMist the Kitchen Pantry, Gym Storage, and both Men's & Women's Restrooms.

1500 All Pro decons out and is off site. Mechanical Room by the library, Kitchen Pantry, Gym Storage, and both Men's & Women's restrooms are all mold air sampled.

1600 Air Sampling is complete and Tanay Ranadive is off site to Q Labs with samples. Justin McFarland is off site.

**QUALITY ENVIRONMENTAL SOLUTIONS & TECHNOLOGIES**

**DAILY PROJECT LOG**

Mold Assessor: Tanay Ranadive  
Date: 9/04/18

Work Site: Pequenakonck E.S.

Encl: Pantry, Gym

Type of Removal: Mold Remediation

Project #:Q18-1941

0900 Tanay Ranadive of QuES&T is on site with All Pro. All mold air samples have passed except for the Mechanical Room near the Library. Crew suits up in full PPE and re-cleans the Mechanical Room. SteraMist is used in the room. All Pro and QuES&T equipment no longer in use is packed up.

1200 Tanay Ranadive re-samples the air in the Mechanical Room.

1230 Tanay Ranadive is off site to Q Labs with samples. All Pro remains on site.

# Appendix E: Certifications

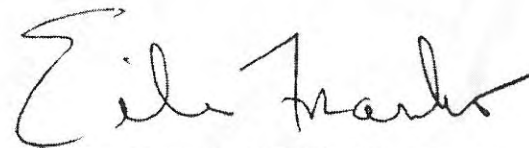
**NEW YORK STATE - DEPARTMENT OF LABOR**  
DIVISION OF SAFETY AND HEALTH  
LICENSE AND CERTIFICATE UNIT  
STATE CAMPUS BUILDING 12

**Mold Assessor Company License**

Quality Environmental Solutions + Technologies, In  
1376 Route 9  
WAPPINGERS FALLS, NY 12590

LICENSE NUMBER 00553  
DATE OF ISSUE: 3/22/2018  
EXPIRATION DATE 3/31/2020

This license is valid only for the contractor named above.



Eileen Franko, Director  
FOR THE COMMISSIONER OF LABOR



STATE OF NEW YORK - DEPARTMENT OF LABOR  
MOLD ASSESSOR



LOUIS JOHNSON

EXPIRES: 03-20

CERT# MA00532



01213 004630568 69

EYES BLU  
HAIR BLN  
HGT 5' 9 "

IF FOUND, RETURN TO:  
NYSOL - L&C UNIT  
ROOM 161A BUILDING 12  
STATE OFFICE CAMPUS  
ALBANY NY 12240





# QuES&T

Quality Environmental Solutions & Technologies, Inc  
1376 Route 9, Wappingers Falls, NY 12590  
Phone 845-298-6031 Fax 845-298-6251  
NYS DOL Training Provider # MTP-028

## Certificate of Completion

This certifies that on 2/22/2016

**Louis Johnson III**  
DMV License Number: 641924292


Successfully completed the 32 hour New York State Department of Labor Approved

Mold Assessor Initial Course

Pursuant to Article 32 of the New York State Labor Law

Attendee Identification Number: 1-16-02-22-028-005

Training Course Location: Wappingers Fall, NY



Kenneth C. Eck CIH, CSP, CFPS, CHMM  
Training Director

This certificate is not a license to perform assessment, remediation or abatement of mold projects



OSHA

001897147



U.S. Department of Labor  
Occupational Safety and Health Administration

Louis Johnson III

has successfully completed a 10-hour Occupational Safety and Health  
Training Course in

Construction Safety & Health

*Walter C. Veit*

06/10/08  
(Date)

(Trainer)

OSHA recommends Outreach Training courses as an orientation to occupational safety and health for workers. Participation is voluntary. Workers must receive additional training on specific hazards of their job. This course completion card does not expire.

For further information see our web site at [www.osha.gov/outreach.html](http://www.osha.gov/outreach.html)

# **QuES&T**

Quality Environmental Solutions & Technologies, Inc  
1376 Route 9, Wappingers Falls, NY 12590  
Phone 845-298-6031 Fax 845-298-6251

HEREBY CERTIFIES THAT

***LOUIS JOHNSON III***

HAS SUCCESSFULLY COMPLETED A TRAINING SEMINAR IN:

***MOLD REMEDIATION***

MEETING THE REQUIREMENTS OF 29 CFR 1926.62 29 CFR  
1910.134 AND HAS BEEN AWARDED THIS CERTIFICATE BY:



**DAVID C. VEIT**  
TRAINING INSTRUCTOR

ON THIS DATE: FEBRUARY 11, 2011

CERTIFICATE NUMBER: 11-MOLD-05-01

STATE OF NEW YORK - DEPARTMENT OF LABOR  
MOLD ASSESSOR



TANAY RANADIVE

EXPIRES: 03-20

CERT# MA00534



01213 004628809 88

EYES BRN  
HAIR BLK  
HGT 5' 10"

IF FOUND, RETURN TO:  
NYS DOL - L&C UNIT  
ROOM 161A BUILDING 12  
STATE OFFICE CAMPUS  
ALBANY NY 12240





12-005398747

This card acknowledges that the recipient has successfully completed a  
10-hour Occupational Safety and Health Training Course in  
**Construction Safety and Health**

**Tanay Ranadive**

David Veit

06/05/2015

Trainer name – print or type

(Course and date)

OSHA recommends Outreach Training Courses as an orientation to occupational safety and health for workers. Participation is voluntary. Workers must receive additional training on specific hazards of their job. This course completion card does not expire.

Use or distribution of this card for fraudulent purposes, including false claims of having received training, may result in prosecution under 18 U.S.C. 1001. Potential penalties include substantial criminal fines, imprisonment up to five years, or both.

For OSHA Outreach Training Program go to "Training" at [www.osha.gov](http://www.osha.gov)





# *QuES&T*

Quality Environmental Solutions & Technologies, Inc  
1376 Route 9, Wappingers Falls, NY 12590  
Phone 845-298-6031 Fax 845-298-6251  
NYS DOL Training Provider # MTP-028

## *Certificate of Completion*

This certifies that on 2/22/2016

**Tanay Ranadive**

DMV License Number: 859664473

Successfully completed the 32 hour New York State Department of Labor Approved

Mold Assessor Initial Course

Pursuant to Article 32 of the New York State Labor Law

Attendee Identification Number: 1-16-02-22-028-004

Training Course Location: Wappingers Fall, NY

Kenneth C. Eck CIH, CSP, CFPS, CHMM  
Training Director

This certificate is not a license to perform assessment, remediation or abatement of mold projects



STATE OF NEW YORK - DEPARTMENT OF LABOR  
MOLD ASSESSOR



FRANCIS MANNA

EXPIRES: 12-19

CERT# MA00052

12345678901234567890



01213 004430934 09

EYES BLK  
HAIR BRN  
HGT 5' 10"

IF FOUND, RETURN TO:  
NYSOL - L&C UNIT  
ROOM 161A BUILDING 12  
STATE OFFICE CAMPUS  
ALBANY NY 12240

STATE OF NEW YORK - DEPARTMENT OF LABOR  
MOLD ASSESSOR



MICHAEL SMITH

EXPIRES: 03-20

CERT# MA01291

1100001 00001 100 0000 10 0000 000



01213 004629882 45

EYES BLU  
HAIR BLN  
HGT 6' 0 "

IF FOUND, RETURN TO:  
NYSDOL - L&C UNIT  
ROOM 161A BUILDING 12  
STATE OFFICE CAMPUS  
ALBANY NY 12240